

HMGB1 polymorphisms in acute lymphoblastic leukemia

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Abstract

Acute lymphoblastic leukemia (ALL) is the most common pediatric malignancy and the leading cause of childhood death in contrast to the 90% cure rates. ALL includes different subtypes described by interrupt collections of somatic chromosomal alterations and sequence mutations that disrupt normal body functions such as lymphoid maturation, cell-cycle regulation, and tumor suppression. Having a significant role in several cancers, the high mobility group box-1 (HMGB1) gene considered an important gene in the development of tumors. Herein, the genetic role of HMGB1 was studied in the 49 Iranian patients with newly diagnosed ALL using Sanger sequencing of HMGB1 coding regions (exons 2 to 5). The results showed that none of the subjects in the study had any promising variants in the coding sequences of the HMGB1. These findings suggest that HMGB1 is not directly associated with ALL incidence and behavior. Further investigations using a large group of patients with different races and ethnicities are required to analyze the possible role of HMGB1 gene polymorphisms in ALL patients.

Introduction

Acute lymphoblastic leukemia (ALL) is the most common pediatric malignancy and the leading cause of childhood death in contrast to the 90% cure rates¹. ALL includes different subtypes described by distinct constellations of somatic chromosomal alterations and sequence mutations that disrupt lymphoid maturation, cell-cycle regulation, kinase signaling, tumor suppression, and chromatin modification². Of these genetic changes, some influence the risk of treatment failure and relapse in addition to leading to leukemogenesis. Notably, *KMT2A* (previously known as *MLL*) rearrangement, *BCR-ABL1* mutation, *ETV6-RUNX1* (due to t(12;21)) fusion, and activating kinase alterations in Ph-like ALL are correlated with poor disease outcome. Deletion or mutation of the *IKZF1* lymphoid transcription factor gene also confers a poor prognosis. Another common translocation is t(1;19), leading to *TCF3-PBX1* (*E2A-PBX1*) fusion, which occurs in approximately 5% of childhood cases as well as in adult ALL³⁻⁵.

Several single gene studies on ALL have been recently focused on single nucleotide polymorphism. However, the role of high mobility group box-1 (*HMGB1*) gene polymorphisms has not been investigated in ALL patients so far. HMGB1 protein is a highly conserved ubiquitous protein which is present in high concentration in the nucleus and cytoplasm of mammalian cells. This protein, previously known as a DNA binding protein, has a crucial role in the nucleosome structure maintenance and gene transcription regulation^{6,7}.

The aim of the current study was to detect the *HMGB1* gene polymorphisms among the Iranian patients with new-onset ALL, as well as its relation with the disease behavior, and patients' survival and outcome.

Materials and Methods

The study was carried out in the Children’s Medical Center in Tehran university of medical sciences. In this study, 49 newly diagnosed ALL patients consist of 21 females and 28 males with a median age of 28 years (range: 14 to 80 years) were included. The diagnosis of the patients was made according to the standard morphological examinations and immunophenotyping. Patient recruitment and all experimental protocols used in this study complied with the Declaration of Helsinki and were approved by the Ethics Committee of the Tehran University of Medical Sciences. Written informed consent was obtained from the patients prior to entering the study.

Peripheral whole blood samples were collected from each patient at the time of diagnosis and DNA was isolated by the standard salting-out method. Primers were designed for 4 coding exons (exons 2-5) of the *HMGB1* gene. DNA samples were amplified in a volume of 25 μ l, containing 40 ng of DNA template, 10 μ M of each reverse and forward primers, and 12 μ l of Taq DNA polymerase 2Xmastermix (Ampliçon). The cycling conditions were as follows: initial denaturing, 5 min at 95 °C, followed by 30 cycles of 30 s at 95 °C, 30 s at 60 °C, and 30 s at 72 °C. An additional extension step of 10 min at 72 °C was also performed. Afterward, PCR products were loaded and visualized on a 1% agarose gel using SYBR-safe dye and subsequently proceeded Sanger sequencing using Applied Biosystems 3130 Genetic Analyzer. Ultimately, the sequenced data of each individual were analyzed for searching for any possible variant in the exons.

Results

In the current study, all five exons of the *HMGB1* gene were analyzed in the subjects. Assuming the potential role in different malignancies according to previous studies, *HMGB1* gene polymorphisms were evaluated in newly diagnosed ALL patients. The results revealed that none of the 49 enrolled patients had any variants in the *HMGB1* gene.

Discussion

Acute lymphoblastic leukemia (ALL) is the most prevalent pediatric cancer and the leading cause of childhood death ¹. In this study, the *HMGB1* gene polymorphisms were used to study in 49 Iranian ALL patients with the purpose of finding the role of this gene in the development of ALL. After analyzing of all exons of the *HMGB1* in the subjects, no promising variant was identified in the coding sequences of the *HMGB1*. These findings suggest that *HMGB1* is not a clinically significant gene in ALL and other genetic changes may have a role in these patients.

The *HMGBs* are a highly conserved family including four members (*HMGB1* , *HMGB2* , *HMGB3* , and *HMGB4*). It is known that knocking out the *HMGB1* , *HMGB2* , and *HMGB3* genes in mice results in noticeable phenotypic changes, although the encoded proteins share approximately 80% amino acid sequence identity. Each *HMGB* has two DNA binding domains termed as HMG boxes A and B. *HMGB1-3* include an acidic C-terminal tail, whereas *HMGB4* lacks this tail ⁸. *HMGB1* , as a member of mammalian HMG-box family, include tandem homologous DNA binding domains, called HMG-box A and B. These domains comprised of about 80 amino acids folded into three α -helices that adopt a characteristic L-shaped structure and followed by a linker abundant in basic amino acid residues and a C-terminal acidic tail contains about 30 consecutive aspartate and glutamate residues⁸.

Besides performing nuclear functions, *HMGB1* has a significant extracellular function as damage-associated molecular pattern molecules (DAMPs) ⁹. Extracellular *HMGB1* functions as a DAMP to alert the innate immune system by recruiting mesangioblasts, stem cells and, smooth muscle cells ¹⁰.

Primary studies demonstrated the role of *HMGB1* as a late mediator of sepsis. Currently, it has been exhibited as a danger signal involved in the pathogenesis of various non-infectious inflammatory diseases^{11,12}. It has been also proved that *HMGB1* plays pivotal rules in tissue repair, remodeling, and preconditioning that these findings made *HMGB1* as an important protein in danger signal¹¹.

The *HMGB1* expression is higher in myeloid cells compared to lymphoid cells. *HMGB1* expression is upreg-

ulated in cancer cells, however, it is downregulated during aging, suggesting a critical role in development and cancer⁹. As a nuclear DNA-binding protein, HMGB1 is involved in the transcription of several cancer-related genes, including E-selectin¹³, TNF α ¹⁴, insulin receptor¹⁵, and BRCA¹⁶. In addition, in necrotic cancer cells, HMGB1 releases into tumor microenvironment that lead to chronic inflammatory and reparative responses that consequently leads to cancer cell survival and metastasis¹⁷. Having a role in metastasis, HMGB1 correlated with poor prognosis in a variety of cancers including breast¹⁸, colon¹⁹, pancreas²⁰, and prostate²¹. However, the role of HMGB1 has not been studied in ALL so far, plays an important role in tumor development, growth, and metastasis. Despite HMGB1 that no evidence is available for proving its role in ALL development, another member of the HMG family, HMGB3 has shown promising effects in improving ALL. It has been proved that HMGB3 in the fusion with NPU98 (HMGB3-NPU98 fusion protein) is an oncogene found in leukemia and augments malignant transformation in recipient mice²². It is known that HMGB1 has been secreted and accumulates in cell membranes during murine erythroleukemia cell differentiation²³. The N-terminal 18 amino acids of the hydrophilic acylated surface protein B, an unclassical secretory signal peptide, could deliver HMGB1 on the cell surface, efficiently²⁴. In erythroblast-macrophage contact, HMGB1 is involved in macrophage-mediated erythroid proliferation and maturation in a homophilic manner²⁵. It has been studied that during platelet activation, HMGB1 translocates to the membrane and is then released²⁶, which mediates neutrophil extracellular traps formation and function^{27,28}. Extracellular HMGB1 leads to chronic lymphocytic leukemia differentiation²⁹. It is exhibited that retreatment with HMGB1 results in endotoxin and lipoteichoic acid tolerance in bone marrow-derived macrophages and the acute monocytic leukemia cell line THP-1 through NF-KB activity downregulation^{9,30}. Interestingly, it is revealed that miR181a impedes the expression of HMGB1 in T- and B ALL cells and consequently results in a decline in cell proliferation and metabolic activity³¹. Moreover, HMGB1-mediated autophagy augments chemoresistance in cancer cells, including leukemia, colon cancer, gastric cancer, ovarian cancer, osteosarcoma, and pancreatic cancer⁹. Despite the comprehensive study of HMGB1 both biologically and clinically, there is not any promising evidence suggesting the role of HMGB1 in ALL. Kang, R. et al showed that the HMGB1 serum levels were significantly higher in ALL initial treatment group compared to the healthy control group and ALL complete remission group. Interestingly, HMGB1 levels had no significant differences between the healthy control group and ALL complete remission group. Moreover, it is shown that HMGB1 treatment of K562 cells, led to secretion and augmentation in the TNF- α level. Using JNK (SP600125), MEK (PD98059), and p38 MAPK (SB203580) inhibitors resulted in HMGB1-induced TNF- α secretion arrest. The serum HMGB1 measurement in the assessment of childhood ALL is beneficial due to HMGB1 stimulates leukemic cells to secrete TNF-alpha through a MAPK-dependent mechanism³².

In sum, HMGB1 has been studied in several biological and medical conditions majorly cancers such as colon, pancreatic, breast, and prostate cancers. A few studies investigated the role of the HMGB1 gene in leukemia however, lacking conclusive information around the hematological malignancies. Studying a large group of ALL patients and from different ethnicities may noticeably help understanding the HMGB1 role in ALL patients.

Conflict of Interest

The authors declare that they have no conflict of interests.

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Authors' contributions

“Elham Rayzan” contributed to the laboratory works and was in charge of the project management. “Saeed Farajzadeh Valilou” drafted the manuscript and contributed to the laboratory works. “Sara Hemmati” and “Amin Sadeghi” helped in improving the study design and proposal preparation. “Hamid Farajifard” designed the primers and analyzed the sanger results. “Sepideh Shahkarami” managed the patients’ recruitment and samplings, proposal drafting, and sanger results analysis. “Nima Rezaei” defined the study protocol and supervised the whole project. All authors read, critically revised, and approved the final manuscript.

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References

1. Hunger SP, Mullighan CG. Acute lymphoblastic leukemia in children. *N Engl J Med.* 2015;373(16):1541-1552.
2. Iacobucci I, Mullighan CG. Genetic basis of acute lymphoblastic leukemia. *J Clin Oncol.* 2017;35(9):975.
3. Mullighan CG, Su X, Zhang J, et al. Deletion of IKZF1 and prognosis in acute lymphoblastic leukemia. *N Engl J Med.*2009;360(5):470-480.
4. Churchman ML, Qian M, Te Kronnie G, et al. Germline genetic IKZF1 variation and predisposition to childhood acute lymphoblastic leukemia. *Cancer Cell.* 2018;33(5):937-948. e938.
5. Hein D, Borkhardt A, Fischer U. Insights into the prenatal origin of childhood acute lymphoblastic leukemia. *Cancer Metastasis Rev.*2020:1-11.
6. Yang H, Wang H, Czura CJ, Tracey KJ. HMGB1 as a cytokine and therapeutic target. *J Endotoxin Res.* 2002;8(6):469-472.
7. Chen G, Ward MF, Sama AE, Wang H. Extracellular HMGB1 as a proinflammatory cytokine. *J Interferon Cytokine Res.*2004;24(6):329-333.
8. Thomas JO, Travers AA. HMG1 and 2, and related 'architectural'DNA-binding proteins. *Trends Biochem Sci.*2001;26(3):167-174.
9. Kang R, Chen R, Zhang Q, et al. HMGB1 in health and disease. *Mol Aspects Med.* 2014;40:1-116.
10. Tang D, Billiar TR, Lotze MT. A Janus tale of two active high mobility group box 1 (HMGB1) redox states. *Mol Med.*2012;18(10):1360-1362.
11. Klune JR, Dhupar R, Cardinal J, Billiar TR, Tsung A. HMGB1: endogenous danger signaling. *Mol Med.* 2008;14(7-8):476-484.
12. Sims GP, Rowe DC, Rietdijk ST, Herbst R, Coyle AJ. HMGB1 and RAGE in inflammation and cancer. *Annu Rev Immunol.* 2009;28:367-388.
13. Aychek T, Miller K, Sagi-Assif O, et al. E-selectin regulates gene expression in metastatic colorectal carcinoma cells and enhances HMGB1 release. *Int J Cancer.* 2008;123(8):1741-1750.
14. Andersson U, Wang H, Palmblad K, et al. High mobility group 1 protein (HMG-1) stimulates proinflammatory cytokine synthesis in human monocytes. *The Journal of experimental medicine.*2000;192(4):565-570.
15. BRUNETTI A, Manfioletti G, CHIEFARI E, GOLDFINE ID, FOTI D. Transcriptional regulation of human insulin receptor gene by the high-mobility group protein HMGI (Y). *The FASEB Journal.*2001;15(2):492-500.
16. Baldassarre G, Battista S, Belletti B, et al. Negative regulation of BRCA1 gene expression by HMGA1 proteins accounts for the reduced BRCA1 protein levels in sporadic breast carcinoma. *Mol Cell Biol.*2003;23(7):2225-2238.
17. Dong XDE, Ito N, Lotze MT, et al. High mobility group box I (HMGB1) release from tumor cells after treatment: implications for development of targeted chemoimmunotherapy. *J Immunother.* 2007;30(6):596-606.

18. Dolde CE, Mukherjee M, Cho C, Resar LM. HMG-I/Y in human breast cancer cell lines. *Breast Cancer Res Treat.* 2002;71(3):181-191.
19. Fedele M, Bandiera A, Chiappetta G, et al. Human colorectal carcinomas express high levels of high mobility group HMGI (Y) proteins. *Cancer Res.* 1996;56(8):1896-1901.
20. Tarbé N, Evtimova V, Burtscher H, Jarsch M, Alves F, Weidle UH. Transcriptional profiling of cell lines derived from an orthotopic pancreatic tumor model reveals metastasis-associated genes. *Anticancer Res.* 2001;21(5):3221-3228.
21. Leman ES, Madigan MC, Brünagel G, Takaha N, Coffey DS, Getzenberg RH. Nuclear matrix localization of high mobility group protein I (Y) in a transgenic mouse model for prostate cancer. *J Cell Biochem.* 2003;88(3):599-608.
22. Petit A, Ragu C, Della-Valle V, et al. NUP98–HMGB3: a novel oncogenic fusion. *Leukemia.* 2010;24(3):654-658.
23. Passalacqua M, Zicca A, Sparatore B, Patrone M, Melloni E, Pontremoli S. Secretion and binding of HMG1 protein to the external surface of the membrane are required for murine erythroleukemia cell differentiation. *FEBS Lett.* 1997;400(3):275-279.
24. Zhu H, Wang L, Ruan Y, et al. An efficient delivery of DAMPs on the cell surface by the unconventional secretion pathway. *Biochem Biophys Res Commun.* 2011;404(3):790-795.
25. Hanspal M, Hanspal JS. The association of erythroblasts with macrophages promotes erythroid proliferation and maturation: a 30-kD heparin-binding protein is involved in this contact. 1994.
26. Maugeri N, Franchini S, Campana L, et al. Circulating platelets as a source of the damage-associated molecular pattern HMGB1 in patients with systemic sclerosis. *Autoimmunity.* 2012;45(8):584-587.
27. Mitroulis I, Kambas K, Chrysanthopoulou A, et al. Neutrophil extracellular trap formation is associated with IL-1 β and autophagy-related signaling in gout. *PLoS One.* 2011;6(12).
28. Tadie J-M, Bae H-B, Jiang S, et al. HMGB1 promotes neutrophil extracellular trap formation through interactions with Toll-like receptor 4. *American Journal of Physiology-Lung Cellular and Molecular Physiology.* 2013;304(5):L342-L349.
29. Jia L, Clear A, Liu F-T, et al. Extracellular HMGB1 promotes differentiation of nurse-like cells in chronic lymphocytic leukemia. *Blood, The Journal of the American Society of Hematology.* 2014;123(11):1709-1719.
30. Aneja RK, Tsung A, Sjodin H, et al. Preconditioning with high mobility group box 1 (HMGB1) induces lipopolysaccharide (LPS) tolerance. *J Leukoc Biol.* 2008;84(5):1326-1334.
31. Dahlhaus M, Schult C, Lange S, Freund M, Junghans C. MicroRNA 181a influences the expression of HMGB1 and CD4 in acute Leukemias. *Anticancer Res.* 2013;33(2):445-452.
32. Kang R, Tang D, Cao L, Yu Y, Zhang G, Xiao X. High mobility group box 1 is increased in children with acute lymphocytic leukemia and stimulates the release of tumor necrosis factor-alpha in leukemic cell. *Zhonghua er ke za zhi= Chinese Journal of Pediatrics.* 2007;45(5):329-333.