Impact of spatial variability in zooplankton grazing rates on carbon export flux

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Abstract

The biological carbon pump is a key controller of how much carbon is stored within the global ocean. This pathway is influenced by food web interactions between zooplankton and their prey. In global biogeochemical models, Holling Type functional responses are frequently used to represent grazing interactions. How these responses are parameterised greatly influences biomass and subsequent carbon export estimates. The half-saturation constant, or k value, is central to the Holling functional response. Empirical studies show k can vary over three orders of magnitude, however, this variation is poorly represented in global models. This study derives zooplankton grazing dynamics from remote sensing products of phytoplankton biomass, resulting in global distribution maps of the grazing parameter k. The impact of these spatially varying k values on model skill and carbon export flux estimates is then considered. This study finds large spatial variation in k values across the global ocean, with distinct distributions for micro- and mesozooplankton. High half-saturation constants, which drive slower grazing, are generally associated with areas of high productivity. Grazing rate parameterisation is found to be critical in reproducing satellite-derived distributions of nanophytoplankton biomass, highlighting the importance of top-down drivers for this size class. Spatially varying grazing dynamics decrease mean total carbon export by >17% compared to globally homogeneous dynamics, with increases in faecal pellet export and decreases in export from algal aggregates. This study highlights the importance of grazing dynamics to both community structure and carbon export, with implications for modelling marine carbon sequestration under future climate scenarios.
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Key Points:

• Inverse modelling predicts strong spatial variability in global grazing dynamics for two zooplankton functional types.
• Locally-tuned zooplankton grazing dynamics improve the model’s ability to reproduce satellite-derived phytoplankton biomass.
• Locally-tuned zooplankton grazing dynamics can decrease mean carbon flux by 17% and modify the routing of carbon export.

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Abstract

The biological carbon pump is a key controller of how much carbon is stored within the global ocean. This pathway is influenced by food web interactions between zooplankton and their prey. In global biogeochemical models, Holling Type functional responses are frequently used to represent grazing interactions. How these responses are parameterised greatly influences biomass and subsequent carbon export estimates. The half-saturation constant, or $k$ value, is central to the Holling functional response. Empirical studies show $k$ can vary over three orders of magnitude, however, this variation is poorly represented in global models. This study derives zooplankton grazing dynamics from remote sensing products of phytoplankton biomass, resulting in global distribution maps of the grazing parameter $k$. The impact of these spatially varying $k$ values on model skill and carbon export flux estimates is then considered. This study finds large spatial variation in $k$ values across the global ocean, with distinct distributions for micro- and mesozooplankton. High half-saturation constants, which drive slower grazing, are generally associated with areas of high productivity. Grazing rate parameterisation is found to be critical in reproducing satellite-derived distributions of nanophytoplankton biomass, highlighting the importance of top-down drivers for this size class. Spatially varying grazing dynamics decrease mean total carbon export by $>17\%$ compared to globally homogeneous dynamics, with increases in faecal pellet export and decreases in export from algal aggregates. This study highlights the importance of grazing dynamics to both community structure and carbon export, with implications for modelling marine carbon sequestration under future climate scenarios.

1 Introduction

The ocean plays a major role in mitigating the impact of climate change (Hoegh-Guldberg & Bruno, 2010). It is thought that over 20% of anthropogenic carbon dioxide emissions are stored within the global ocean (Friedlingstein et al., 2022). The biological carbon pump describes a suite of processes which can transport organic carbon from the surface ocean, to depths of over 1000m (Turner, 2015). This pathway is responsible for approximately 10% of the ocean’s carbon inventory (DeVries, 2022). As carbon dioxide emissions are predicted to increase over the 21st Century, it is essential to fully understand the processes underlying carbon sequestration via the biological pump and predict how they will change in the future (Siegel et al., 2022).

Organic carbon is exported out of the surface ocean as the faecal pellets of consumers or as aggregates of phytoplankton (Siegel et al., 2022). The rate that these forms of particulate carbon are exported via the biological carbon pump is directly influenced by zooplankton grazing. Grazing rates impact the biomass of both predator and prey (Rohr et al., 2023b) and consequently the production of algal aggregates and faecal pellets. Slower grazing rates, for example, reduce the amount of fecal pellets produced by consumers, which decreases the contribution of this pathway to carbon export.

Biogeochemical (BGC) models can estimate particulate carbon using a combination of grazing rates and mortalities (e.g. Aumont et al. (2015)). In BGC models, grazing dynamics between predator and prey can be described by a food limited functional response (Gentleman & Neuheimer, 2008; Anderson et al., 2010; Vallina et al., 2014). This dictates how ingestion rates change with prey density (Gentleman & Neuheimer, 2008). The choice and parameterisation of grazing functional responses can impact estimates of carbon export (Anderson et al., 2010). Holling Type II or Type III (Holling, 1959) grazing formulations are commonly used (Kearney et al., 2021; Rohr et al., 2022). These formulations require two parameters: the maximum grazing rate, $g$, and the half-saturation constant, $k$. The half-saturation constant represents the concentration of prey at which half the maximum grazing rate is reached (Gentleman & Neuheimer, 2008). Together these two parameters describe the shape and magnitude of the functional response. Ecologically, they represent the time taken to capture and consume prey (Rohr et al., 2022) – characteristics that vary with
species physiology (Hansen et al., 1997; Hirst & Bunker, 2003). Although the functional response is described by both parameters, population dynamics are most sensitive to change in the $k$ value (Rohr et al., 2022), which is the focus of this study.

Laboratory measurements of $k$ values (Hansen et al., 1997; Hirst & Bunker, 2003) show a large range in $k$ values, spanning 0.96-6000 mgC m$^{-3}$. Laboratory (Hansen et al., 1997), ecological (Barton et al., 2013) and modelling (Rohr et al., 2023b) studies also point towards strong spatial variability in $k$ values. However, even the most complex BGC models have fixed, globally homogenous, $k$ values. These models can simulate spatial variability in grazing dynamics through the competition of multiple plankton functional types, but this likely does not capture the full physiological variability. Some models use mechanisms such as multi-prey responses (e.g. Anderson et al. (2010, 2015)) and prey preferences (e.g. Aumont et al. (2015)) to further emulate this variability, but these mechanisms are limited and there is little observational data to confirm what emergent grazing dynamics should be (Rohr et al., 2023b). Furthermore, there is uncertainty around the impact of zooplankton grazing on carbon flux, which contributes to the large variability in global estimates of carbon export (Siegel et al., 2014; Boyd, 2015; Rohr et al., 2023b).

We address these gaps by using an inverse modelling approach to estimate spatial variation in zooplankton $k$ values. We find large variations with notable implications for carbon export. We first describe our approach, then our findings and then the implications of these for our overall understanding of zooplankton and carbon export.

2 Methodology

2.1 Overview

This study builds on the work of Siegel et al. (2014) which used satellite-derived estimates of Net Primary Productivity (NPP) and phytoplankton biomass to predict global grazing rates and subsequent estimates of carbon export. The work by Siegel et al. (2014) was extended by Archibald et al. (2019) to include diel vertical migrations (DVM) by zooplankton, allowing organic particulate to be exported via both passive sinking and the vertical movements of organisms. We modified the model by Archibald et al. (2019) to include explicit grazing and zooplankton biomass pools.

Here, we use a 0-D BGC box model to infer the optimal $k$ parameters for both microzooplankton and mesozooplankton, within each grid cell of a 1x1 degree global domain. We force this model with observed bottom-up controls (phytoplankton cell division rates) but allow it to prognostically compute Net Primary Productivity (NPP), phytoplankton biomass, zooplankton biomass and carbon export. Phytoplankton and zooplankton biomass pools are divided into two functional groups each (2P2Z). We then run a suite of simulations to determine what combination of $k$ values is required to best match satellite derived phytoplankton biomass and thus infer the spatial distribution of grazing dynamics. Finally, to understand how more realistic zooplankton behavioural diversity influences marine carbon cycling, we compare global prognostic ecosystem biomass and carbon export from three model scenarios: a run using non-optimised, globally homogenous $k$ values derived from literature (Baseline scenario); a run using optimised, globally homogenous $k$ values (Global-$k$ scenario) and a run using optimised, locally tuned $k$ values (Local-$k$ scenario). The model inputs (§2.2), the ecosystem sub-model (§2.3), the approach to determine optimised $k$ values (§2.4), the carbon export sub-model (§2.5) and results analysis (§2.6) are discussed below.

2.2 Input Data

The model is forced by satellite-derived phytoplankton community mean growth rates, $\mu$. The use of $\mu$ ensures coupling between NPP and grazing dynamics. Without this coupling, the top-down influence of grazing dynamics would be removed. This would make
overgrazing of phytoplankton an impossibility, as there would always be NPP regardless of what the free-running biomass population is. \( \mu \) was selected as an input over the explicit representation of nutrients to ensure observational forcing remained.

In the Carbon-based Productivity Model (CbPMv2) (Westberry et al., 2008), \( \mu \) is computed from satellite derived chlorophyll-to-carbon ratios (Behrenfeld et al., 2005). Net Primary Productivity (NPP) can then be derived using the relationship between \( \mu \) and satellite-derived estimates of phytoplankton carbon biomass \( P_{\text{obs}} \), where

\[
NPP = \mu P_{\text{obs}} \quad (1)
\]

In CbPMv2, \( \mu \) is computed for the bulk phytoplankton population; however, in this study we needed to differentiate the growth rates of two phytoplankton classes, to force our 2P2Z model. The distribution of particle backscatter can partition phytoplankton carbon biomass across size classes, but not their respective growth rates. To estimate the partitioning of growth rates into two size classes \( \mu_i \) we assume a fixed allometric ratio, then determine the values required to produce bulk NPP from the two biomass pools. The following three equations are satisfied at each time step and location:

\[
P_{\text{obs}} = PS_{\text{obs}} + PL_{\text{obs}} \quad (2)
\]

\[
NPP = \mu PS_{\text{obs}} + \mu PL_{\text{obs}} \quad (3)
\]

\[
\frac{\mu PL}{\mu PS} = \left( \frac{M_{PL}}{M_{PS}} \right)^{-0.25} \quad (4)
\]

Both \( NPP \) and \( P_{\text{obs}} \) are derived from monthly climatologies presented in detail in Siegel et al. (2014). These climatologies are then interpolated to produce daily data. \( NPP \) values come from the Carbon-based Productivity Model (CbPMv2) (Westberry et al., 2008), which uses observations made by the Sea-viewing Wide-Field-of-view (SeaWiFS) satellite ocean colour mission between 1997 and 2008 (McClain, 2009; Siegel et al., 2014, 2013). Phytoplankton biomass values \( P_{\text{obs}} \) are estimated using particulate backscattering coefficient data (Behrenfeld et al., 2005; Westberry et al., 2008; Kostadinov et al., 2010; Siegel et al., 2013). \( P_{\text{obs}} \) is partitioned into two size classes \( (PS \& PL) \) using the slope of the particle size spectrum (Kostadinov et al., 2010; Siegel et al., 2014). \( M_i \) represents body size for the two size classes which has an allometric scaling constant of \(-0.25\) applied in accordance with metabolic theory (e.g. West et al. (1997)). This ensures the growth rate of nanoplatoplanckton is always faster than microphytoplankton. For \( M_i \), the same lower size limit implemented to partition \( P_{\text{obs}} \) is used, i.e. 20 \( \mu m \) and 0.5 \( \mu m \) for \( PL \) and \( PS \) respectively (Kostadinov et al., 2010; Siegel et al., 2014). A maximum value for \( \mu_i \) is set at 2 \( d^{-1} \) to correspond with the CbPMv2 data (Westberry et al., 2008). Observed minimum growth rates are approximately 0.1 \( d^{-1} \), however the CbPMv2 model extrapolates this towards 0 (Westberry et al., 2008). In our study we use a minimum growth rate within the range of these two values (0.01 \( d^{-1} \)).

In the CbPMv2, all properties are assumed to be constant and distributed evenly within the mixed layer (Westberry et al., 2008). Within this study, phytoplankton biomass is assumed to be homogeneous across the mixed layer and negligible below the mixed layer depth as in Siegel et al. (2014). Integrated NPP is assumed constant across the euphotic depth as per Siegel et al. (2014). To enable the calculation of \( \mu \), depth integrated NPP is divided by the greater of euphotic zone depth \( (Z_{eu}) \) or mixed layer depth \( (Z_{ml}) \). Depth data is interpolated from monthly climatologies also presented in detail in Siegel et al. (2014).
2.3 Ecosystem Sub-Model

A simple Phytoplankton-Zooplankton (2P2Z) model is constructed (Table 1). To run the ecosystem model, the global ocean is divided into a 1 degree latitude/longitude grid. The model is only run in grid cells with remote sensing products for a minimum of 10 out of 12 months. This limits the model to roughly between 50 and −50°N, covering approximately \(2.93 \times 10^8\) km\(^2\) of the global ocean or just over 80% of its total surface area. This avoids estimation bias in polar regions due to seasonal ice and cloud cover. The model is run with a daily time step and spun up until quasi-equilibrium is reached. Results are taken from the last year of the model run.

The rate of change, per day, in biomass within the mixed layer (\(Z_{ml}\)) for each size class is given by

\[
\frac{dPS}{dt} = \mu_{PS}PS - G_{ZS} - \text{agg}_{PS}PS^2 - m_P(PS - PS_0) - \frac{PS}{Z_{ml}} \frac{dz_{ml}}{dt} H\left(\frac{dz_{ml}}{dt}\right)
\]

\[
\frac{dPL}{dt} = \mu_{PL}PL - G_{ZL,PL} - \text{agg}_{PL}PL^2 - m_P(PL - PL_0) - \frac{PL}{Z_{ml}} \frac{dz_{ml}}{dt} H\left(\frac{dz_{ml}}{dt}\right)
\]

\[
\frac{dZS}{dt} = b_{ZS}G_{ZS} - m_{ZS}(ZS - ZS_0) - G_{ZL,ZS}
\]

\[
\frac{dZL}{dt} = b_{ZL}(G_{ZL,PL} + G_{ZL,ZS}) - m_{ZL}(ZL - ZL_0) - p_{ZL}ZL^2
\]

where \(PS, PL, ZS,\) and \(ZL\) represent biomass of nanophytoplankton (2-20\(\mu\)m), microphytoplankton (20-200\(\mu\)m), microzooplankton (20-200\(\mu\)m) and mesozooplankton (>200\(\mu\)m) respectively (Moriarty & O’Brien, 2013; Calbet & Calbet, 2008; Sieburth et al., 1978). The model does not resolve vertical or horizontal movement, therefore, biomass represents the mean concentration within the mixed layer, with the assumption of even distribution. Natural mortality \(m_i\) terms have a lower threshold applied of 0.2 for phytoplankton (Aumont et al., 2015) and 1.0 for zooplankton (Archibald et al., 2019) for model stability. Algal aggregates are represented as quadratic mortality terms \(\text{agg}_i\) of plankton biomass (Aumont et al., 2015). This enables changes in biomass to be reflected in algal export. The influence of shear on aggregate formation (Aumont et al., 2015) is not represented due to the lack of vertical movement and other physical dynamics within the model. The last term in Equations 5 and 6 describes the dilution of biomass as the depth of the mixed layer increases (Archibald et al., 2019; Siegel et al., 2014). \(H=1\) if the change in mixed layer depth is less than or equal to zero, or \(H=0\) otherwise (Archibald et al., 2019; Siegel et al., 2014; Evans & Parslow, 1985). The sub-model is closed by a quadratic mortality term \(p_{ZL}\) for mesozooplankton, which represents grazing by higher trophic levels.

Zooplankton growth is the product of gross growth efficiency \(b_i\) and grazing (Anderson et al., 2015). Grazing rates \(G_i\) are based on Holling Type III (Holling, 1959) functional responses, where

\[
G_{ZS} = \frac{g_{ZS}PS^2}{k_0^2 + PS^2}ZS
\]

\[
G_{ZL,PL} = \frac{g_{ZL}PL^2}{k_1^2 + PL^2}ZL
\]
Table 1. Model variables, parameters and forcing fields.

<table>
<thead>
<tr>
<th>Parameters</th>
<th>Symbol</th>
<th>Description</th>
<th>Value</th>
<th>Unit</th>
<th>Refs.</th>
</tr>
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<tr>
<td>dt</td>
<td>Model time step</td>
<td>1</td>
<td>day</td>
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<td>-</td>
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<tr>
<td>$k_0$</td>
<td>Microzooplankton half-saturation constant</td>
<td>-</td>
<td>mgC m$^{-3}$</td>
<td>-</td>
<td>-</td>
</tr>
<tr>
<td>$k_1$</td>
<td>Mesozooplankton half-saturation constant</td>
<td>-</td>
<td>mgC m$^{-3}$</td>
<td>-</td>
<td>-</td>
</tr>
<tr>
<td>$b_{ZS}$</td>
<td>Microzooplankton gross growth efficiency</td>
<td>0.3</td>
<td>-</td>
<td>(1)</td>
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<tr>
<td>$b_{ZL}$</td>
<td>Mesozooplankton gross growth efficiency</td>
<td>0.5</td>
<td>-</td>
<td>(2)(3)</td>
<td></td>
</tr>
<tr>
<td>$g_{ZS}$</td>
<td>Maximum grazing rate of microzooplankton</td>
<td>2</td>
<td>d$^{-1}$</td>
<td>(4)</td>
<td></td>
</tr>
<tr>
<td>$g_{ZL}$</td>
<td>Maximum grazing rate of mesozooplankton</td>
<td>2</td>
<td>d$^{-1}$</td>
<td>(4)</td>
<td></td>
</tr>
<tr>
<td>$m_{P}$</td>
<td>Phytoplankton mortality</td>
<td>0.1</td>
<td>d$^{-1}$</td>
<td>(2)(5)</td>
<td></td>
</tr>
<tr>
<td>$m_{ZS}$</td>
<td>Natural mortality microzooplankton</td>
<td>0.05</td>
<td>d$^{-1}$</td>
<td>(1)(6)</td>
<td></td>
</tr>
<tr>
<td>$m_{ZL}$</td>
<td>Natural mortality mesozooplankton</td>
<td>0.005</td>
<td>d$^{-1}$</td>
<td>(1)</td>
<td></td>
</tr>
<tr>
<td>$PS_{0}/PL_{0}$</td>
<td>Phytoplankton mortality refuge</td>
<td>0.2</td>
<td>mgC m$^{-3}$</td>
<td>(1)</td>
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</tr>
<tr>
<td>$Z_{ml}$</td>
<td>Mixed layer depth</td>
<td>m</td>
<td>(2)(5)</td>
<td></td>
<td></td>
</tr>
<tr>
<td>$Z_{eu}$</td>
<td>Depth of euphotic layer</td>
<td>m</td>
<td>(2)(5)</td>
<td></td>
<td></td>
</tr>
<tr>
<td>$\mu$</td>
<td>Phytoplankton Growth Rate</td>
<td>d$^{-1}$</td>
<td>(12-14)</td>
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<table>
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<tr>
<th>Forcing Fields</th>
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<th>Description</th>
<th>Unit</th>
<th>Refs.</th>
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<td>$Z_{ml}$</td>
<td>Mixed layer depth</td>
<td>m</td>
<td>(2)(5)</td>
</tr>
<tr>
<td></td>
<td>$Z_{eu}$</td>
<td>Depth of euphotic layer</td>
<td>m</td>
<td>(2)(5)</td>
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<td></td>
<td>$\mu$</td>
<td>Phytoplankton Growth Rate</td>
<td>d$^{-1}$</td>
<td>(12-14)</td>
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<th>Prognostic Variables</th>
<th>Symbol</th>
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<th>Unit</th>
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<tr>
<td>NPP</td>
<td>Net Primary Productivity</td>
<td>mgC m$^{-2}$d$^{-1}$</td>
<td></td>
</tr>
<tr>
<td>PS</td>
<td>Nanophytoplankton biomass</td>
<td>mgC m$^{-3}$</td>
<td></td>
</tr>
<tr>
<td>PL</td>
<td>Microphytoplankton biomass</td>
<td>mgC m$^{-3}$</td>
<td></td>
</tr>
<tr>
<td>ZS</td>
<td>Microzooplankton biomass</td>
<td>mgC m$^{-3}$</td>
<td></td>
</tr>
<tr>
<td>ZL</td>
<td>Mesozooplankton biomass</td>
<td>mgC m$^{-3}$</td>
<td></td>
</tr>
<tr>
<td>G$_{ZS}$</td>
<td>Microzooplankton grazing rate on Nanophytoplankton</td>
<td>mgC m$^{-3}$d$^{-1}$</td>
<td></td>
</tr>
<tr>
<td>G$_{ZL,PL}$</td>
<td>Mesozooplankton grazing rate on Microphytoplankton</td>
<td>mgC m$^{-3}$d$^{-1}$</td>
<td></td>
</tr>
<tr>
<td>G$_{ZL,ZS}$</td>
<td>Mesozooplankton grazing rate on Microzooplankton</td>
<td>mgC m$^{-3}$d$^{-1}$</td>
<td></td>
</tr>
<tr>
<td>G$_{ZL}$</td>
<td>Mesozooplankton combined grazing rate</td>
<td>mgC m$^{-3}$d$^{-1}$</td>
<td></td>
</tr>
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<td>$F_{eu}$</td>
<td>Total POC flux out of the euphotic zone</td>
<td>mgC m$^{-2}$d$^{-1}$</td>
<td></td>
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<tr>
<td>$F_{alg}$</td>
<td>Flux of algal aggregates out of the euphotic zone</td>
<td>mgC m$^{-2}$d$^{-1}$</td>
<td></td>
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<tr>
<td>$F_{fec}$</td>
<td>Flux of fecal pellets out of the euphotic zone</td>
<td>mgC m$^{-2}$d$^{-1}$</td>
<td></td>
</tr>
<tr>
<td>$J_{dvm}$</td>
<td>DVM-mediated export flux</td>
<td>mgC m$^{-2}$d$^{-1}$</td>
<td></td>
</tr>
<tr>
<td>$J_{met}$</td>
<td>Respired DIC produced in twilight zone by migrating ZL</td>
<td>mgC m$^{-2}$d$^{-1}$</td>
<td></td>
</tr>
<tr>
<td>$J_{f_{fc}}$</td>
<td>Faecal pellets produced in twilight zone by migrating ZL</td>
<td>mgC m$^{-2}$d$^{-1}$</td>
<td></td>
</tr>
<tr>
<td>ER</td>
<td>Export Ratio</td>
<td>-</td>
<td></td>
</tr>
<tr>
<td>DER</td>
<td>DVM Export Ratio</td>
<td>-</td>
<td></td>
</tr>
<tr>
<td>DRR</td>
<td>Respiration Ratio</td>
<td>-</td>
<td></td>
</tr>
<tr>
<td>RD</td>
<td>Weighted depth of respiration</td>
<td>-</td>
<td></td>
</tr>
<tr>
<td>$f_{fc}$</td>
<td>Fraction of fecal pellets in the euphotic zone</td>
<td>-</td>
<td></td>
</tr>
<tr>
<td>$p_{met}$</td>
<td>Fraction of metabolism in the twilight zone</td>
<td>-</td>
<td></td>
</tr>
</tbody>
</table>

\[ G_{ZL,ZS} = \frac{g_{ZL} ZS^2}{k_1^2 + ZS^2} ZL \]  

(11)

\( k_0 \) and \( k_1 \) are half-saturation constants and \( g_i \) are maximum grazing rates. There is no prey preference (Aumont et al., 2015) for mesozooplankton grazing and no multiple prey feeding response (Anderson et al., 2010, 2015) as this fundamentally changes the relationship between \( k \) and the prey distribution (Gentleman et al., 2003; Rohr et al., 2022; Anderson et al., 2015). The grazing terms allow for two-way coupling between zooplankton and their prey, so that grazing rates are influenced by both predator and prey biomass. A Type III functional response is chosen due to increased stability, its suitability to coarse resolution global model and improved reproduction of seasonal population dynamics compared to Type II (Rohr et al., 2023b, 2022).

### 2.4 Optimisation of Grazing Dynamics

This study aims to assess the impact of locally-tuned grazing dynamics on outputs from a coupled ecosystem-carbon export model. To do this three scenarios are considered (§2.1). The same ecosystem-carbon export model and bottom-up forcing is used for all three scenarios and all parameters except \( k \) are kept constant. For the Baseline and Global-\( k \) scenarios, the same pair of \( k \) values is used for every grid cell location (i.e. they are globally homogeneous). For the Baseline scenario, the median \( k \) values from 40 models reviewed by Rohr et al. (2022) are used. These are 40 and 80 mgC m\(^{-3}\) for microzooplankton and mesozooplankton, respectively. For the Global-\( k \) scenario, a single pair of globally optimised values are used. For the Local-\( k \) scenario, \( k \) values are locally tuned, at every grid cell location. The optimisation process for both the globally optimised pair, and locally-tuned values is detailed below.

For each model grid point, the optimum half saturation constant for both microzooplankton (\( k_0 \)) and mesozooplankton (\( k_1 \)) grazing is assessed using an inverse modelling approach. Multiple simulations of the model are run, each with a different set of \( k_0 \) and \( k_1 \) values. The output of the model is then compared to the climatological seasonal cycle of phytoplankton biomass (Behrenfeld et al., 2005; Westberry et al., 2008; Siegel et al., 2013, 2014). The \( k \) values that most closely reproduce these satellite-derived biomass values are then selected as the ‘optimum’ \( k \) values. To find the optimum values, a cost function is used. Several different cost functions were analysed which produced consistent results (Figures S1 & S2). The cost function presented here (Equation 12) is the sum of the normalised absolute average error (nAAE) (Stow et al., 2009) for both nanophytoplankton and microphytoplankton, computed across the full seasonal cycle. This represents the degree of agreement in the size and alignment of the seasonal cycle between model and observations. Here, the term ‘observations’ refers to the satellite-derived phytoplankton biomass. A value of zero indicates a perfect match and alignment with observations.

\[
\text{Cost} = n \text{AAE}_{PS} + n \text{AAE}_{PL}
\]  

(12)

where,

\[
n \text{AAE}_{P(i)} = \frac{\text{AAE}_{P(i)}}{\sigma_o}
\]  

(13)

\[
\text{AAE}_{P(i)} = \sum \frac{|P(i)_{\text{obs}} - P(i)_{\text{mod}}|}{n}
\]  

(14)

\( \sigma_o \) is the standard deviation of the observed data, which represents observed temporal variance in the seasonal cycle; \( n \) is the total number of observations across the climatological
year; i is the size class and \( P(i)_{\text{obs}} \) and \( P(i)_{\text{mod}} \) are observation and model values of phytoplankton respectively. The absolute average error is normalised by the standard deviation of the observed climatology, to enable comparisons of relative errors in high and low productivity regions. Due to the uncertainty in zooplankton observational estimates (Strömberg et al., 2009) a zooplankton term was not included in the cost function.

To maximise computational efficiency, two routines of \( k \) optimisation are carried out. The first coarse resolution routine uses 15 log spaced values of \( k_0 \) and \( k_1 \) (mgC m\(^{-3}\)): 16, 20, 26, 33, 43, 54, 70, 89, 114, 146, 187, 239, 306, 392 and 501. These are within the range of empirical and model estimates presented in Rohr et al. (2022). The model is run for each possible pair of half-saturation constants, at every grid cell (i.e. a total of 225 (15x15) runs at each grid cell). The pair of half-saturation constants that produces the lowest cost are then selected as the optimal values at that location. The result is a distribution of optimal \( k \) values across the global ocean.

To improve the resolution of our optimisation, a second optimisation routine is then carried out. At each location, the sampling input of \( k \) values is calculated by first taking the optimum \( k \) values from the coarse optimisation run. Next, an upper and lower limit for input values is calculated by \( \pm10\% \) in each direction from the optimised values. Numbers are rounded to the nearest integer. The model is then run for every pairing of integers between these two limits at that location. The cost function is then reevaluated at each grid cell location and a new optimised pairing of \( k \) values selected. For example, if the coarse optimisation identifies \( k_0 = 20 \) & \( k_1 = 26 \) as optimal, we then rerun the simulation for all integer values (and pairings) of \( 18 <= k_0 <= 22 \) and \( 23 <= k_1 <= 29 \).

For the Global-\( k \) scenario, the globally homogenous pair of \( k \) values are estimated by globally integrating the cost function for each \( k \) pairing. The pair that produce the lowest cost value from this global integration are selected for the Global-\( k \) scenario. The results from the coarse resolution optimisation routine are used for this integration. For the Local-\( k \) scenario, the optimum pair of \( k \) values estimated for every grid cell (from the second optimisation routine) are used.

### 2.5 Carbon Export Sub-Model

The Archibald et al. (2019) carbon export model is used to examine the impact of grazing parametrisation in this study. The carbon export model consists of two modules: the euphotic and twilight zone modules. In this study, the twilight zone component has remained unchanged from its description in Archibald et al. (2019). However, a few changes have been made to the euphotic zone module to reflect the use of fully-coupled grazing terms and the use of \( \mu \) as an input instead of NPP. These changes are described below.

Carbon export out of the euphotic zone (Total Export Flux) is the sum of the passive sinking flux (\( F_{\text{eu}} \)) and DVM-mediated flux (\( J_{\text{dvm}} \)).

\[
\text{Total Export Flux} = F_{\text{eu}} + J_{\text{dvm}} \tag{15}
\]

\[
F_{\text{eu}} = F_{\text{alg}} + F_{\text{fec}} \tag{16}
\]

\( F_{\text{eu}} \) is the sum of microphytoplankton algal aggregates (\( F_{\text{alg}} \)) and faecal pellets produced by mesozooplankton grazing (\( F_{\text{fec}} \)). Sinking algal aggregates from the euphotic layer (\( F_{\text{alg}} \)) are estimated as

\[
F_{\text{alg}} = Z_{\text{eu}} (1 - \text{bact}) \ (agg_{ZL} \ PL^2 + 0.5 \ m_{PL} \ PL) \tag{17}
\]
where the microphytoplankton aggregation term and 50 % of microphytoplankton linear
mortality contributes to sinking aggregates (Aumont et al., 2015). As in Aumont et al.
(2015), the remaining 50 % of the linear mortality term is classed as small POC, which
is retained in the euphotic zone. $b_{act}$ is the proportion lost to bacterial remineralisation
(Aumont et al., 2015). Nanophytoplankton does not contribute to algal export in the model,
as its smaller cell size means aggregates are assumed to contribute to the microbial web in
the euphotic zone, rather than sinking export flux (Archibald et al., 2019; Calbet & Landry,
2004).

Euphotic zone sinking faecal pellets ($F_{fec}$) and faecal pellets produced in the twilight
($J_{fec}$) are estimated as

$$F_{fec} = (p_{dvm} f_{fec} + (1 - p_{dvm})) (m_{fec} G_{ZL}) Z_{eu}$$

(18)

$$J_{fec} = p_{dvm} (1 - f_{fec}) (m_{fec} G_{ZL}) Z_{eu}$$

(19)

where $G_{ZL}$ is the combined grazing rate for mesozooplankton on both prey types ($G_{ZL,PL} +
G_{ZL,PS}$), $p_{dvm}$ is the proportion of mesozooplankton that participate in DVM, $m_{fec}$ is the
fraction of grazed carbon expelled as faecal pellets and $f_{fec}$ is the proportion of faecal
pellets expelled in the euphotic zone. DVM is treated as a single event and particulate
organic carbon (POC) is a single pool of carbon that decays exponentially (Archibald et
al., 2019). All carbon export parameter values (Table 1) were kept consistent with those
detailed in Archibald et al. (2019). In this study, microzooplankton do not vertically migrate
and their faecal pellets do not contribute to export flux as their smaller pellet size means
they are assumed to be consumed by the microbial loop in the euphotic zone (Archibald et
al., 2019; Calbet & Landry, 2004).

Finally, the production of respired dissolved inorganic carbon (DIC) in the twilight zone
($J_{met}$) is estimated as

$$J_{met} = p_{met} p_{dvm} f_{met} (G_{ZL} - m_{fec} G_{ZL}) Z_{eu}$$

(20)

where $p_{met}$ is the fraction of total metabolism that occurs in the twilight zone and $f_{met}$
is the fraction of absorbed carbon that is metabolised. The contribution of both $J_{met}$ and
$J_{fec}$ to DVM-mediated flux ($J_{dvm}$) is then described by

$$J_{dvm} = J_{met} + J_{fec}$$

(21)

2.6 Analysis

To analyse the output data from the optimisation of grazing dynamics and the ecosystem-
carbon export model, several additional metrics are considered.

2.6.1 Grazing Dynamics

The biomass-weighted $k$ value (BW-$k$) combines both $k_0$ and $k_1$ values. It considers
the optimal $k$ value for each zooplankton class and their relative abundance at every grid
cell location (BW-$k = (k_0 ZS + k_1 ZL)/Z$). This reflects the emergent grazing dynamics of
the entire zooplankton community. The half-saturation constant and maximum grazing rate
can be related to the prey capture efficiency, $\varepsilon$. The prey capture efficiency is calculated by
dividing the maximum grazing rate (See Table 2) by the half-saturation constant for each
grid cell (Rohr et al., 2022) ($\varepsilon = g / k^2$). To understand the relationship between $k$ and
both NPP and prey biomass, a linear regression is fitted to log-normalised data.
2.6.2 Carbon Export

The export ratio represents the proportion of NPP exported as carbon from the euphotic zone. DVM export ratio (DER) is DVM-mediated export as a fraction of total carbon exported from the euphotic zone. The DVM respiration ratio (DRR) is the amount of respiration carried out by migrating zooplankton as a fraction of the integrated respiration from the twilight zone (Zeu – 1000m). The weighted depth of respiration (RD) is the increase in depth of dissolved inorganic carbon (DIC) production and oxygen utilisation, as a result of zooplankton vertical migrations. Carbon export metrics are described in further detail in Archibald et al. (2019).

3 Results

3.1 Distribution of Locally Tuned Grazing Dynamics

Local tuning of k values results in high variability in inferred grazing dynamics (Figure 1 & S1-4). k values span a range of 537 mgC m\(^{-3}\) (Table 2). High k values are generally associated with highly productive regions (Figure 1c). Lower values are generally associated with the less productive subtropical oligotrophic gyres, with the exception of the eastern South Pacific, where maximum k values (551 mgC m\(^{-3}\)) are estimated for microzooplankton. Maximum k values are also found in the high latitudes of the southern hemisphere.

Zooplankton functional groups are characterised by different grazing dynamics. Microzooplankton k values estimated from local optimisation are, on average, lower than mesozooplankton (median k values are 18 mgC m\(^{-3}\) and 27 mgC m\(^{-3}\) respectively), suggesting faster grazing for the smaller size class (Table 2 & S1). In addition, the globally optimised pair of k values (estimated for the Global-k scenario - see §2.4) are 33 mgC m\(^{-3}\) and 392 mgC m\(^{-3}\) for micro- and mesozooplankton respectively. Microzooplankton are generally characterised by more efficient grazing than mesozooplankton, with the exception of the oligotrophic gyres (Figure S5).

The distribution of grazing dynamics also differs between zooplankton size classes. This is particularly evident in equatorial upwelling regions, where microzooplankton and mesozooplankton communities are characterised by low and high k values respectively (Figure 1a, b). These differences result in divergent relationships between k values, NPP and prey biomass for the two size classes. High k\(_0\) values are associated with low NPP and prey biomass, whilst high k\(_1\) values are associated with high NPP and prey biomass (Figure S6).

Table 2. Locally-tuned microzooplankton (k\(_0\)) and mesozooplankton (k\(_1\)) half-saturation constants estimated using the cost function. Half-saturation constant values are in mgC m\(^{-3}\). Global-k and Baseline scenario k values are included below for comparison (NB: average statistics cannot be provided for these two scenarios due to the same value being used for every grid cell in the model domain).

<table>
<thead>
<tr>
<th></th>
<th>Local-k</th>
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<tbody>
<tr>
<td></td>
<td>k(_0)</td>
<td>k(_1)</td>
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</tr>
<tr>
<td>Median</td>
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<tr>
<td>Geometric Mean</td>
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<td>38</td>
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</tr>
<tr>
<td>Biomass-weighted Mean</td>
<td>31</td>
<td>49</td>
<td></td>
</tr>
<tr>
<td>Range</td>
<td>14-551</td>
<td>14-551</td>
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<tr>
<td>IQR</td>
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<td>38(14-52)</td>
<td></td>
</tr>
<tr>
<td>Global-k</td>
<td>33</td>
<td>392</td>
<td></td>
</tr>
<tr>
<td>Baseline</td>
<td>40</td>
<td>80</td>
<td></td>
</tr>
</tbody>
</table>
Figure 1. Locally-tuned $k$ values. (a) Microzooplankton half-saturation constants ($k_0$) estimated using the cost function (Equation 12). b) Mesozooplankton half-saturation constants ($k_1$) estimated using the cost function. $k$ values are in mgC m$^{-3}$. c) Biomass-weighted $k$ values (BW-$k$) which considers the optimal $k$ value for each zooplankton size class and their relative abundance. BW-$k$ reflects the overall grazing dynamics of the entire zooplankton community. The maximum and minimum values on the colourbar represent the maximum/minimum $k$ values sampled in the optimisation.

3.2 Impact of Locally Tuned Grazing Dynamics on Model Skill

Locally-tuned $k$ values improve model skill in comparison to globally homogenous $k$ values (Figure 2). Mean cost (Equation 12) is reduced by 43% in the Local-$k$ scenario, compared to the Global-$k$ scenario. Therefore, the use of locally tuned $k$ values improves
Figure 2. Cost values representing the difference between observations of phytoplankton biomass and the model phytoplankton biomass (Equation 12). a) Cost values from the Baseline model scenario (non-optimised $k$ values). b) Percentage change in cost values in the Global-$k$ scenario compared to the Baseline scenario. c) Percentage change in cost values in the Local-$k$ scenario compared to the Baseline.

The model’s ability to reproduce satellite-derived phytoplankton biomass (Figure 3). In comparison, the use of optimised globally homogenous $k$ values (Global-$k$) has a limited ability (-14%) in reducing model cost from the Baseline scenario. Reduction in cost values due to local tuning is most evident in the tropics and subtropics, particularly productive upwelling regions (Figure 2). Despite improvements, microphytoplankton biomass estimates are the greatest source of error (78%) in the Local-$k$ cost function (Figure S7-8; Table S3).
3.3 Ecosystem Impact

The reproduction of the remotely sensed nanophytoplankton biomass distribution is greatly improved with the implementation of locally tuned $k$ values (Figure 3 & Figure 4a-d). The use of optimised globally homogenous $k$ values (Global-$k$) improves the reproduction of observed nanophytoplankton but shows little of the regional variability found in observations (Figure 4a & c). This regional variability is only reproduced when locally tuned grazing dynamics are implemented (Figure 4d) highlighting the importance of top-down drivers for this size class. The Local-$k$ model does a good job in estimating global nanophytoplankton biomass with an annual mean (± S.D.) of 9.40±3 mgC m$^{-3}$, in comparison to 11.89±6 mgC m$^{-3}$ from satellite-derived estimates (Siegel et al., 2014) (Figure 4a). NPP is 52 and 43 Gt C yr$^{-1}$ for the Local-$k$ and Global-$k$ runs respectively, compared to 87 Gt C yr$^{-1}$ for the non-optimised baseline run (Figure S9).

Microplankton observational distributions are reproduced well in all three model versions showing little difference when locally tuned grazing dynamics are applied (Figure 4e-h), suggesting the distribution for this functional group is determined primarily by realistic bottom-up drivers. However, all three models overestimate biomass in equatorial upwelling areas (Figure 3). In the Local-$k$ model, microplankton has a global mean (± S.D.) of 3.05±3 mgC m$^{-3}$, compared to 2.62±5 mgC m$^{-3}$, from satellite-derived estimates (Siegel et al., 2014) (Figure 4e). In the subtropical oligotrophic gyres, microplankton biomass estimates appear to closely emulate observations, despite higher cost values in this region. This is due to the low biomass in the region, which results in small changes producing large error values with normalisation.

Local-tuning of $k$ values improves zooplankton biomass estimates in comparison to observations (Figure 4i-l & Figure S10). When only optimised homogenous global values are used, global mesozooplankton biomass is underestimated, with a mean (± S.D.) of 1.72±0.81 mgC m$^{-3}$ compared to 5.52±9 mgC m$^{-3}$ from (Strömberg et al., 2009) (Figure 4i). This is as a result of the very high $k$ value for mesozooplankton (392 mgC m$^{-3}$), resulting in very low grazing and therefore biomass. In contrast, mean (± S.D.) mesozooplankton biomass from the Local-$k$ model is 5.07±3 mgC m$^{-3}$. All model versions underestimate mesozooplankton biomass in the higher latitudes of the northern hemisphere. Mean microzooplankton biomass estimates are greatly reduced with the implementation of locally-tuned grazing dynamics, from 29.09±23 mgC m$^{-3}$ in the Global-$k$ run to 8.85±11 mgC m$^{-3}$ in the Local-$k$ run (Table 3).

3.4 Impact on Carbon Export

Local tuning of $k$ values decreases mean total carbon export by >17% (Table 3). The magnitude of change depends on the homogenous $k$ values used for comparison (-35.64% in comparison to the Baseline and -17.07% in comparison to the Global-$k$ scenario). Export values are generally high, with a total export flux of 7.19 PgC yr$^{-1}$ for the Local-$k$ scenario and 8.16 Pg C yr$^{-1}$ and 10.94 Pg C yr$^{-1}$ for the Global-$k$ and Baseline scenarios respectively.

The routing of carbon is impacted by the implementation of locally-tuned grazing dynamics (Figure 5). In the Local-$k$ scenario, more carbon is exported as faecal pellets and less as algal aggregates, compared to the Global-$k$ scenario (Table 3). When $k$ values are locally tuned, carbon exported from pellets and aggregates are more similar in magnitude (annual mean of 36.91 and 20.52 mgC m$^{-2}$d$^{-1}$ respectively) compared to model runs with homogenous $k$ values. In contrast, in both the Baseline and Global-$k$ model runs carbon export is dominated by algal aggregates. Local-tuning of $k$ values also results in increased export (>21%) via vertically migrating zooplankton compared to the Global-$k$ model run (Figure 5, Table 3).

Productive upwelling regions, that are characterised by high carbon export rates, are the regions of greatest change from the local tuning of $k$ values (Figure 5). These patterns
Figure 3. Absolute bias in modelled nanophytoplankton (PS) and microphytoplankton (PL) biomass in comparison to satellite derived biomass (P(i)mod - P(i)obs). Three model scenarios are shown: Baseline (non-optimised $k$ values), Global-$k$ (globally optimised $k$ value) and Local-$k$ (locally tuned $k$ values). Observational phytoplankton biomass values were calculated as per Siegel et al. (2014), using particulate backscattering coefficient data (Behrenfeld et al., 2005; Westberry et al., 2008; Kostadinov et al., 2010; Siegel et al., 2013).

4 Discussion

One of the largest sources of uncertainty in the marine carbon cycle is zooplankton grazing (Rohr et al., 2023a). In this study we used an inverse modelling approach to estimate spatial variation in zooplankton grazing dynamics and explored the subsequent
impact of these dynamics on modelling marine ecosystems and carbon export. The focus of this study was the grazing parameter $k$, which is frequently used in global biogeochemical (BGC) models. We found that local tuning of $k$ results in high variability of inferred grazing dynamics. The local-tuning of $k$ values improved the model’s ability to reproduce satellite-derived phytoplankton biomass. Consequently, estimates of mean total carbon export decreased by >17% compared to global tuning, with a greater proportion of export as faecal pellets and less as algal aggregates.

4.1 High Variability of Inferred Grazing Dynamics

Local optimisation suggested high $k$ values were generally associated with eutrophic ocean regions. This is consistent with a study in review (Rohr et al., 2023b) which used an inverse modelling approach to infer high community $k$ values for a single zooplankton group (combining $k_0$ and $k_1$) in equatorial upwelling regions and higher latitudes. In equatorial upwelling regions, communities are dominated by suspension-feeding copepods (Steinberg & Landry, 2016), whose slower grazing rates enable diatom blooms to form (Rohr et al., 2023b). In the higher latitudes, higher half-saturation constants reduce grazing pressure on nutrient-limited phytoplankton stocks with slow growth rates, which prevents prey stocks from being fully depleted (Schmoker et al., 2013). Here, higher $k$ values could characterise prey switching events or increased handling time. Christaki et al. (2021) found mesozooplankton in the Southern Ocean preferentially graze on microzooplankton over phytoplankton, due to slow growth rates of the latter.

Generally low $k$ values were estimated in the subtropical oligotrophic gyres, where communities are dominated by faster grazing microzooplankton, in particular pico- and nano-sized flagellates (Calbet & Calbet, 2008). Anomalous high $k_0$ values inferred in the hyper-oligotrophic South Pacific gyre (Ras et al., 2007) are in disagreement to the study by Rohr et al. (2023b) and coincide with an underestimation of NPP, nanophytoplankton...
Table 3. Comparison of mean global carbon export estimates from the three model scenarios. NPP = Net Primary Productivity. $F_{alg}$ = Euphotic export flux of algal aggregates. $F_{fec}$ = Euphotic export flux of faecal pellets. $J_{dvm}$ = DVM-mediated export flux. $G_{ZL}$ = Mesozooplankton grazing rate on all prey types. $G_{ZS}$ = Microzooplankton grazing rate on all prey types. $G_{ZL,PL}$ = Mesozooplankton grazing on microphytoplankton. $G_{ZL,ZS}$ = Mesozooplankton grazing on microzooplankton. $PS$ = Nanophytoplankton biomass. $PL$ = Microphytoplankton biomass. $ZS$ = Microzooplankton biomass. $ZL$ = Mesozooplankton biomass. DER = DVM export ratio. DRR = DVM respiration ratio.

<table>
<thead>
<tr>
<th></th>
<th>Baseline</th>
<th>Global-k</th>
<th>Local-k</th>
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<tbody>
<tr>
<td></td>
<td>Mean</td>
<td>S.D.</td>
<td>Mean</td>
</tr>
<tr>
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<td>127.46</td>
<td>69.44</td>
</tr>
<tr>
<td>NPP (mgC m$^{-2}$d$^{-1}$)</td>
<td>752.18</td>
<td>715.79</td>
<td>379.36</td>
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<tr>
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<tr>
<td>PS (mgC m$^{-3}$)</td>
<td>13.95</td>
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</tr>
<tr>
<td>PL (mgC m$^{-3}$)</td>
<td>4.04</td>
<td>3.52</td>
<td>4.38</td>
</tr>
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<tr>
<td>$G_{ZL,PL}$ (mgC m$^{-3}$d$^{-1}$)</td>
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<td>0.25</td>
<td>1.7x10$^{-4}$</td>
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<td>$G_{ZL,ZS}$ (mgC m$^{-3}$d$^{-1}$)</td>
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<td>1.91</td>
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<td>$F_{alg}$ (mgC m$^{-2}$d$^{-1}$)</td>
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<td>68.28</td>
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<tr>
<td>$F_{fec}$ (mgC m$^{-2}$d$^{-1}$)</td>
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<td>33.14</td>
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<td>$J_{dvm}$ (mgC m$^{-3}$d$^{-1}$)</td>
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<tr>
<td>DER</td>
<td>0.03</td>
<td>0.02</td>
<td>3.2x10$^{-3}$</td>
</tr>
<tr>
<td>DRR</td>
<td>0.03</td>
<td>0.02</td>
<td>2.7x10$^{-3}$</td>
</tr>
</tbody>
</table>

Biomass and near-zero growth rates. This suggests that in this region, nanophytoplankton growth rates used to force the model may be too low. Growth rates are derived from observed NPP, which is divided by the greater of euphotic and mixed layer depth. The South Pacific gyre is characterised by a deep euphotic layer, which may have produced unrealistically low growth rates and anomalous $k$ values for this size class. In addition, the lack of explicit representation of temperature within the model may affect growth and grazing rates in these extreme environments. The 1P1Z 3D model in Rohr et al. (2023b) uses modelled rather than observationally derived bottom-up controls which may negate these issues.

In this study, mesozooplankton showed on average, higher $k$ values, and therefore slower prey capture times. This is consistent with ecological understanding (Barton et al., 2013). As the maximum grazing rate, $g$, was held constant across the model domain, variation in the $k$ value represents variation in the prey capture rate, rather than consumption (Rohr et al., 2022). For both size classes, average half-saturation constants were in the lower quartile of empirical values (Rohr et al., 2022; Hansen et al., 1997; Hirst & Bunker, 2003). However, empirical estimates are from laboratory measurements of samples collected from a very narrow range of locations, with the majority from coastal regions in the northern hemisphere (e.g. fjords in Norway, coastal USA and UK, Japan) and none representing the open ocean (Hansen et al., 1997; Hirst & Bunker, 2003). These are also of individual species and are unlikely to be representative of the community mean values estimated here for each 1 degree grid cell (Rohr et al., 2022).
Figure 5. Changes in carbon export due to grazing parameterisation. Three model runs are presented: Baseline, Global-k and Local-k. The outputs from the Baseline run are presented in plots a-f. Plots g-l show the absolute change when changing the model input from the baseline run (non-optimised $k$ values) to the Global-k run (globally optimised $k$ values). Plots m-r show the absolute change when changing the model input from the Global-k run (globally optimised $k$ values) to the Local-k run (locally tuned $k$ values). $F_{alg}$ = Euphotic export flux of algal aggregates. $F_{fec}$ = Euphotic export flux of faecal pellets. $J_{dvm}$ = DVM-mediated export flux. $G_{ZL}$ = Mesozooplankton grazing rate on all prey types. $G_{ZS}$ = Microzooplankton grazing rate on all prey types.

Locally-tuned $k$ values produced global distributions of grazing rates ($G_i$) that are consistent with other studies (Siegel et al., 2014; Archibald et al., 2019). Archibald et al. (2019) used satellite-derived estimates of Net Primary Productivity (NPP) and phytoplankton biomass to predict global grazing rates. Mean $G_{ZS}$ was 4.17 mgC m$^{-3}$ d$^{-1}$ and mean $G_{ZL,PL}$ was 0.98 mgCm$^{-3}$d$^{-1}$ in Archibald et al. (2019). These represent grazing mortalities from satellite observations on phytoplankton, so include grazing losses by other groups not considered here (e.g. mesozooplankton grazing on nanophytoplankton). However, the Local-k model improves the reproduction of these observationally derived grazing rates in comparison to globally homogenous $k$ values (Table 3), with the potential to help address the uncertainty in global zooplankton grazing dynamics.
4.2 Implications of Improved Model Skill for Future Studies

With potential to improve model skill, the local optimisation of grazing dynamics could be advisable in future BGC modelling studies. This study shows that the competition generated by two zooplankton functional types isn’t sufficient to emulate the global variability in grazing suggested by the locally tuning \( k \) values. Other models currently emulate this dynamical variability using different methods, such as increased numbers of plankton functional groups (PFTs) (e.g. Dutkiewicz et al. (2021), prey switching (e.g. Anderson et al. (2010)), or prey preferences (e.g. Aumont et al. (2015)). In this study, four PFTs were used, in line with several modelling studies (e.g. Siegel et al. (2014)), however, this groups together species with different functional traits, with different geographic distributions (Barton et al., 2013). Gelatinous salps, for example, graze preferentially on nanophytoplankton, which leads to their prevalence in subtropical oligotrophic gyres (Barton et al., 2013). By explicitly representing more PFTs and their prey preferences, some of the impact of locally-tuning \( k \) values may be reduced. A study by Le Quéré et al. (2016) showed that the explicit representation of krill in the Southern Ocean improved model skill in reproducing zooplankton dynamics. However, increasing the number of PFTs is computationally costly and unlikely to encompass the full extent of physiological and behavioural diversity found within the plankton community. The global distributions of the grazing parameter \( k \), produced in this study, could provide a platform for varying zooplankton grazing parameters in larger, more complicated BGC models, with environmental conditions. If the variability in local tuned \( k \) values correlates with key environmental variables, then grazing dynamics could be implemented as a function of covariates. This could improve model skill in comparison to the use of globally homogenous \( k \) values, whilst remaining computationally effective. This work is beyond the scope of this study but is an area for further work.

4.3 Reducing Uncertainty in Modelling Zooplankton

Large uncertainty exists in quantifying zooplankton biomass (Petrik et al., 2022), however the use of locally tuned \( k \) values improves the models ability to reproduce observational estimates. The observed mean (±S.D.) biomass of mesozooplankton is estimated as 5.9±10.6 mgC m\(^{-3}\) by Buitenhuis et al. (2013); Moriarty and O’Brien (2013), which compares to 5.07 ±2.7 mgC m\(^{-3}\) in the Local-\( k \) model scenario. In contrast, optimised globally homogenous \( k \) values worsen the model’s ability to reproduce observational mesozooplankton biomass in comparison to the Baseline scenario (Table 3). This highlights a potential limitation of global optimisation of grazing dynamics when only two zooplankton functional groups are used. This also suggests a possible reason why several BGC models underestimate mean global mesozooplankton biomass by a similar magnitude (Figure S10) (Aumont et al., 2015; Lovato et al., 2022). In addition, mean global microzooplankton estimates from the Global-\( k \) scenario are much greater than observations by Buitenhuis et al. (2013), where biomass is estimated to have a mean of 9.3±17.1 mgC m\(^{-3}\) and a median of 3.1 mgC m\(^{-3}\). These limitations occur despite improved model skill at reproducing the magnitude and distribution of satellite-derived phytoplankton. Local-\( k \) mean global biomass for microphytoplankton is within the range of estimates by Buitenhuis et al. (2013)(Table 3).

4.4 Implications for Predicting Carbon Export under a Changing Climate

This study shows large variability in carbon export estimates driven by inferred grazing dynamics. There is a high degree of uncertainty in global export flux estimates, which vary between 5-12 PgC yr\(^{-1}\) (Siegel et al., 2022). Locally tuning grazing dynamics modifies carbon export estimates by >17% to coincide with this range (7.19 PgC yr\(^{-1}\)). The Global-\( k \) scenario estimates high carbon export, despite underestimating mesozooplankton biomass and therefore fecal export. This is due to a large proportion of algal aggregates. The substantial influence of this one model component on carbon export highlights one possible cause for uncertainty in carbon sequestration estimates (Laufkötter et al., 2016). It is vital to reduce this uncertainty when modelling under different climate scenarios.
Algal and fecal estimates were closer to contributing equally to carbon export (as found within previous studies (Stock et al., 2014; Steinberg & Landry, 2016)) when grazing dynamics were locally optimised. The relationship between algal aggregates and faecal export highlights the balance between natural and grazing mortalities within BGC models, as the amount of biomass available for aggregates from mortality rates is impacted by grazing. The proportion of faecal versus algal export is determined by the zooplankton species present and their grazing dynamics (Steinberg & Landry, 2016). Local tuning of grazing dynamics therefore has important consequences for climate models as the impact of climate change on plankton communities differs between species, trophic levels and geographic location (Cael et al., 2021), further influencing these two routes of carbon export.

The modelled distribution of carbon export corresponds to estimates by Stock et al. (2020), with coastal upwelling areas experiencing the greatest change from the local tuning of $k$. These areas produce the highest export rates due to more efficient diatom-copepod food chains (Schmoker et al., 2013). However, the fast growth rates of diatoms makes them more susceptible to abrupt changes over the 21st Century in response to climate change (Cael et al., 2021). It is therefore vital to decrease uncertainty in grazing and export estimates in these areas. The subtropical oligotrophic gyres and the Southern Ocean are areas of lower carbon export due to the presence of smaller phytoplankton species which are lighter, sinking less carbon into the ocean interior (Murphy et al., 2021; Schmoker et al., 2013; Calbet & Landry, 2004). The highest flux estimates in the Southern Ocean occur closer to the Antarctic shelf edge, predominantly during summer months (Stock et al., 2020). However the polar extremes are out of the scope of this study due to the limitations of satellite observations in these areas (Siegel et al., 2014), which is a common issue with many plankton models (Cael et al., 2021; Dutkiewicz et al., 2021).

4.5 Limitations

There are several limitations of this modelling study. Firstly, non-$k$ parameters are held constant across the model domain, when in reality they are likely to vary in space and time. If these other parameters were tuned, the non-optimised (Baseline scenario) estimate of NPP may be reduced to coincide with the observed range (45-60 GtC yr$^{-1}$) (Westberry et al., 2008; Le Quéré et al., 2016) alongside the two optimised model scenarios. $G_{Z1,PL}$ and $G_{Z1,ZS}$ also use the same $k$ value, which obscures whether changes in $k_1$ are biased to improve microphytoplankton estimates or nanophytoplankton estimates (via microzooplankton) during the cost analysis. This may contribute to the overestimation of microphytoplankton in all model runs. Secondly, grazing formulas are based on the Holling Type III functional response; however there is a lack of consensus within the modelling community about the most suitable functional response. Anderson et al. (2010) found that the use of different grazing formulations caused large variations in biomass, with diatoms most greatly affected. This resulted in carbon export predictions varying by as much as 25%.

Thirdly, within the centre of the oligotrophic gyres, microzooplankton were characterised by less efficient prey capture rates compared to mesozooplankton (Figure S5), however prey capture efficiency should decline with size (Rohr et al., 2022; Hansen et al., 1997). This highlights a possible limitation of the model, potentially the functional groups used or model parameters. In this study, the same maximum grazing rate was used for both size classes, so prey capture efficiency is dominated by the half-saturation constant, or capture rate. In oligotrophic regions, smaller plankton dominate, so prey capture efficiencies for microzooplankton are more likely to be driven by consumption rather than capture rates, suggesting higher maximum grazing rates are needed to represent realistic capture efficiencies in these environments. Fourthly, CbPMv2 was selected as the NPP forcing variable for consistency with Archibald et al. (2019), however other NPP models such as the The Carbon, Absorption, and Fluorescence Euphotic-resolving (CAFE) (Silsbe et al., 2016) have been found to be more realistic, with consequences for carbon export (Bisson et al., 2018). Finally, the carbon export model does have several limitations which are discussed in detail in Archibald et al. (2019). In particular, the model is very sensitive to three parameters – $f_{sec}$, $f_{met}$.
and pdcm, however these parameters remain unchanged to enable comparisons. The model also doesn't include small fecal pellets produced by microzooplankton in carbon export estimates as they are assumed to be retained in the euphotic zone. However, some studies suggest fecal export by this size class could contribute a significant portion of export flux, particularly in the subtropical oligotrophic gyres Bisson et al. (2020).

4.6 Future Considerations

The simplification of the coupled ecosystem-carbon export model means the results of this study should be considered as an example of an ecosystem model with and without spatially varying $k$ values. Here, we have shown that highly spatially heterogeneous grazing dynamics are required to reproduced observed biomass when forced with observed bottom-up controls. This heterogeneity exceeds what is achievable from the explicit competition between two zooplankton functional types and has profound implications for the routing and magnitude of carbon export. Future models, particularly those concerned with ecosystem dynamics, high trophic levels and carbon export must reconcile with the possibility that even two zooplankton groups are insufficient to capture the true variability in top-down controls across the globe. More realistic representation of the global variability in zooplankton grazing dynamics may help shed light on the uncertainty in carbon export estimates under future climate scenarios.

5 Open Research

Climatologies used here are presented in detail in Siegel et al. (2014). CbPMv2 (Westberry et al., 2008) Net Primary Productivity, particulate backscatter (used to derive phytoplankton carbon biomass) and mixed layer depth data can be sourced from: http://orca.science.oregonstate.edu/app_products.php. World Ocean Atlas temperature and oxygen data used in the Archibald et al. (2019) carbon export model can be found at: (https://coastwatch.pfeg.noaa.gov/erddap/index.html).

Acknowledgments

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References


Supporting Information for "Impact of spatial variability in zooplankton grazing rates on carbon export flux"

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Figure S1. Locally-tuned microzooplankton half-saturation constant ($k_0$) estimated using alternate cost functions. a) MWAAE = Absolute average error (Stow et al., 2009) normalised using the observational mean instead of the standard deviation (as per the main article). b) RMSE = the Root Mean Squared Error (Stow et al., 2009), normalised by standard deviation of observations. c) Bias = Average Error or Bias (Stow et al., 2009), normalised by standard deviation of observations. Plots show very similar results are produced, no matter the cost function used.
Figure S2. Locally-tuned mesozooplankton half-saturation constant ($k_1$) estimated using alternate cost functions. a) MWAAE = Absolute average error (Stow et al., 2009) normalised using the observational mean instead of the standard deviation (as per the main article). b) RMSE = the Root Mean Squared Error (Stow et al., 2009), normalised by standard deviation of observations. c) Bias = Average Error or Bias (Stow et al., 2009), normalised by standard deviation of observations. Plots show very similar results are produced, no matter the cost function used.
Table S1. Average locally-tuned microzooplankton ($k_0$) and mesozooplankton ($k_1$) half-saturation constants estimated using alternate cost functions. Units are in mgC m$^{-3}$. MWAAE = absolute average error (Stow et al., 2009), normalised using the mean of observations. RMSE = the Root Mean Squared Error (Stow et al., 2009), normalised using the standard deviation of observations. Bias = annual average Error or Bias (Stow et al., 2009), normalised using standard deviation of observations.

<table>
<thead>
<tr>
<th>Cost function</th>
<th>$k_0$</th>
<th>$k_1$</th>
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<td>Median</td>
<td>Mean</td>
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<td>71</td>
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<tr>
<td>RMSE</td>
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<td>72</td>
</tr>
<tr>
<td>Bias</td>
<td>18</td>
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</table>

Figure S3. Locally-tuned microzooplankton half-saturation constant ($k_0$) estimated using nAAE cost function (Equation 12) from a) December-February, b) March-May, c) June-August and f) September-November. Figures show consistency across seasons.
**Figure S4.** Locally-tuned mesozooplankton half-saturation constant ($k_1$) estimated using nAAE cost function (Equation 12) from a) December-February, b) March-May, c) June-August and f) September-November. Figures show consistency across seasons.

**Figure S5.** Prey capture efficiencies. Values represent the log ratio of prey capture efficiencies for micro- and mesozooplankton, or $\varepsilon_0$ and $\varepsilon_1$, respectively. The prey capture efficiency is calculated by dividing the maximum grazing rate (See Table 1) by the half-saturation constant for each grid cell (Rohr et al., 2022)
Table S2. Seasonal averages of locally-tuned microzooplankton ($k_0$) and mesozooplankton ($k_1$) half-saturation constants estimated using nAAE cost function (Equation 12). Units are in mgC m$^{-3}$. DJF = December-February, MMA = March-May, JJA = June-August and SON = September-November.

<table>
<thead>
<tr>
<th>Season</th>
<th>$k_0$ Median</th>
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<th>$k_1$ Median</th>
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<td>29</td>
<td>94</td>
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<tr>
<td>MMA</td>
<td>18</td>
<td>71</td>
<td>29</td>
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<tr>
<td>JJA</td>
<td>18</td>
<td>72</td>
<td>29</td>
<td>93</td>
</tr>
<tr>
<td>SON</td>
<td>18</td>
<td>72</td>
<td>290</td>
<td>95</td>
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</table>

Table S3. Breakdown of mean cost estimates form the Local-$k$ model run into its constituent parts (Equations 12-14). Cost consist of normalised Absolute Average Error values (nAAE) for each size class. Cost indicates model fit against satellite observations of phytoplankton biomass. A value of zero indicates a perfect match with satellite observations. DJF = December-February, MAM = March-May, JJA = June-August, SON = September-November, PS = nanophytoplankton, PL = microphytoplankton.

<table>
<thead>
<tr>
<th>Time Period</th>
<th>AAE PS</th>
<th>AAE PL</th>
<th>nAAE PS</th>
<th>nAAE PL</th>
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<td>2.76</td>
<td>4.02</td>
<td>18.95</td>
<td>-</td>
</tr>
</tbody>
</table>

December 19, 2023, 4:06pm
Figure S6. Comparison of half-saturation constant values from local optimisation with variables a) Microzooplankton half-saturation constant ($k_0$) and model-derived NPP from nanophytoplankton (NPPn), b) Mesozooplankton half-saturation constant ($k_1$) and model-derived NPP from microphytoplankton (NPPm), c) Microzooplankton half-saturation constant ($k_0$) and Nanophytoplankton Biomass (PS), d) Mesozooplankton half-saturation constant ($k_1$) and Microphytoplankton Biomass (PL). All values are log-transformed. Units are mgC m$^{-3}$ for biomass and $k$ values, and mgC m$^{-2}$d$^{-1}$ for NPP. Red line = linear regression.
Figure S7. Cost values from the Local-$k$ model are the sum of the a) Normalised Absolute Average Error for nanophytoplankton biomass, and the b) Normalised Absolute Average Error for microphytoplankton biomass.
Figure S8. Normalised Absolute Average Error (nAAE) values for observations and model phytoplankton biomass estimates for each season. DJF = December-February. MAM = March-May). JJA = June-August. SON = September-November. Estimates are for both nanophytoplankton (a,c,e,g) and microphytoplankton biomass (b,d,f,h).
Figure S9. a) Local-\(k\) model Net Primary Productivity (NPP), b) Satellite-derived NPP from Westberry et al. (2008).
Figure S10. Global mean mesozooplankton (Z) estimates from this study, compared with other model and empirical values. Blue indicates estimates derived from observations. Green indicates model based estimates. Purple indicates estimates from this study from three different scenarios: Baseline scenario with non-optimised globally homogenous \( k \) values; Global-\( k \) scenario with globally optimised homogenous \( k \) value for each size class; Local-\( k \) scenario with locally optimised \( k \) values. Sources of data are: STROM= Strömberg et al. (2009); MARE= Buitenhuis et al. (2013); GLMM= Heneghan et al. (2020); CPOD=Moriarty and O’Brien (2013); CMCC= BFMv5.2 (Lovato et al., 2022); IPSL = PISCES2.0 model (Aumont et al., 2015), UK = MEDUSA2.1 model (Yool et al., 2013, 2021), GFDL = COBALTv2 model (Stock et al., 2020). See Petrik et al. (2022) for description of zooplankton estimates from other model and empirical estimates.
Figure S11.  a) Local-$k$ model nanophytoplankton growth rate, b) Local-$k$ model microphytoplankton growth rate, c) $Z_{eu}$, depth of the euphotic layer, d) $Z_{ml}$, mixed layer depth. Note the different scales for the growth rate figures.
Figure S12. Changes in carbon export due to grazing parameterisation. Three model runs are presented: Baseline, Global-$k$ and Local-$k$. The outputs from the Baseline run are presented in the left-hand column. Plots in the middle column show the absolute change when changing the model input from the baseline run (non-optimised $k$ values) to the Global-$k$ run (globally optimised $k$ values). Plots in the right column show the absolute change when changing the model input from the Global-$k$ run (globally optimised $k$ values) to the Local-$k$ run (locally tuned $k$ values). $f_{Eu}$ = Total euphotic zone export flux as a fraction of NPP. $f_{DVM}$ = DVM-mediated export flux as a fraction of NPP. NPP = Net primary Productivity, ER= export ratio, DER=DVM export ratio, DRR= DVM respiration ratio. RD = Respiration Depression.
Figure S13. Microzooplankton biomass estimated by the model under three scenarios: Baseline, Global-\(k\) and Local-\(k\).