# Variable species establishment in response to microhabitat indicates different likelihoods of climate-driven range shifts

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October 9, 2023

#### Abstract

Climate change is causing geographic range shifts globally, and understanding the factors that influence species' range expansions is crucial for predicting future changes in biodiversity. A common, yet untested, assumption in forecasting approaches is that species will shift beyond current range edges into new habitats as they become macroclimatically suitable, even though microhabitat variability could have overriding effects on local population dynamics. We aim to better understand the role of microhabitat in range shifts through its impacts on establishment by i) examining microhabitat variability along large macroclimatic gradients, ii) testing which of these microhabitat variables explain plant recruitment and seedling survival, and iii) predicting microhabitat suitability beyond species range limits. We transplanted seeds of 25 common tree, shrub, forb, and graminoid species across and beyond their current elevational ranges in the Washington Cascade Range, USA, along a large elevational gradient spanning a broad range of macroclimates. Over five years, we recorded recruitment, survival, and microhabitat characteristics rarely measured in biogeographic studies. We asked whether microhabitat variables correlate with elevation, which variables drive species establishment, and whether microhabitat variables important for establishment are already suitable beyond leading range limits. We found that only 30% of microhabitat parameters covaried in the expected way with elevation. We further observed extremely low recruitment and moderate seedling survival in our study system, and these were generally only weakly explained by microhabitat. Moreover, species and life stages responded in contrasting ways to soil biota, soil moisture, temperature, and snow duration. Microhabitat suitability predictions suggest that distribution shifts are likely to be species-specific, as different species have different suitabilities, and availabilities, of microhabitat beyond their present ranges, thus calling into question large-scale macroclimatic projections that will miss such complexities. We encourage further research on species responses to microhabitat and the inclusion of microhabitat in range shift forecasts.

### Introduction

The most pressing environmental issues of our times are understanding the ecological effects of ongoing climate change and predicting the ensuing implications for maintaining biodiversity (Lovejoy & Hannah, 2019). Foundational to these issues are an understanding of species range limits, which are the geographic limits to species' spatial distributions. Projection inaccuracy of how species ranges will shift with climate change is alarming (Urban, 2019), and it is unclear whether our inability to project species range shifts comes from our poor knowledge of which climatic variables are important for which species or from the many other factors that can influence range shifts (e.g. microhabitat variation, dispersal limitation, species interactions, and many others). Even explaining the mechanisms underlying observed shifts is difficult, and it is clear that responses are highly species-specific (Freeman et al., 2018; Rumpf et al., 2018).

To address this issue, ecologists are urgently seeking to understand species niche limits in order to project species distribution shifts with climate change. These are commonly predicted with correlative Species Distribution Models (SDMs; Elith et al., 2010; Pacifici et al., 2015), which correlate species occurrences to current macro-climate in order to predict the relative probability of occurrence in space and time (Wiens et al., 2009). Such models have become an integral part of biogeographical research and progress has been made to address some of their shortcomings (Franklin, 2023). While these models implicitly assume that they include sufficient niche variables to accurately describe a species distribution, few models actually test what niche components are necessary to quantify population growth rate and shape species distributions (Pulliam, 2000).

A main driver of population growth rate is the fine-scale microhabitat that mediates the success of individual plants through abiotic (e.g., canopy cover, above-ground temperature, soil moisture) and biotic (e.g., soil biota, soil nutrients) conditions (Kephart & Paladino, 1997; Zurbriggen et al., 2013; Oldfather & Ackerly, 2019; Tanner et al., 2021; Sanczuk et al., 2023; Allsup et al., 2023; Kemppinen et al., 2023). However, studies that quantify population response to microhabitat variation along species ranges are rare, impeding an understanding of which aspects of fine-scale microhabitat are important in shaping species broad-scale distributions (but see Tourville et al., 2022). Despite this, recent work has still shown that incorporating microhabitat into SDMs can provide substantially improved predictions (Maclean & Early, 2023).

How microhabitat and microclimate themselves vary along large elevational, and therefore macroclimatic, gradients that characterize species ranges is poorly understood, and research is increasingly finding that microclimate is often decoupled from macroclimate (Scherrer & Körner, 2010; Ford et al., 2013; Lembrechts, 2023). It is, however, now widely accepted that microclimate differs from macroclimate due to factors such as forest canopy cover (De Frenne et al., 2019; Haesen et al., 2021) and topography (Lawson et al., 2014). This can modify species response to habitat use (Lawson et al., 2014) and create climate change refugia (Dobrowski, 2011; Pradhan et al., 2023) that mediate species response to climate change (Morelli et al., 2020). Very few studies have been able to document how variable plant-relevant environmental drivers of performance are across macroscale gradients, although substantial progress is underway in including microclimate as part of biogeographical research (Kemppinen et al., 2023).

Testing how sensitive species' life stages are to variations in microhabitat can address many of the sources of bias that distribution models face and accounting for demographic processes can greatly improve our understanding of range dynamics (Fig. 1; Normand et al., 2014; Copenhaver-Parry et al., 2020). For plant species, how early life stages (i.e. recruitment and seedling survival; henceforth establishment) respond to microhabitat may be the key to determining species' ability to expand their ranges (Kroiss & HilleRisLambers, 2015), as establishment is essential for a range expansion to occur. Understanding how early life stages respond to variations in microhabitat is thus critical to understanding microhabitat suitability for range shifts and population persistence, especially since life stages can respond differently to environmental conditions (Goodwin & Brown, 2023). Yet, few studies test how microhabitat influences establishment, and important microhabitat variables, such as soil moisture and temperature, are often left out of such work (Schurr et al., 2012).

Species show remarkable variability in their responses to climatic aspects of microhabitat (Bullied et al., 2012) and aspects of microhabitat not directly influenced by climate (Selmants et al., 2016; Castro et al., 2022). There are no general trends, except for that diverse components of microhabitat are important for establishment (Table S1). With regards to climatic aspects of microhabitat, soil moisture and temperature, plant-height temperature, and snow duration can affect establishment in a myriad of ways (Szeicz & Macdonald, 1995; Thompson & Naeem, 1996; Yates et al., 1996; Graae et al., 2009; Santana et al., 2010; Rodríguez-García et al., 2011; Bullied et al., 2012; Moyes et al. 2013; Caldeira et al., 2014; Mondoni et al., 2015; Renard et al., 2016; Kueppers et al., 2017; Andrus et al., 2018; Elliott & Petruccelli, 2018; Lett & Dorrepaal, 2018; Dolezal et al., 2021; Ósvaldsson et al., 2022)

How early life stages respond to microhabitat variables not directly related to climate is also variable by species, with linear, unimodal, or no responses to canopy cover (Lloret et al., 2005; Käber et al., 2021), soil

fungus and bacterial content (Rigg et al., 2016; van der Heijden et al., 2016; Tobias et al., 2017; Xi et al., 2018), soil carbon and nitrogen (Monaco et al., 2003; Pérez-Fernández et al., 2006; Li et al., 2011; Pröll et al., 2011; Bateman et al., 2017; Kołodziejek et al., 2017; Zhong et al., 2019), and water holding capacity (Moser et al., 2017; Smithers, 2017; James et al., 2019; Khurana & Singh, 2000). However, studies examining responses to microhabitat beyond range edges are rare and can show inconclusive effects of microhabitat (Lee-Yaw et al., 2016), highlighting the need to measure responses to microhabitat along multiple species' ranges.

To comprehensively understand how microhabitat, and other factors, influence species distributions, transplant experiments beyond species' ranges have been suggested as a promising approach (Lee-Yaw et al., 2016; Morris & Ehrlén, 2015). Transplant experiments show either congruence with SDM predictions (Sanczuk et al., 2022) or markedly different responses of populations than predicted by SDMs (Greiser et al., 2020), highlighting the importance of these experiments. Such experiments are particularly valuable at the leading edge of a species range (i.e. edge expanding with climate change), where novel species interactions are likely to be found (Thuiller et al., 2008). Transplant experiments often find that species are establishment, not dispersal, limited, and this can be due to unfavorable microsite conditions (Clark et al., 2007; Davis & Gedalof, 2018). How microhabitat suitability is distributed beyond species current leading edge therefore likely determines species' ability to shift their ranges (Tourville et al., 2022), yet few studies quantify microhabitat along and above species distributions. We utilized the large elevational macroclimatic gradients of the West and East sides of the Washington Cascade Range, USA for a seed transplant experiment encompassing common grasses, forbs, shrubs, and trees to ask:

- 1. How does microhabitat variation within sites compare to among-site macroclimate variation, and does microhabitat covary expectedly along elevational gradients?
- 2. Does microhabitat explain establishment of common species in our system, and if so, are responses consistent among species?
- 3. If microhabitat variables explain establishment, does the distribution of microhabitats along macroclimatic gradients suggest range shifts will be facilitated, constrained, or unaffected by the availability of suitable microhabitat?

# Materials and Methods

#### Study site

We utilized the large climatic gradients of the Cascade Range in Washington, USA to transplant seeds along each of 2 large transects spanning ~1200 m on the West and East side of the Cascade Crest on the traditional lands of the Nlaka'pamux, Nooksack, Okanagan, and Methow peoples. Both transects are characterized by topographically complex terrain and differ substantially in their macroclimatic characteristics. The West transect (Mount Baker National Forest) is warmer and wetter than the East transect (Okanagan National Forest) (Supporting Information). Both transects are characterized by montane to subalpine species common in the Pacific Northwest, with an abundance of cedars, firs, heathers, and understory forbs and grasses. We selected 15 sites along each of our 2 transects (n = 30 sites) using satellite images of accessible areas, and identifying areas (i.e. blocks) that had both low and high tree canopy openness. In the field, we established two blocks per site (n = 60 blocks) to encompass different levels of canopy openness, with one block in the relatively most open area and the other block in the relatively most closed canopy.

#### Species data

We sowed 25 species encompassing tree (Abies grandis, A. lasiocarpa, Picea sitchensis, P. engelmannii, Pinus ponderosa, P. contorta), shrub (Mahonia nervosa, M. aquifolium, Rubus ursinus, R. spectabilis, Sambucus cerulea, S. racemosa, Sorbus sitchensis, Vaccinium parvifolium, V. deliciosum), forb (Eriophyllum lanatum,

Anemone occidentalis, Erigeron perigrinus, Lupinus latifolius, Maianthemum dilatum, M. racemosum, Tolmiea menziesii, Tellima grandiflora ), and graminoid (Carex stipata, Carex spectabilis ) growth forms. We chose congeneric and confamilial pairs of species that have different regional distributional characteristics (e.g. lower vs. higher elevation, or wetter vs. drier side of the Cascade Crest), thus aiming to capture a range of species potential responses. To facilitate field identification of seedlings, we sowed these pairs onto separate quadrats within each block for three plots (n = 180 plots) of three 0.25 x 0.25 m quadrat replicates (n = 540 replicates), leaving the third quadrat unmanipulated to control for background recruitment. We also included unpaired species with large seeds (L. latifolius ) or with high regional prevalence (S. sitchensis, A. occidentalis ). We opportunistically sourced seeds from nearby areas in 2016 and purchased native seeds for those species for which we had no, or not enough, locally sourced seeds.

We homogenized all seed sources for a given species and sowed seeds in a sand mixture in September-October 2017. We recorded recruitment (i.e. survived to end of first growing season) and yearly survival of seedlings during the growing season (May-September) in 2018, 2019, and 2020, and recorded only surviving seedlings in 2022. We surveyed sites three times during the growing season in 2018 and 2019, and visited sites once at the end of the growing season in 2020 and 2022. We were not able to access 10 of our sites in 2018 (wildfire closures) or any of our field sites in 2021 (closed USA border).

#### Microhabitat data

We measured a suite of abiotic and biotic parameters to quantify microhabitat (Table S1) and categorize these parameters as being directly influenced by climate change or not. We measured some of these variables annually throughout the first three years of the study period and others in just the last year of the study (due to logistical and financial constraints). Here, we assume that the rank order of microhabitat differences among sites remained constant across years.

We measured aspects of microhabitat directly related to climate by recording soil and air temperature, duration of snow cover, and soil moisture at each block with two different data loggers (TMS-4 data logger, TOMST, Prague Czech Republic; Wild et al., 2019; HOBO 64K Pendant Temperature/Alarm data loggers, Onset, Bourne, Massachusetts USA). We calculated seasonal variables from these data loggers, using some functions from package 'myClim' (Man et al., 2023; Man, Kalčík, Macek, Wild, et al., 2023) to generate the following biologically meaningful microclimate explanatory variables: summer maximum soil temperature, winter minimum soil temperature, spring days of snow coverage, summer minimum soil moisture, and summer minimum plant-height temperature (Table S1, Supporting Information).

We also measured aspects of microhabitat not directly related to climate by quantifying abiotic and biotic soil microhabitat, namely soil fungal, bacterial, organic carbon and nitrogen content, and water holding capacity at each replicate to capture variability within each site. We also measured canopy openness at each block. Upon individual site snow melt-out in 2022, we took soil cores to conduct *in situ*measurements of fungus:bacteria ratio (F:B) with a microBIOMETER rest Kit (Prolific Earth Sciences, Montgomery, New York, USA). We then quantified soil organic carbon to nitrogen ratio (C:N) and water holding capacity in the lab (Supporting Information).

#### Statistical analyses: microhabitat variation

To answer how variation within sites compares to among sites of each of our microhabitat parameters, we calculated the variance partition coefficient from intercept-only linear mixed models (LMMs; package 'lmerTest'; Kuznetsova et al., 2017) that included block nested within site as a random effect (microhabitat variable  $\sim 1 + (1|\text{site}/\text{block})$ ). In cases where LMMs did not converge, we changed the random effect to include only site. To test if microhabitat parameters covary with elevation, we fit LMMs with quadratic elevation, transect, and their interaction and included site as a random effect (microhabitat variable  $\sim$  poly(elevation, 2) \* transect + (1|site)). We also used a principal component analysis plot to identify any clustering in the microhabitat data. We conducted all data processing and analyses in R version 4.2.3 (R Core Team, 2023).

#### Statistical analyses: species establishment

To interpret our results in terms of likelihood of recruitment, recruit numbers, and seedling survival, we first transformed our species-level yearly recruitment and seedling survival data to binary or proportional responses by calculating: binary recruitment, relative recruit counts in the first three years (2018, 2019, 2020), 1-year (2018-2019 or 2019-2020), 2-year (2018-2020), and multi-year (survived to 2022) survival of seedlings. We controlled for background recruitment by subtracting the recruit counts in control plots from paired seed addition plots. From this, we then calculated relative recruit counts: recruit counts<sub>species i</sub>, site-rep j / maximum recruit counts<sub>species i</sub>. We calculated the proportion of surviving seedlings in all replicates, including control replicates, using all non-zero recruitment data and not allowing survival probability > 1.

While we have general *a priori* hypotheses of how certain microhabitat conditions affect species establishment (Table S1), previous studies have shown that recruitment and seedling survival vary greatly by species and we have limited prior knowledge of which microhabitat parameters are important for each species. We thus chose a data exploratory framework and used an information-theoretic approach with model averaging ('MuMIn' package, Bartoń, 2022) to compare ecologically meaningful microhabitat variables to identify the most suitable ones for each species (Tredennick et al., 2021). We fit separate binomial generalized linear models (GLMs) for each species' life stage to determine which of the microhabitat variables described above (uncorrelated with Pearson's correlation coefficient < 0.7) are important for establishment (i.e. recruitment and seedling survival). We only analyzed species for which we observed a response in at least 8 plots for any given life stage.

Each of our global GLMs included all 8 microhabitat variables described above, plus any quadratic effects identified as important by AICc in a reduced model ('AICcmodavg' package; Mazerolle, 2020; Supporting Information). To account for soil temperature effects on recruitment and plant-height temperature on seedling survival, we fit recruitment models with soil temperature and seedling survival models with plant-height temperature. To account for microhabitat variation not captured by any of these parameters, we further included canopy openness and transect as fixed effects (life stage success  $\sim$  microhabitat variables + canopy openness + transect, family = binomial(link = 'logit')). In 1-year survival models, we also included year as a fixed effect to account for differences among years in overall seedling survival and different frequencies of site visits (life stage success  $\sim$  microhabitat variables + canopy openness + transect + year, family = binomial(link = 'logit')).

We removed variables with a variance inflation factor > 5 ('car' package; Fox et al., 2023) to reduce parameter collinearity, and restricted the models used in model selection to have N/10 maximum parameters to avoid fitting overly complex models. Where there were multiple GLMs within 2 AICc points of the best model, we calculated the full model averaged coefficients ('MuMIn' package; Bartoń, 2022), and otherwise we selected the model with the lowest AICc as the best model. Because of the link function in our GLMs, our results can be interpreted as increasing or decreasing the log odds of recruitment or seedling survival, corresponding to lower or higher likelihoods, respectively.

#### Statistical analyses: microhabitat suitability

To answer how microhabitat suitability changes at and beyond species' range edges, we used the model results from the species establishment analyses above to predict likelihood of recruitment, relative recruit counts, and 1-year as well as 2-year seedling survival at each plot (predict(model, type = 'response')) ('Mu-MIn' package; Bartoń, 2022). We considered these predicted values as proxies for microhabitat suitability, with increasing suitability at and beyond range edge indicating greater likelihood of range expansion and decreasing suitability indicating a lower likelihood for range expansion. For these analyses, we used only the species where sites extend beyond their thermal range limit (Fig. S2) and with an observed response in at least 8 plots.

# Results

### Microhabitat variability

Microhabitat variation within sites was higher than among site variation for half of the microhabitat parameters for which we could fit nested LMMs. Out of these 9 microhabitat parameters that we measured, only 3 varied predictably with elevation (canopy openness, winter minimum soil temperature, and spring days of snow). The parameters that followed an elevational pattern are both expected to be directly (winter minimum temperature, spring days of snow) as well as indirectly (canopy openness) affected by climate (Fig. 2).

Our principal component analysis plot shows that the microhabitat variables we measured cluster broadly by soil characteristics (carbon:nitrogen, fungus:bacteria, water holding capacity), soil moisture (summer minimum soil moisture, spring days of snow), light availability (canopy openness), soil temperature (summer maximum soil temperature), and above-ground temperature (summer minimum plant-height temperature) (Fig. S3). An additional parameter describing soil temperature (winter minimum soil temperature) clusters in the middle of the microhabitat space.

#### Species establishment

Out of the 25 species that we sowed, 84% (21/25) recruited in at least 1 plot and 56% (14/25) recruited in 8 plots or more (Table S2). Of the species that recruited, seedlings (i.e. at least one seedling) survived for one year for 57% (8/14) of species and survived for two years for 43% (6/14) of species. Almost all species that had sites beyond their range edge recruited and survived beyond their range edge, a pattern seen for species with either high- or low-elevation range edges (Table S2). Almost all of our models met the GLM assumption of independence of residuals. However, many of our models did not meet homogeneity of variance, variance did not equal the mean, or the link function was sometimes inappropriate (> 0.5 difference from 1 for slope of link function). Together with the low proportion of variance (Table S3) from each set of candidate models, we are therefore cautious in interpreting our results.

Overall, we found microhabitat parameters both related and not related to climate identified as the most important parameters in model selection of the effects of microhabitat on establishment, but models had low explanatory power (Tables 1, S3). At the community level, the main patterns we found were that certain microhabitat variables had largely negative or positive effects, but the direction of these effects changed with different life stages (Fig. 3). At the species level, we found that some species only had few microhabitat parameters chosen in model selection, whereas others had many (Fig. S4). Within the same species, parameters usually had the same directionality of effect on likelihood of recruitment, recruit counts, and seedling survival except for T. grandiflora. For two species, A. lasiocarpa and E. lanatum, the directionality of effects was consistent across life stages.

### Microhabitat suitability

We generated predictions of microhabitat suitability to better understand if the distribution of microhabitats along climatic gradients suggests range shifts will likely be facilitated (i.e. where suitability increases with elevation), constrained (i.e. decreases), or unaffected (i.e. is constant). We note that due to poor model fit in the original models, our predictions cannot be used to make definitive forecasts of which species are likely to shift their ranges with climate change, but rather use these predictions to assess how microhabitat suitability changes across a large elevational gradient. Our predictions indicate that only range shifts for L. *latifolius* are likely to be facilitated with increasing microhabitat suitability with elevation (Fig. 4), although our experimental sites extended only just beyond the range limit for the species. For all other species, microhabitat suitability either declines or has no pattern with elevation and thus range shifts are likely to either be constrained or unaffected, respectively (Figs. S5, S6, S7). We also found that suitability patterns were modified by transect and life stage.

Together with these suitability predictions, the microhabitat parameters that significantly vary with elevation can give further insight on what aspects of microhabitat can facilitate or constrain range shifts. For example, the increasing microhabitat suitability for *L. latifolius* (Fig. 4a) together with the positive effects of canopy openness on the species (Table 1) and canopy openness increasing with elevation (Fig. 2a) point to a likely facilitated range expansion for the species. Spring snow cover shows the same pattern, and thus is a further microhabitat variable that will likely facilitate range expansion for this species. However, winter minimum soil temperature decreases with elevation in the Okanagan transect but positively affects *L. latifolius* recruit counts and seedling survival, so this microhabitat variable could constrain range expansion.

### Discussion

While many species ranges are on average shifting in the direction predicted by SDMs, there is large variation among species that is difficult to explain. Microhabitat variability is increasingly recognized as influencing population dynamics across species ranges (Oldfather & Ackerly, 2019) and may be a key factor in understanding and predicting range shifts (Lembrechts, Nijs, et al., 2019; Maclean & Early, 2023; Stickley & Fraterrigo, 2023). However, how microhabitat suitability, and variability therein, is distributed along species ranges is vastly understudied, impeding an understanding of how microhabitat can facilitate or constrain range shifts. We found that microhabitat is highly variable across species ranges and is largely decoupled from the macro-scale at which SDMs are typically constructed. We further found variable, albeit weak, effects of microhabitat for different species and life stages, suggesting that the drivers of establishment are complex and difficult to detect. Our microhabitat suitability predictions show microhabitat will either constrain range expansions, or have no effect, for most species, with the range shifts of just one species (*L. latifolius*) likely facilitated by microhabitat.

#### Complex microhabitat patterns across elevation gradients

Surprisingly, most of our microhabitat variables, even ones related to climate (summer soil moisture, summer soil temperature, summer plant height temperature) did not follow an elevational pattern representative of the macroclimate (Fig. S8) commonly used in SDMs. This is consistent with many studies finding that microhabitat is often decoupled from elevation and regional climate (Ford et al., 2013; Lembrechts, Lenoir, et al., 2019). We further found that none of the soil composition parameters we measured (fungus:bacteria, carbon:nitrogen, water holding capacity) followed an elevational pattern, even though other alpine studies have found soil microbial community (Hiiesalu et al., 2023) and soil carbon:nitrogen (Weintraub et al., 2016) to covary with elevation.

We further found that only canopy openness, winter minimum soil temperature, and spring snow days covary with elevation. Despite higher elevation areas characterized by more open forest canopies, the high variability at each site (Fig. 2) is in line with the large body of work showing that forest canopies can provide climate refugia (De Frenne et al., 2019; Haesen et al., 2021). As species light requirements can be variable, light availability and canopy cover can shape species ranges (Muñoz Mazon et al., 2023) and therefore can be an important component of range shifts (Tourville et al., 2022).

#### Low and variable effects of microhabitat on establishment

Our results agree with other studies, which find overall low recruitment in seed addition sites (Clark et al., 2007) and decreasing fitness of transplants beyond the range (Lee-Yaw et al., 2016; Stanton-Geddes et al., 2012). Plant establishment beyond current range edges is necessary for plant species to shift their ranges upward, however this is complicated by plants usually being recruitment limited. This means that they are

limited by some aspect of their environment (e.g., microhabitat unsuitability, seed predation) and not by dispersal (Clark et al., 2007). While we did not control for seed predation, herbivory, or fungal infections, we captured mortality with our methods. Even though seed predation rates decline in higher elevation areas (Hargreaves et al., 2019), we still removed fleshy fruits around seeds to deter predation. We also did not collect data to test for effects of community composition on establishment, but with low recruitment rates, plants in our system would still be considered recruitment limited.

We did not find that establishment responses to any particular microhabitat were restricted to certain growth forms, with species-specific relationships to microhabitat. This is not surprising, as species have unique environmental requirements (Table S1) and species characteristics even impact predictive power in SDMs (Guisan et al., 2007). We also did not find differences in effects of the microhabitat aspects that are expected to be directly versus indirectly affected by climate change. Interestingly, we found that some parameters, such as winter soil temperature, increased likelihood of recruitment but these effects switched to decreasing likelihood of seedling survival. One exception was spring days with snow, which generally had positive effects, and this is similar to findings by Davis & Gedalof (2018), who found positive effects of winter snow cover on recruitment. The importance of microbial community in plant dynamics is also increasingly acknowledged (Castro et al., 2022), although soil fungus:bacteria ratio was not selected more than other variables in our system. Microbial communities could even be an important factor in determining species range shifts, such as favorable soil microbial composition mediating climate tolerance to promote tree seedling survival (Allsup et al., 2023).

#### Predicting microhabitat suitability

We found that microhabitat suitability beyond the leading range edge was species- and life stage-specific, with most species showing either decreased suitability or no pattern with elevation. We posit that the role that microhabitat may play in facilitating or constraining range shifts for any given species is closely tied to how microhabitat itself varies with elevation. For example, if a favorable microhabitat parameter for establishment decreases with elevation, establishment will overall not be favored and a range shift could be constrained. We found this in our system, with increasing likelihood of L. latifolius recruit counts and seedling survival with warmer winter minimum soil temperatures, but soil temperature decreases with elevation. If a favorable microhabitat parameter increases with elevation, however, this could lead to a facilitated range shift. This is the case for canopy openness, which increases with elevation and also increases likelihood of L. latifolius establishment. Since microhabitat suitability increases beyond the range for this species, these cases highlight the complex ways in which microhabitat parameters may interact to mediate range shifts. The alternative scenarios, where an unfavorable microhabitat parameter either declines with elevation to inversely favor establishment or increases with elevation to inhibit establishment, can also facilitate or inhibit range shifts, respectively.

Macroclimatic parameters vary predictably with elevation (Fig. S8) in our study system, however only a third of our microhabitat variables show an elevational pattern. Since species distributions can be driven by microhabitat, understanding how microhabitat parameters vary across species' ranges can give insights into how microhabitat may facilitate or inhibit range expansion (Lembrechts et al., 2017; Tourville et al., 2022; Kemppinen et al., 2023). Incorporating microhabitat suitability into ecological studies is not new, including quantifying habitat suitability in the field (Jabis & Ayers, 2014), with remote sensing (Falco et al., 2019), and at the leading range edge (Mamet & Kershaw, 2013). Transplant studies often find decreasing macroclimate suitability beyond the range (Lee-Yaw et al., 2016), but to our knowledge no studies exist that assess microhabitats in SDMs (Lembrechts, Nijs, et al., 2019), the vast majority of SDMs still focus on macroclimatic variables and this may lead to the mismatches between predicted and observed range shifts, or lack thereof.

In our system, we found that suitable microhabitat is unchanged or reduced beyond the leading edge for many species, yet we also find that macroclimatic conditions enable an upwards shift in species' recruitment optima at the community level (*unpublished data*). However, this pattern is only evident at the community level, with high species-level variability and no effect of canopy cover. This matches the large variability, and low explanatory power, seen in our microhabitat suitability predictions, and suggests that species can find pockets of suitable microhabitat to recruit beyond their macroclimatic cold range edge. Microhabitat suitability may be just high enough to allow for successful establishment, but is overall very low and the complicated ways in which microhabitat acts to affect species establishment makes it difficult to detect patterns. We also found continued species recruitment at the warm edge of the range (*unpublished data*) and this buffering of contractions at the warm edge could be due to microhabitat refugia (De Lombaerde et al., 2022) created by the range of microhabitat found at the lower elevation sites (Fig. 2)

#### Limitations

Our experimental design is not without limitations. Seed provenance might cause different recruitment responses across microhabitat gradients, however variation in germination rates was not biologically meaningful (Fig. S1). Since we sowed all seeds in the same year, we also cannot test if we sowed in favorable versus unfavorable years and aimed to capture a large germination window by recording recruitment over three years. Finally, we collected some of our data on species responses and microhabitat measurements asynchronously and therefore emphasize that only differences between sites, not absolute values, should be interpreted for their effects. Finally, we strongly urge future studies to incorporate increased replication at each microhabitat value for higher statistical power in explaining results.

#### Conclusion

To our knowledge, our work is one of the first studies to measure key demographic life stages of a large group of species along a microhabitat gradient both within and beyond their current range limit. As such, this work yields a more comprehensive understanding of the mechanisms that set species range limits and indicates ways in which species shift their distributions with climate change. Our results suggest that complex ways in which microhabitat parameters influence early life stages are complex and difficult to detect, which complicates range shift predictions. A greater understanding of the role of macrohabitat in shaping species' distribution limits will ultimately improve predictions of how species distributions will shift with climate change. We emphasize that predictions accounting for complex microhabitat drivers are necessary to create tailored conservation and management decisions in order to mitigate ongoing biodiversity loss.

## Acknowledgements

#### [REMOVED FOR PEER REVIEW]

#### Data archiving statement

All data used in this study are available on Dryad [DATA DEPOSITED AT TIME OF PUBLICATION]. All R scripts are available at [LINK REMOVED FOR PEER REVIEW].

# **Conflict of Interest Statement**

The authors declare no conflict of interest.

# **Ethics Statement**

Approval for field work was granted by the Mount Baker and Okanagan National Forests. No special use permits were needed for this work.

# **Funding Statement**

[REMOVED FOR PEER REVIEW]

# Author Contributions

[REMOVED FOR PEER REVIEW]

### References

Allsup, C. M., George, I., & Lankau, R. A. (2023). Shifting microbial communities can enhance tree tolerance to changing climates. *Science*, 380 (6647), 835–840. https://doi.org/10.1126/science.adf2027

Bartoń, K. (2022). *MuMIn: Multi-Model Inference* (1.47.1) [Computer software]. https://cran.r-project.org/web/packages/MuMIn/index.html

Bateman, A., Erickson, T. E., Merritt, D. J., & Muñoz-Rojas, M. (2017). Effects of inorganic amendments (urea, gypsum) on seed germination and seedling recruitment of 20 native plant species used in dryland restoration. *Geophysical Research Abstracts*, 19 (EGU2017-15744). https://meetingorganizer.copernicus.org/EGU2017/EGU2017-15744.pdf

Bullied, W. J., Van Acker, R. C., & Bullock, P. R. (2012). Review: Microsite characteristics influencing weed seedling recruitment and implications for recruitment modeling. *Canadian Journal of Plant Science*, 92 (4), 627–650. https://doi.org/10.4141/cjps2011-281

Caldeira, M. C., Ibáñez, I., Nogueira, C., Bugalho, M. N., Lecomte, X., Moreira, A., & Pereira, J. S. (2014). Direct and indirect effects of tree canopy facilitation in the recruitment of M editerranean oaks. *Journal of Applied Ecology*, 51 (2), 349–358.

Castro, D., Concha, C., Jamett, F., Ibáñez, C., & Hurry, V. (2022). Soil Microbiome Influences on Seedling Establishment and Growth of Prosopis chilensis and Prosopis tamarugo from Northern Chile. *Plants*, 11 (20), 2717. https://doi.org/10.3390/plants11202717

Clark, C. J., Poulsen, J. R., Levey, D. J., & Osenberg, C. W. (2007). Are Plant Populations Seed Limited? A Critique and Meta-Analysis of Seed Addition Experiments. *The American Naturalist*, 170 (1), 128–142. https://doi.org/10.1086/518565

Copenhaver-Parry, P. E., Carroll, C. J. W., Martin, P. H., & Talluto, M. V. (2020). Multi-scale integration of tree recruitment and range dynamics in a changing climate. *Global Ecology and Biogeography*, 29 (1), 102–116. https://doi.org/10.1111/geb.13012

Davis, E. L., & Gedalof, Z. (2018). Limited prospects for future alpine treeline advance in the Canadian Rocky Mountains. *Global Change Biology*, 24 (10), 4489–4504. https://doi.org/10.1111/gcb.14338

De Frenne, P., Zellweger, F., Rodriguez-Sanchez, F., Scheffers, B. R., Hylander, K., Luoto, M., Vellend, M., Verheyen, K., & Lenoir, J. (2019). Global buffering of temperatures under forest canopies. *Nature Ecology & Evolution*, 3 (5), Article 5. https://doi.org/10.1038/s41559-019-0842-1

De Lombaerde, E., Vangansbeke, P., Lenoir, J., Van Meerbeek, K., Lembrechts, J., Rodriguez-Sanchez, F., Luoto, M., Scheffers, B., Haesen, S., Aalto, J., Christiansen, D. M., De Pauw, K., Depauw, L., Govaert, S., Greiser, C., Hampe, A., Hylander, K., Klinges, D., Koelemeijer, I., ... De Frenne, P. (2022). Maintaining forest cover to enhance temperature buffering under future climate change. *Science of The Total Environment*, 810, 151338. https://doi.org/10.1016/j.scitotenv.2021.151338

Dobrowski, S. Z. (2011). A climatic basis for microrefugia: The influence of terrain on climate . 1022–1035. https://doi.org/10.1111/j.1365-2486.2010.02263.x

Dolezal, J., Jandova, V., Macek, M., Mudrak, O., Altman, J., Schweingruber, F. H., & Liancourt, P. (2021). Climate warming drives Himalayan alpine plant growth and recruitment dynamics. *Journal of Ecology*, 109 (1), 179–190. https://doi.org/10.1111/1365-2745.13459

Elith, J., Kearney, M., & Phillips, S. (2010). The art of modelling range-shifting species. *Methods in Ecology* and Evolution ,1, 330–342. https://doi.org/10.1111/j.2041-210X.2010.00036.x

Elliott, G. P., & Petruccelli, C. A. (2018). Tree recruitment at the treeline across the Continental Divide in the Northern Rocky Mountains, USA: The role of spring snow and autumn climate. *Plant Ecology & Diversity*, 11 (3), 319–333. https://doi.org/10.1080/17550874.2018.1487475

Falco, N., Wainwright, H., Dafflon, B., Leger, E., Peterson, J., Steltzer, H., Wilmer, C., Rowland, J. C., Williams, K. H., & Hubbard, S. S. (2019). Investigating Microtopographic and Soil Controls on a Mountainous Meadow Plant Community Using High-Resolution Remote Sensing and Surface Geophysical Data. *Jour*nal of Geophysical Research: Biogeosciences, 124 (6), 1618–1636. https://doi.org/10.1029/2018JG004394

Ford, K. R., Ettinger, A. K., Lundquist, J. D., Raleigh, M. S., & Lambers, J. H. R. (2013). Spatial Heterogeneity in Ecologically Important Climate Variables at Coarse and Fine Scales in a High-Snow Mountain Landscape. *PLOS ONE*, 8 (6), e65008. https://doi.org/10.1371/journal.pone.0065008

Fox, J., Weisberg, S., Price, B., Adler, D., Bates, D., Baud-Bovy, G., Bolker, B., Ellison, S., Firth, D., Friendly, M., Gorjanc, G., Graves, S., Heiberger, R., Krivitsky, P., Laboissiere, R., Maechler, M., Monette, G., Murdoch, D., Nilsson, H., ... R-Core. (2023).car: Companion to Applied Regression (3.1-2) [Computer software]. https://cran.r-project.org/web/packages/car/index.html

Franklin, J. (2023). Species distribution modelling supports the study of past, present and future biogeography phies. Journal of Biogeography, n/a (n/a). https://doi.org/10.1111/jbi.14617

Freeman, B. G., Lee-Yaw, J. A., Sunday, J. M., & Hargreaves, A. L. (2018). Expanding, shifting and shrinking: The impact of global warming on species' elevational distributions. *Global Ecology and Biogeography*, *May*, 1–9. https://doi.org/10.1111/geb.12774

Goodwin, K. J. A., & Brown, C. D. (2023). Integrating demographic niches and black spruce range expansion at subarctic treelines. *Oecologia*, 201 (1), 19–29. https://doi.org/10.1007/s00442-022-05293-7

Graae, B. J., Ejrnaes, R., Marchand, F. L., Milbau, A., Shevtsova, A., Beyens, L., & Nijs, I. (2009). The effect of an early-season short-term heat pulse on plant recruitment in the Arctic. *Polar Biology*, 32 (8), 1117–1126. https://doi.org/10.1007/s00300-009-0608-3

Greiser, C., Hylander, K., Meineri, E., Luoto, M., & Ehrlen, J. (2020). Climate limitation at the cold edge: Contrasting perspectives from species distribution modelling and a transplant experiment. *Ecography*, 43, 1–11. https://doi.org/10.1111/ecog.04490

Guisan, A., Zimmermann, N. E., Elith, J., Graham, C. H., Phillips, S., & Peterson, A. T. (2007). What Matters for Predicting the Occurrences of Trees: Techniques, Data, or Species' Characteristics? *Ecological Monographs*, 77 (4), 615–630.

Haesen, S., Lembrechts, J. J., De Frenne, P., Lenoir, J., Aalto, J., Ashcroft, M. B., Kopecky, M., Luoto, M., Maclean, I., Nijs, I., Niittynen, P., van den Hoogen, J., Arriga, N., Brůna, J., Buchmann, N., Čiliak, M., Collalti, A., De Lombaerde, E., Descombes, P., ... Van Meerbeek, K. (2021). ForestTemp – Sub-canopy microclimate temperatures of European forests. *Global Change Biology*, 27 (23), 6307–6319. https://doi.org/10.1111/gcb.15892

Hargreaves, A. L., Suárez, E., Mehltreter, K., Myers-Smith, I., Vanderplank, S. E., Slinn, H. L., Vargas-Rodriguez, Y. L., Haeussler, S., David, S., Muñoz, J., Almazán-Núñez, R. C., Loughnan, D., Benning, J. W., Moeller, D. A., Brodie, J. F., Thomas, H. J. D., & Morales, P. A. (2019). Seed predation increases from the Arctic to the Equator and from high to low elevations. *Science Advances*, 5 (2), 1–11. htt-ps://doi.org/10.1126/sciadv.aau4403

Hiiesalu, I., Schweichhart, J., Angel, R., Davison, J., Doležal, J., Kopecký, M., Macek, M., & Řehakova, K. (2023). Plant-symbiotic fungal diversity tracks variation in vegetation and the abiotic environment along an extended elevational gradient in the Himalayas. *FEMS Microbiology Ecology*, fiad092. https://doi.org/10.1093/femsec/fiad092

Jabis, M. D., & Ayers, T. J. (2014). Habitat Suitability as a Limiting Factor for Establishment in a Narrow Endemic: Abronia alpina (Nyctaginaceae). Western North American Naturalist, 74 (2), 185–200. https://doi.org/10.3398/064.074.0204

James, J. J., Sheley, R. L., Leger, E. A., Adler, P. B., Hardegree, S. P., Gornish, E. S., & Rinella, M. J. (2019). Increased soil temperature and decreased precipitation during early life stages constrain grass seedling recruitment in cold desert restoration. *Journal of Applied Ecology*, 56 (12), 2609–2619. https://doi.org/10.1111/1365-2664.13508

Käber, Y., Meyer, P., Stillhard, J., De Lombaerde, E., Zell, J., Stadelmann, G., Bugmann, H., & Bigler, C. (2021). Tree recruitment is determined by stand structure and shade tolerance with uncertain role of climate and water relations. *Ecology and Evolution*, 11 (17), 12182–12203. https://doi.org/10.1002/ece3.7984

Kemppinen, J., Lembrechts, J. J., Van Meerbeek, K., Carnicer, J., Chardon, N. I., Kardol, P., Lenoir, J., Liu, D., Maclean, I., Pergl, J., Saccone, P., Senior, R. A., Shen, T., Słowińska, S., Vandvik, V., von Oppen, J., Aalto, J., Ayalew, B., Bates, O., ... De Frenne, P. (2023). *Microclimate, an inseparable part of ecology* and biogeography. Zenodo. https://doi.org/10.5281/zenodo.7973314

Kephart, S. R., & Paladino, C. (1997). Demographic change and microhabitat Variability in a Grassland Endemic, Silene Douglasii Var. Oraria (Caryophyllaceae). *American Journal of Botany*, 84 (2), 179–189. https://doi.org/10.2307/2446079

Khurana, E., & Singh, J. S. (2000). Influence of Seed Size on Seedling Growth of Albizia procera Under Different Soil Water Levels. *Annals of Botany*, 86 (6), 1185–1192. https://doi.org/10.1006/anbo.2000.1288

Kołodziejek, J., Patykowski, J., & Wala, M. (2017). Effect of light, gibberellic acid and nitrogen source on germination of eight taxa from dissapearing European temperate forest, Potentillo albae-Quercetum. *Scientific Reports*, 7 (1), 13924. https://doi.org/10.1038/s41598-017-13101-z

Kroiss, S. J., & HilleRisLambers, J. (2015). Recruitment limitation of long-lived conifers: Implications for climate change responses. *Ecology*, 96 (5), 1286–1297. https://doi.org/10.1890/14-0595.1

Kueppers, L. M., Conlisk, E., Castanha, C., Moyes, A. B., Germino, M. J., de Valpine, P., Torn, M. S., & Mitton, J. B. (2017). Warming and provenance limit tree recruitment across and beyond the elevation range of subalpine forest. *Global Change Biology*, 23 (6), 2383–2395. https://doi.org/10.1111/gcb.13561

Lawson, C. R., Bennie, J., Hodgson, J. A., Thomas, C. D., & Wilson, R. J. (2014). Topographic microclimates drive microhabitat associations at the range margin of a butterfly. *Ecography*, 37 (8), 732–740. https://doi.org/10.1111/ecog.00535

Lee-Yaw, J. A., Kharouba, H. M., Bontrager, M., Mahony, C., Csergo, A. M., Noreen, A. M. E., Li, Q., Schuster, R., & Angert, A. L. (2016). A synthesis of transplant experiments and ecological niche models suggests that range limits are often niche limits. *Ecology Letters*, 19 (6), 710–722. https://doi.org/10.1111/ele.12604

Lembrechts, J. J. (2023). Microclimate alters the picture. Nature Climate Change, 13 (5), Article 5. https://doi.org/10.1038/s41558-023-01632-5

Lembrechts, J. J., Lenoir, J., Nuñez, M. A., Pauchard, A., Geron, C., Bussé, G., Milbau, A., & Nijs, I. (2017). Microclimate variability in alpine ecosystems as stepping stones for non-native plant establishment above their current elevational limit. *Ecography*, 41 (6), 900–909. https://doi.org/10.1111/ecog.03263

Lembrechts, J. J., Lenoir, J., Roth, N., Hattab, T., Milbau, A., Haider, S., Pellissier, L., Pauchard, A., Ratier Backes, A., Dimarco, R. D., Nuñez, M. A., Aalto, J., & Nijs, I. (2019). Comparing temperature data sources for use in species distribution models: From in-situ logging to remote sensing. *Global Ecology and Biogeography*, 28 (11), 1578–1596. https://doi.org/10.1111/geb.12974

Lembrechts, J. J., Nijs, I., & Lenoir, J. (2019). Incorporating microclimate into species distribution models. *Ecography*, 42 (7), 1267–1279. https://doi.org/10.1111/ecog.03947

Li, Y., Yang, H., Xia, J., Zhang, W., Wan, S., & Li, L. (2011). Effects of increased nitrogen deposition and precipitation on seed and seedling production of potentilla tanacetifolia in a temperate steppe ecosystem. *PLoS ONE*, 6 (12). https://doi.org/10.1371/journal.pone.0028601

Lloret, F., Peñuelas, J., & Estiarte, M. (2005). Effects of vegetation canopy and climate on seedling establishment in Mediterranean shrubland. *Journal of Vegetation Science*, 16 (1), 67–76. https://doi.org/10.1111/j.1654-1103.2005.tb02339.x

Lovejoy, T. E., & Hannah, L. (2019). *Biodiversity and Climate Change: Transforming the Biosphere*. Yale University Press.

Maclean, I. M. D., & Early, R. (2023). Macroclimate data overestimate range shifts of plants in response to climate change. *Nature Climate Change*, 13 (5), 484–490. https://doi.org/10.1038/s41558-023-01650-3

Mamet, S. D., & Kershaw, G. P. (2013). Multi-scale Analysis of Environmental Conditions and Conifer Seedling Distribution Across the Treeline Ecotone of Northern Manitoba, Canada. *Ecosystems*, 16 (2), 295–309. https://doi.org/10.1007/s10021-012-9614-3

Man, M., Kalčík, V., Macek, M., Brůna, J., Hederová, L., Wild, J., & Kopecký, M. (2023). MyClim: Microclimate data handling and standardised analyses in R - Man—Methods in Ecology and Evolution—Wiley Online Library. *Methods in Ecology and Evolution*, *Early View*. https://doi.org/10.1111/2041-210X.14192

Man, M., Kalčík, V., Macek, M., Wild, J., Kopecký, M., Brůna, J., Hederová, L., & Sciences, I. of B. of the C. A. of. (2023).myClim: Microclimatic Data Processing (1.0.1) [Computer software]. https://cran.r-project.org/web/packages/myClim/index.html

Mazerolle, M. J. (2020). AICcmodavg: Model Selection and Multimodel Inference Based on (Q)AIC(c) (2.3-1) [Computer software]. https://cran.r-project.org/web/packages/AICcmodavg/index.html

Monaco, T. A., Mackown, C. T., Johnson, D. A., Jones, T. A., Norton, J. M., Norton, J. B., & Redinbaugh, M. G. (2003). Nitrogen effects on seed germination and seedling growth. In *Plant Physiologist and Plant Geneticist* (Vol. 56, pp. 73036–2144). Plants, Soils.

Mondoni, A., Pedrini, S., Bernareggi, G., Rossi, G., Abeli, T., Probert, R. J., Ghitti, M., Bonomi, C., & Orsenigo, S. (2015). Climate warming could increase recruitment success in glacier foreland plants. *Annals of Botany*, 116 (6), 907–916. https://doi.org/10.1093/aob/mcv101

Morelli, T. L., Barrows, C. W., Ramirez, A. R., Cartwright, J. M., Ackerly, D. D., Eaves, T. D., Ebersole, J. L., Krawchuk, M. A., Letcher, B. H., Mahalovich, M. F., Meigs, G. W., Michalak, J. L., Millar, C. I., Quiñones, R. M., Stralberg, D., & Thorne, J. H. (2020). Climate-change refugia: Biodiversity in the slow lane. *Frontiers in Ecology and the Environment*, 18 (5), 228–234. https://doi.org/10.1002/fee.2189

Morris, W. F., & Ehrlén, J. (2015). Predicting changes in the distribution and abundance of species under environmental change. *Ecology Letters*, 18 (3), 303–314. https://doi.org/10.1111/ele.12410

Moser, B., Walthert, L., Metslaid, M., Wasem, U., & Wohlgemuth, T. (2017). Spring water deficit and soil conditions matter more than seed origin and summer drought for the establishment of temperate conifers. *Oecologia*, 183 (2), 519–530. https://doi.org/10.1007/s00442-016-3766-3

Muñoz Mazon, M., Klanderud, K., & Sheil, D. (2023). Exploring how disturbance and light availability shape the elevation ranges of multiple mountain tree and shrub species in the tropics. Landscape Ecology . https://doi.org/10.1007/s10980-023-01670-6

Normand, S., Zimmermann, N. E., Schurr, F. M., & Lischke, H. (2014). Demography as the basis for understanding and predicting range dynamics. *Ecography*, 37, 1149–1154. https://doi.org/10.1111/ecog.01490

Oldfather, M. F., & Ackerly, D. D. (2019). Microclimate and demography interact to shape stable population dynamics across the range of an alpine plant. *New Phytologist*, 222 (1), 193–205. https://doi.org/10.1111/nph.15565

Ósvaldsson, A., Chesler, M. K., & Burns, J. H. (2022). Effects of snow on reproduction of perennial Thalictrum dioicum: Plants survive but seedlings fail to recruit with reduced snow cover. *American Journal of Botany*, 109 (3), 406–418. https://doi.org/10.1002/ajb2.1829

Pacifici, M., Foden, W. B., Visconti, P., Watson, J. E. M., Butchart, S. H. M., Kovacs, K. M., Scheffers, B. R., Hole, D. G., Martin, T. G., Akçakaya, H. R., Corlett, R. T., Huntley, B., Bickford, D., Carr, J. A., Hoffmann, A. A., Midgley, G. F., Pearce-Kelly, P., Pearson, R. G., Williams, S. E., ... Rondinini, C. (2015). Assessing species vulnerability to climate change. *Nature Climate Change*, 5, 215–225. https://doi.org/10.1038/nclimate2448

Pérez-Fernández, M. A., Calvo-Magro, E., Montanero-Fernández, J., & Oyola-Velasco, J. A. (2006). Seed germination in response to chemicals: Effect of nitrogen and pH in the media. *Journal of Environmental Biology*, 27 (1), 13–20.

Pradhan, K., Ettinger, A. K., Case, M. J., & Hille Ris Lambers, J. (2023). Applying climate change refugia to forest management and old-growth restoration. *Global Change Biology*, 29 (13), 3692–3706. https://doi.org/10.1111/gcb.16714

Proll, G., Dullinger, S., Dirnbock, T., Kaiser, C., & Richter, A. (2011). Effects of nitrogen on tree recruitment in a temperate montane forest as analysed by measured variables and Ellenberg indicator values. *Preslia*, 83, 111–127.

Pulliam, H. R. (2000). On the relationship between niche and distribution. Ecology Letters , 3 , 349–361. https://doi.org/10.1046/j.1461-0248.2000.00143.x

R Core Team, R. (2023). R: A Language and Environment for Statistical Computing [Computer software]. R Foundation for Statistical Computing, Vienna, Austria. http://www.r-project.org

Renard, S. M., McIntire, E. J. B., & Fajardo, A. (2016). Winter conditions – not summer temperature – influence establishment of seedlings at white spruce alpine treeline in Eastern Quebec. *Journal of Vegetation Science*, 27 (1), 29–39. https://doi.org/10.1111/jvs.12347

Rigg, J. L., Offord, C. A., Singh, B. K., Anderson, I., Clarke, S., & Powell, J. R. (2016). Soil microbial communities influence seedling growth of a rare conifer independent of plant–soil feedback. *Ecology*, 97 (12), 3346–3358. https://doi.org/10.1002/ecy.1594

Rodriguez-Garcia, E., Bravo, F., & Spies, T. A. (2011). Effects of overstorey canopy, plant–plant interactions and soil properties on Mediterranean maritime pine seedling dynamics. *Forest Ecology and Management*, 262 (2), 244–251.

Rumpf, S. B., Hulber, K., Klonner, G., Moser, D., Schutz, M., Wessely, J., Willner, W., Zimmermann, N. E., & Dullinger, S. (2018). Range dynamics of mountain plants decrease with elevation. *Proceedings of the National Academy of Sciences*, 201713936–201713936. https://doi.org/10.1073/PNAS.1713936115

Sanczuk, P., De Lombaerde, E., Haesen, S., Van Meerbeek, K., Van der Veken, B., Hermy, M., Verheyen, K., Vangansbeke, P., & De Frenne, P. (2022). Species distribution models and a 60-year-old transplant experiment reveal inhibited forest plant range shifts under climate change. *Journal of Biogeography*, 49 (3), 537–550. https://doi.org/10.1111/jbi.14325

Sanczuk, P., De Pauw, K., De Lombaerde, E., Luoto, M., Meeussen, C., Govaert, S., Vanneste, T., Depauw, L., Brunet, J., Cousins, S. A. O., Gasperini, C., Hedwall, P.-O., Iacopetti, G., Lenoir, J., Plue, J., Selvi, F., Spicher, F., Uria-Diez, J., Verheyen, K., ... De Frenne, P. (2023). Microclimate and forest density drive plant population dynamics under climate change. *Nature Climate Change*, 13 (8), Article 8. https://doi.org/10.1038/s41558-023-01744-y

Santana, V. M., Bradstock, R. A., Ooi, M. K. J., Denham, A. J., Auld, T. D., Baeza, M. J., Santana, V. M., Bradstock, R. A., Ooi, M. K. J., Denham, A. J., Auld, T. D., & Baeza, M. J. (2010). Effects of soil temperature regimes after fire on seed dormancy and germination in six Australian Fabaceae species. *Australian Journal of Botany*, 58 (7), 539–545. https://doi.org/10.1071/BT10144

Scherrer, D., & Korner, C. (2010). Infra-red thermometry of alpine landscapes challenges climatic warming projections. *Global Change Biology*, 16 (9), 2602–2613. https://doi.org/10.1111/j.1365-2486.2009.02122.x

Schurr, F. M., Pagel, J., Cabral, J. S., Groeneveld, J., Bykova, O., O'Hara, R. B., Hartig, F., Kissling, W. D., Linder, H. P., Midgley, G. F., Schroder, B., Singer, A., & Zimmermann, N. E. (2012). How to understand species' niches and range dynamics: A demographic research agenda for biogeography. *Journal of Biogeography*, 39, 2146–2162. https://doi.org/10.1111/j.1365-2699.2012.02737.x

Selmants, P. C., Adair, K. L., Litton, C. M., Giardina, C. P., & Schwartz, E. (2016). Increases in mean annual temperature do not alter soil bacterial community structure in tropical montane wet forests. *Ecosphere*, 7 (4). https://doi.org/10.1002/ecs2.1296

Smithers, B. (2017). Soil Preferences in Germination and Survival of Limber Pine in the Great Basin White Mountains. *Forests*, 8 (11), 423. https://doi.org/10.3390/f8110423

Stanton-Geddes, J., Tiffin, P., & Shaw, R. G. (2012). Role of climate and competitors in limiting fitness across range edges of an annual plant. *Ecology*, 93 (7), 1604–1613. https://doi.org/10.1890/11-1701.1

Stickley, S. F., & Fraterrigo, J. M. (2023). Microclimate species distribution models estimate lower levels of climate-related habitat loss for salamanders. *Journal for Nature Conservation*, 72, 126333. https://doi.org/10.1016/j.jnc.2023.126333

Szeicz, J. M., & Macdonald, G. M. (1995). Recent White Spruce Dynamics at the Subarctic Alpine Treeline of North-Western Canada. *Journal of Ecology*, 83 (5), 873–885. https://doi.org/10.2307/2261424

Tanner, K. E., Moore-O'Leary, K. A., Parker, I. M., Pavlik, B. M., Haji, S., & Hernandez, R. R. (2021). Microhabitats associated with solar energy development alter demography of two desert annuals. *Ecological Applications*, 31 (6), e02349. https://doi.org/10.1002/eap.2349

Thompson, L. J., & Naeem, S. (1996). The effects of soil warming on plant recruitment. *Plant and Soil*, 182 (2), 339–343. https://doi.org/10.1007/BF00029064

Thuiller, W., Albert, C., Araujo, M. B., Berry, P. M., Cabeza, M., Guisan, A., Hickler, T., Midgley, G. F., Paterson, J., Schurr, F. M., Sykes, M. T., & Zimmermann, N. E. (2008). Predicting global change impacts on plant species' distributions: Future challenges. *Perspectives in Plant Ecology, Evolution and Systematics*, 9, 137–152. https://doi.org/10.1016/j.ppees.2007.09.004

Tobias, T. B., Farrer, E. C., Rosales, A., Sinsabaugh, R. L., Suding, K. N., & Porras-Alfaro, A. (2017). Seed-associated fungi in the alpine tundra: Both mutualists and pathogens could impact plant recruitment. *Fungal Ecology*, 30, 10–18. https://doi.org/10.1016/j.funeco.2017.08.001

Tourville, J. C., Wason, J. W., & Dovciak, M. (2022). Canopy gaps facilitate upslope shifts in montane conifers but not in temperate deciduous trees in the Northeastern United States. *Journal of Ecology*, 110 (12), 2870–2882. https://doi.org/10.1111/1365-2745.13993

Tredennick, A. T., Hooker, G., Ellner, S. P., & Adler, P. B. (2021). A practical guide to selecting models for exploration, inference, and prediction in ecology. *Ecology*, 102 (6), e03336. https://doi.org/10.1002/ecy.3336

Urban, M. C. (2019). Projecting biological impacts from climate change like a climate scientist. Wiley Interdisciplinary Reviews: Climate Change, 10, e585–e585. https://doi.org/10.1002/wcc.585

van der Heijden, M. G., Bruin, S. de, Luckerhoff, L., van Logtestijn, R. S., & Schlaeppi, K. (2016). A widespread plant-fungal-bacterial symbiosis promotes plant biodiversity, plant nutrition and seedling recruitment. *The ISME Journal*, 10 (2), Article 2. https://doi.org/10.1038/ismej.2015.120

Weintraub, S. R., Cole, R. J., Schmitt, C. G., & All, J. D. (2016). Climatic controls on the isotopic composition and availability of soil nitrogen across mountainous tropical forest. *Ecosphere*, 7 (8). https://doi.org/10.1002/ecs2.1412

Wiens, J. A., Stralberg, D., Jongsomjit, D., Howell, C. A., & Snyder, M. A. (2009). Niches, models, and climate change: Assessing the assumptions and uncertainties. *Proceedings of the National Academy of Sciences*, 106 (Supplement 2), 19729–19736. https://doi.org/10.1073/pnas.0901639106

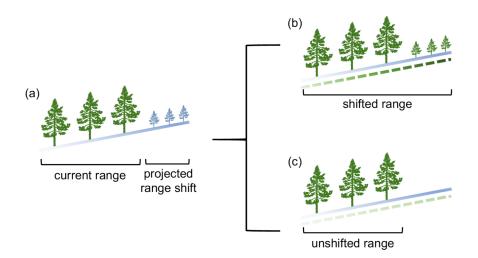
Wild, J., Kopecky, M., Macek, M., Šanda, M., Jankovec, J., & Haase, T. (2019). Climate at ecologically relevant scales: A new temperature and soil moisture logger for long-term microclimate measurement. *Agricultural and Forest Meteorology*, 268, 40–47. https://doi.org/10.1016/j.agrformet.2018.12.018

Xi, N., Chu, C., & Bloor, J. M. G. (2018). Plant drought resistance is mediated by soil microbial community structure and soil-plant feedbacks in a savanna tree species. *Environmental and Experimental Botany*, 155, 695–701. https://doi.org/10.1016/j.envexpbot.2018.08.013

Yates, C. J., Hobbs, R. J., & Bell, R. W. (1996). Factors Limiting the Recruitment of Eucalyptus salmonophloia in Remnant Woodlands. III. Conditions Necessary for Seed Germination. *Australian Journal of Botany*, 44 (3), 283–296. https://doi.org/10.1071/bt9960283

Zhong, M., Miao, Y., Han, S., & Wang, D. (2019). Nitrogen addition decreases seed germination in a temperate steppe. *Ecology and Evolution*, 9 (15), 8441–8449. https://doi.org/10.1002/ece3.5151

Zurbriggen, N., Hättenschwiler, S., Frei, E. S., Hagedorn, F., & Bebi, P. (2013). Performance of germinating tree seedlings below and above treeline in the Swiss Alps. *Plant Ecology*, 214 (3), 385–396. https://doi.org/10.1007/s11258-013-0176-z



**Figure 1.** Most projections of range shifts (a) only include macroclimate (blue line), which varies predictably with elevation, and thus range shifts are commonly predicted beyond current upper elevational range limits (blue seedlings). However, it is ultimately successful establishment (i.e. success in early life stages) in response to microhabitat (dashed green line), not macroclimate, beyond the current range limit that will determine if a range shift is facilitated (b; green seedlings) by suitable microhabitat (dark green) or is constrained by unsuitable microhabitat (c; light green). Darker colors in both blue (macroclimate) and green (microhabitat) lines indicate increased suitability for establishment.

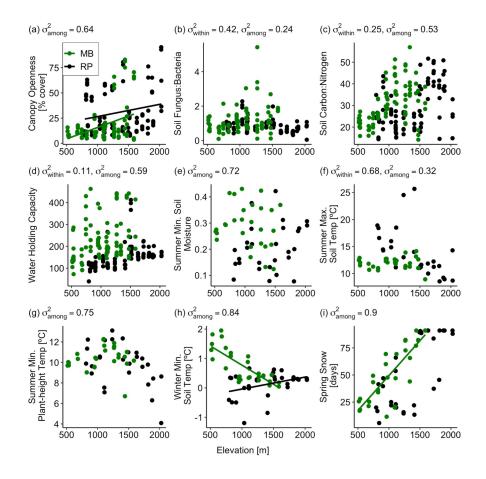
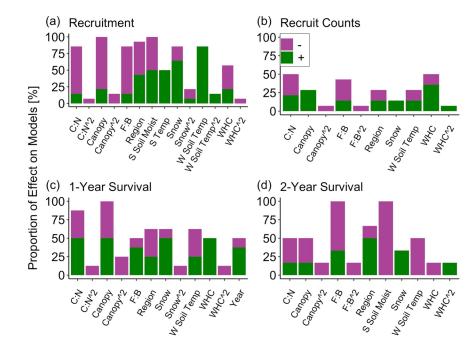
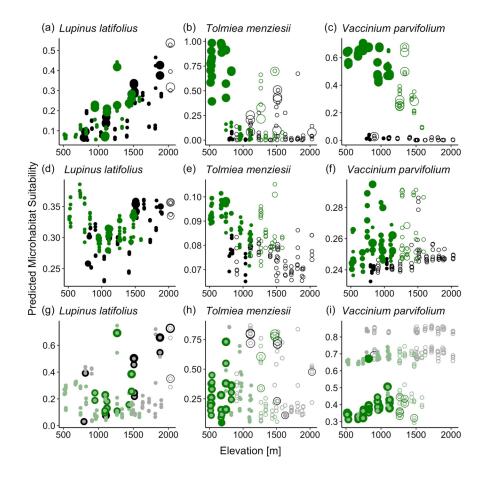


Figure 2. Both microhabitat parameters that are not expected to be directly affected by climate change (a-d) or expected to directly respond to climate change (e-i) differed in their within-site variability and show variable relationships with elevation. We show within  $(\sigma^2_{\text{within}})$  and among  $(\sigma^2_{\text{among}})$  site percent variation explained. These variance partitioning results are from Linear Mixed Models (LMMs) of the microhabitat parameter on the y-axis and block nested within site (microhabitat parameter ~ 1 + (1|site/block). In cases of model fitting issues, we ran the LMM with only (1|site) as a random effect and thus only report among-site percent variation explained. We also fit LMMs to test the effects of quadratic elevation, transect, and their interaction on the microhabitat parameter indicated on the y-axis, with a random effect of site (microhabitat parameter ~ poly(elevation, 2) \* transect + (1|site)). We show one fitted line for models for significant (alpha = 0.05) effects of elevation (line is shown for MB transect only), and two fitted lines for significant elevation\*transect interaction even if elevation itself was not significant. Note that fitted lines do not include random effects. Legend for all plots is as in (a) with different colors indicating the warmer, wetter West (MB) transect or the cooler, driver East (RP) transect.



**Figure 3.** Microhabitat parameters identified as most important vary for likelihood of recruitment (a), relative recruit counts (b), 1-year seedling survival (c), and 2-year seedling survival (d). Percentages calculated from the total times a parameter was used in model selection for all species, with pink indicating a negative effect and green indicating a positive effect. Legend in all panels is as in (b). Note that each panel corresponds to a different number of models (see Tables 1, S3). Summer temperature and minimum soil moisture were only used in model selection for two species because of missing data at many sites. Parameter abbreviations are C:N - soil carbon:nitrogen; Canopy - canopy openness; F:B - soil fungus:bacteria; Region - transect; S Soil Moist - summer minimum soil moisture; S Temp - summer maximum soil temperature for (a), (b) and summer minimum plant-height temperature for (c), (d); Snow - spring days of snow cover; W Soil Temp - winter minimum soil temperature; WHC - water holding capacity, with '^2' indicating a quadratic effect.



**Figure 4.** Microhabitat suitability predicted for each plot across the elevation gradient of our study system for three representative species with sites beyond their current thermal range limit (below range limit = solid points; beyond = hollow points; see Fig. S2) varies by species and life stage. Predictions are generated from model averaging results testing the effects of microhabitat on likelihood of recruitment (a-c), relative recruit counts (d-f), and 1-year seedling survival (g-i). Note that these predictions are not based on elevation, and that y-axis scales differ. Points are colored by the warmer, wetter West (green) or cooler, drier East (black) transects and size of points corresponds to observed recruitment (a-c), relative recruit counts (d-f), or proportion of seedlings that survived to one year (g-i) in a plot. Lighter colors for both transects indicate plots without recruitment and thus this size does not reflect an observed value. Full predictions are shown in Fig. S7.

### Tables

**Table 1.** Model averaging results (or best model, when no other models were within  $\Delta$  AIC < 2, indicated with '\*') for assessing importance of different microhabitat variables in explaining likelihood of recruitment (i.e. did seeds germinate or not), relative recruit counts (i.e. how many seedlings recruited relative to maximum recruit counts for the species), and 1- or 2-year seedling survival. '+' indicates a positive parameter estimate and ''' indicates a negative parameter estimate. Columns are colored by the expectation of those microhabitat variables being directly (blue) or indirectly (green) affected by climate change. Blank cells indicate that the parameter was not chosen in model selection. Because many of our soil loggers were compromised by animal disturbance, we only used those variables for species that showed no response at

sites with compromised data: recruitment and recruit counts for *M. nervosa, E. lanatum* (summer maximum soil temperature, summer minimum soil moisture); seedling survival for *R. ursinus* (summer plant-height minimum temperature, summer minimum soil moisture). These variables are listed as 'NA' for other species. Summer plant-height minimum temperature was never selected so it is not included in this table. Because of the link function in our binomial GLMs, the results can be interpreted as increasing or decreasing the log odds of recruitment or seedling survival, corresponding to lower or higher likelihoods, respectively. Abbreviations are as follows: Yr = year; Tr = transect; Canopy = canopy openness; F:B = fungus: bacteria; C:N = carbon:nitrogen; WHC = water holding capacity; Sum Soil Temp = summer maximum soil temperature; Sum Soil Moist = summer minimum soil moisture; Wint Soil Temp = winter minimum soil temperature; Snow = spring days with snow.

Species	Model	Ν	Yr	$\mathbf{Tr}$	Canopy	$Canopy^2$	F:B	$F:B^2$	C:N	C:N^2	WI
Abies lasiocarpa	recruitment	174		+	-		-		+		-
Abies lasiocarpa	recruit counts	10							+		
Anemone occidentalis	recruitment	174		+	-		-		-		
Anemone occidentalis	recruit counts	10					+		-		
Eriophyllum lanatum	recruitment	165		+	-				-		+
Eriophyllum lanatum	recruit counts $*$	8									
Erigeron peregrinus	recruitment	174		+	-	-	+		-		
Erigeron peregrinus	recruit counts	23			+		-	-			+
Erigeron peregrinus	1-yr survival	31	+		+				-		
Erigeron peregrinus	2-yr survival	28		+	-		-				
Lupinus latifolius	recruitment	174		-	+		+				+
Lupinus latifolius	recruit counts	30		+	+		-				
Lupinus latifolius	1-yr survival	33			+	-	-				+
Lupinus latifolius	2-yr survival	28		+			-				
Mahonia aquifolium	recruitment	174		+	-		-		-		-
Mahonia aquifolium	recruit counts	8							+		+
Mahonia nervosa	recruitment	165		-	-		-		-		-
Mahonia nervosa	recruit counts	9									+
Picea engelmannii	recruitment	173		-	-		-		-	-	
Picea engelmannii	recruit counts	11									I
Rubus ursinus	recruitment	171		-	-				-		+
Rubus ursinus	recruit counts	33							-		+
Rubus ursinus	1-yr survival	39		-	+		+		+		
Rubus ursinus	2-yr survival*	28		+			+				
Sorbus sitchensis	recruitment	173		-	+		-		+		
Sorbus sitchensis	recruit counts	76		+	+	-	-		+		+
Sorbus sitchensis	1-yr survival	92	+	-	-	-			+		
Sorbus sitchensis	2-yr survival	100		-	+	-	-	-	-		-
Tellima grandiflora	recruitment	170		+	-		-		-		-
Tellima grandiflora	recruit counts	29		-	+		+				-
Tellima grandiflora	1-yr survival	57	+		-				-	-	+
Tellima grandiflora	2-yr survival	46			-		+		+		
Tolmiea menziesii	recruitment	171			-		-		-		-
Tolmiea menziesii	recruit counts	33		-			-		-		
Tolmiea menziesii	1-yr survival	53		+	-				-		+
Tolmiea menziesii	2-yr survival*	32					-		-		
Vaccinium deliciosum	recruitment	174		-	-	-	-				
Vaccinium deliciosum	recruit counts	24							-		-
Vaccinium deliciosum	1-yr survival	26		-	+		+		+		
	recruitment	174		-	+		_		_		
Vaccinium parvifolium	recruitment	174		-	+		-		-		

Vaccinium parvifolium	recruit counts	43						+
Vaccinium parvifolium	1-yr survival	43	-	+	-	+	+	+