

Navigating Uncharted Waters: Could COVID-19 and/or Certain COVID-19 Vaccines Promote Malignancy?

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September 21, 2023

Title: Navigating Uncharted Waters: Could COVID-19 and/or Certain COVID-19 Vaccines Promote Malignancy?

Keywords: Cancer, COVID-19, SARS-CoV-2, spike glycoprotein, vaccines, malignancy, recurrence, metastasis

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Word count: 6,485

Charts, tables or figures: 1 table, 2 figures

THIS MANUSCRIPT HAS BEEN SUBJECTED TO PEER REVIEW BY *FRONTIERS IN ONCOLOGY* (Molecular and Cellular Oncology Section). While three (out of four) external reviewers endorsed the manuscript for publication, the study was rejected on editorial grounds.

ABSTRACT

Cancer is a complex and dynamic disease. The “Hallmarks of Cancer” were proposed by Hanahan and Weinberg (2000) as a set of biological capabilities acquired by human cells as they make their way from normalcy to neoplastic transformation. These capabilities include self-sufficiency in proliferative signaling, insensitivity to growth-suppressive signals and immune surveillance, ability to evade cell death, enabling replicative immortality, reprogramming energy metabolism, inducing angiogenesis, and activating tissue invasion and metastasis. Underlying these capabilities are genome instability, which expedites their acquisition, and inflammation, which fosters their function/s. Additionally, cancer exhibits another dimension of complexity: a heterogeneous repertoire of infiltrating and resident host cells, secreted factors, and extracellular matrix, known as the tumor microenvironment, that through a dynamic and reciprocal relationship with cancer cells supports immortality, local invasion, and metastatic dissemination. This staggering intricacy calls for caution when advising all people with cancer (or a previous history of cancer) to receive the COVID-19 primary vaccine series plus additional booster doses. Moreover, because these patients were not included in the pivotal clinical trials, considerable uncertainty remains regarding vaccine efficacy, safety, and the risk of interactions with anticancer therapies, which could reduce the value and innocuity of either medical treatment. After reviewing the available literature, we are particularly concerned that COVID-19 vaccination may predispose some (stable) oncologic patients to cancer progression, recurrence and/or metastasis. This hypothesis is based on biological plausibility (i.e., induction of lymphopenia and inflammation; downregulation of ACE2 expression; activation of oncogenic cascades; sequestration of tumor suppressor proteins; dysregulation of the G4-RNA-protein binding system and type I IFN responses; unsilencing of LINE-1 retrotransposons) together with growing anecdotal evidence and reports filed to Vaccine Adverse Effects Report System (VAERS) suggesting that some cancer patients experienced disease exacerbation or recurrence following COVID-19 vaccination. In light of the above, and because some of these concerns also apply to cancer patients infected with SARS-CoV-2, we encourage the scientific and medical community to urgently evaluate the impact of both COVID-19 and COVID-19 vaccination on cancer biology, adjusting public health recommendations accordingly.

INTRODUCTION

A number of estimates and modelling studies highlight the millions of lives that COVID-19 vaccines might have saved globally (1-6). Yet, the COVID-19 crisis has negatively impacted the health and well-being of many people, particularly those living with cancer. Three years into the pandemic, healthcare authorities keep recommending that people with active and prior cancer get vaccinated against COVID-19 (7). Booster doses are encouraged (7,8) because vaccine effectiveness wanes with time (9) and some cancers and cancer treatments affect the immune system, rendering the vaccines less efficient (10). While clinical trials for COVID-19 vaccines overlooked patients with cancer (11-15), the assumption is that those with a compromised immune system are at higher risk for severe disease, so getting even some protection from the vaccine is better than no protection. However, a growing body of anecdotal evidence (16-21) suggests that some individuals with active or prior cancer experienced disease exacerbation following COVID-19 vaccination. Reports registered in VAERS (22), a national self-reporting vaccine safety surveillance system co-managed by the U.S. Centers for Disease Control and Prevention (CDC) and U.S. Food and Drug Administration (FDA), also revealed a noncausal association between COVID-19-vaccination (namely mRNA-based vaccines) and cancer, relative to other vaccines (23).

While malignancies are generally understood to take months or, more commonly, years to progress such that the existence of a potential long-term health threat cannot be fully ascertained at present, some fast-

91 acting cancers and the reawakening of dormant cancer cells (DCCs), which is associated with cancer
92 recurrence and metastasis, are often aggressive processes that can be rapidly detected (24,25). To our
93 knowledge, prospective pharmacovigilance and/or monitoring of vaccinated recipients versus matched
94 unvaccinated controls have not been pursued in well-designed clinical trials. Additionally, national
95 estimates of cancer recurrence are not routinely collected by cancer registries (26). This article aims to
96 highlight the pressing need to study and compare the incidence of cancer complications after COVID-19
97 vaccination with the incidence of similar events after SARS-CoV-2 infection (in the unvaccinated
98 population). Advancing research on this topic/s will help health authorities to a) properly assess the risk-
99 benefit ratio of COVID-19 vaccination in a population at increased risk of severe COVID-19 outcomes
100 (27) and b) draw more robust conclusions with regard to vaccination (or appropriate alternatives) in
101 patients with a current cancer diagnosis or cancer history.

102 **THE HYPOTHESIS**

103
104 Based on the supporting evidence discussed below, we hypothesize that COVID-19 and/or certain
105 COVID-19 vaccines generate a pro-tumorigenic milieu that predispose some (stable) cancer patients and
106 survivors to disease progression and/or (metastatic) recurrence. Focus is placed on vaccines that promote
107 the endogenous production of SARS-CoV-2 spike (S) glycoprotein, namely mRNA vaccines
108 (Pfizer/BioNTech, Moderna) and adenovirus-vectorized vaccines (Johnson & Johnson,
109 Oxford/AstraZeneca) (28). We acknowledge that other clinical and social factors resulting from the
110 pandemic, such as adverse effects related to SARS-CoV-2 infection (29,30); steep declines in cancer
111 screening, diagnosis and treatment (31); adoption of unhealthy behaviors (i.e., increased alcohol
112 consumption, reduced physical activity) during long pandemic lockdowns (32); stress induced by the
113 COVID-19 crisis (33); and the assumption that millions of adults will remain unemployed and without
114 health insurance; will independently contribute to cancer mortality in the months and years to come.

115 **SUPPORTING EVIDENCE**

116
117 SARS-CoV-2 spike glycoprotein-based vaccines, and particularly mRNA vaccines, have the potential to
118 initiate a set of biological mechanisms that may collectively generate a (transient) pro-tumorigenic
119 environment favorable to cancer progression and/or reactivation of dormant cancer cells (DCCs). These
120 adverse effects may be attributed to the proinflammatory action of the lipid nanoparticles (LNPs); the
121 impaired type I interferon (IFN) response and/or translational dysregulation of cellular microRNAs
122 triggered by structurally modified mRNA (mRNA vaccines); as well as to the unique nature, expression
123 pattern, binding profile, and proinflammatory and tumorigenic effects of the produced antigens, namely
124 the SARS-CoV-2 spike protein and/or its subunits S1 and S2 (mRNA and adenovirus-vectorized
125 vaccines) (Fig.1). In addition, high levels of soluble spike and/or its subunits and peptide fragments have
126 been found in the circulation of vaccinees, where they persist for weeks, or even months. It is thus
127 plausible that the sustained and systemic distribution of spike within the human body (viral spike will
128 not, in most cases, impact tissues and organs other than the respiratory tract) may promote a range of
129 unforeseen interactions with angiotensin-converting enzyme 2 (ACE2), the entry receptor for SARS-
130 CoV-2, either in its soluble circulating form or expressed in cells from various tissues and organs.
131 For the foregoing reasons, it is imperative to understand the effects of COVID-19 and COVID-19
132 vaccination on cancer cells and their microenvironment.

133 **Lymphopenia is a hallmark of both severe coronavirus disease (COVID-19) and COVID-19 vaccination.**

134
135 Lymphopenia, a condition defined by abnormally low counts of lymphocytes, is a feature of severe
136 COVID-19 compared with non-severe disease (34-36). Possible underlying causes for the observed

lymphopenia, especially the decrease in T cell counts, include: T cell redistribution into infected organs, activation-induced exhaustion, apoptosis, and pyroptosis (37). While T cell exhaustion is observed in other viral infections (38), it seems to be more rapid, profound, and long-lasting in the setting of COVID-19. A recent study suggests that lymphopenia in severe COVID-19 patients is likely to result from SARS-CoV-2 infection of T cells in a spike-ACE2-independent manner (39). Additionally, it has been reported that the expression of S alone is sufficient to induce a rapid membrane fusion to produce syncytium, which could readily internalize multiple lines of lymphocytes to form typical cell-in-cell structures, leading to the death of internalized cells (40).

Lymphopenia has also been associated with COVID-19 vaccination. Phase-I/II clinical trials with the BNT162b1 (Pfizer/BioNTech) (41) and ChAdOx1 (Oxford/AstraZeneca) (42) vaccines described a dose-dependent decrease in plasma lymphocytes 6-8 days post-vaccination in 45-46% of the participants. Consistently, two pre-prints based on the immunization programs in Israel (BNT162b1 vaccine) (43) and England (BNT162b1 and ChAdOx1 vaccines) (44) reported an initial surge in infection risk up to 9 days following vaccination. Nonetheless, T-lymphocytes specific to SARS-CoV-2 viral antigens have been shown to ultimately increase after immunization with both genetic vaccines (i.e., spike-specific T cells) and traditional platforms such as the multiantigen modified vaccinia virus Ankara (MVA)-based COVID-19 vaccine COH04S1 (i.e., membrane-, nucleoprotein-, and spike-specific T cells) (45,46).

Even though the molecular mechanisms that underlie lymphopenia in both COVID-19 infection and vaccination are not fully understood, lymphopenia has long been associated with increased cancer incidence and risk of malignancy (47). Lymphocyte alterations are frequent in patients with cancer and strongly impact prognosis and survival (47,48). Severe CD4⁺ T cell lymphopenia is one of the hallmarks of human immunodeficiency virus (HIV) infection. People who have HIV/AIDS are at higher risk of developing certain types of tumors (i.e., Kaposi sarcoma) than people without the disease (49-51). CD8⁺ T cells have a crucial function in immune-mediated dormancy, and their depletion releases the brakes on DCCs leading to metastatic outgrowth (52,53). Anesthetic-induced immunosuppression can promote cancer relapses depending on dose, duration and timing of use (54). Exposure to immunosuppressive drugs that prevent organ rejection in organ transplant recipients, impairs cancer surveillance and facilitates the action of oncogenic viruses, increasing the post-transplant risk of neoplastic complications (55). Analogously, organ transplant recipients accepting an organ from a cancer survivor donor might develop malignancy because exposure to the immunosuppressant drugs allows hidden latent metastases (transplanted with the organ) to spring to life (56). Of note, 25% of cancers developed in patients with organ transplants, experience a clinical remission when the administered dose of the immunosuppressive drug is drastically reduced (57). This strongly suggests that recovery of immune function results in eradication of tumor cells. Remarkably, some types of cancer treatment, such as chemotherapy, radiation, and the combination of chemotherapy and immunotherapy can also cause severe lymphopenia, which is correlated with reduced survival (47,58,59).

Given that lymphopenia, together with inflammation-related factors (described below), contributes to create a microenvironment favorable to cancer progression and/or reawakening of DCCs, extreme caution is needed when recommending COVID-19 vaccination (up to 5 doses) (8) to oncologic patients, especially those undergoing anticancer treatment. Comprehensive studies concerning the molecular mechanisms that lead to overall lymphocyte reduction in both COVID-19 patients and vaccinees should help identify improved vaccination strategies and/or alternative interventions that prevent this major immunological abnormality and its consequences.

The SARS-CoV-2 spike glycoprotein and its S1 subunit elicit cell signaling *in vitro* that might be conducive to tumorigenesis *in vivo*.

SARS-CoV-2 contains a spike (S) protein that consists of two subunits: S1 and S2. S1 aids the virus to infect human cells by binding to angiotensin-converting enzyme 2 (ACE2), a multifunctional protein mostly expressed on the surface of many cells (60,61). S2 mediates the membrane fusion process (62). In addition to facilitate the entry of SARS-CoV-2 into the host cells, the interaction between spike and ACE2 elicits cell signaling in those cells expressing ACE2 (63). Data show that, in lung vascular cells and cells implicated in the development of pulmonary arterial hypertension, the S1 subunit of spike alone, activated MEK, the modulator of Extracellular Signal-Regulated Kinase (ERK) (63), which is a signal transduction mechanism for cell growth (64). In addition, Patra and collaborators (65) conveyed that the full-length spike, through the downregulation of ACE2 expression, promoted an Angiotensin II Type I receptor (AT₁R)-mediated signaling cascade; induced the transcriptional regulatory molecules nuclear factor- κ B (NF- κ B) and activator protein 1 (AP-1)/c-Fos via MAPK activation; and increased interleukin 6 (IL6) levels in epithelial cells (65) (Fig.2). NF- κ B activation in cancer cells promotes proliferation, chemoresistance and invasion whereas, in the tumor microenvironment, stimulates angiogenesis and immune suppression, collectively supporting the metastatic process (66). The mitogen-activated protein kinase Ras/Raf/MEK/ERK cascade is frequently involved in malignancy (67). Indeed, over 30% of all human cancers are driven by Ras genes (68-75). Elevated levels of IL-6 correlate with increased rates of tumor relapse in breast cancer and head and neck cancer (76,77). By contrast, inhibition of IL-6/STAT3 signaling reduced cancer recurrence in preclinical models of breast, head and neck, and hepatocellular carcinoma (78-80). The AT₁R-mediated signaling cascade also activates phosphatidylinositol-3-kinase (PI3K), a component of one of the most important intracellular pathways (PI3K/AKT/mTOR) and a master regulator for cancer (67,81). Overactivation of this pathway is present in many human malignancies and has been implicated in cancer progression. Consistently, the use of PIK3 inhibitors is a common approach in the treatment of tumors (82).

Considering that a) human cells sensitively respond to spike and/or its S1 subunit to elicit ACE2 cell signaling, and b) ACE2 exerts multiple anti-tumoral and anti-invasive effects, including inhibition of cancer angiogenesis and metastasis, the prolonged (or even transient) spike-mediated ACE2 downregulation (or loss) could *per se* promote tumor progression (83-86). Remarkably, free-floating spike, S subunits, and S peptide fragments have been found to enter the circulation and persist in the body for weeks (87,88) and even months (89) following COVID-19 vaccination at concentrations comparable to those found in severe COVID-19 patients (89,90) (Table I). It is hence imperative to monitor the mid- and long-term consequences of COVID-19 vaccines that introduce spike into the human body. Most importantly, appropriate experimental animal models should be developed to understand the contribution and functional implications of these signaling cascades in relation to cancer progression, recurrence and/or sensitivity to cancer therapies.

The mRNA vaccines are designed to deactivate the host innate immunity via Toll-Like Receptors (TLRs), compromising type I IFN responses.

DNA and RNA stimulate the mammalian innate immune system through the activation of Toll-Like Receptors (TLRs), a class of proteins mostly expressed in sentinel cells (i.e., dendritic cells, macrophages) that constitute the first line of defense against invading pathogens and endogenous molecules released from dying or damaged cells (91). TLRs trigger multiple signaling pathways involving nuclear factor- κ B (NF- κ B), interferon regulatory factors (IRFs), and mitogen-activated protein kinases (MAPKs) for the production of various cytokines that play important roles in many diseases, including cancer. RNA particularly signals through human endosomal TLR3, TLR7 and TLR8; however, incorporation of modified nucleosides into the RNA molecule ablates TLR activity (92,93). COVID-19 mRNA vaccines have all uridines in the SARS-CoV-2 spike mRNA sequence synthetically

replaced by N1-methyl pseudouridines (m1Ψ) (94,95). Such replacement increases biological stability, promotes mRNA translation, and dramatically inhibits innate immune sensing since uncontrolled immune activation might lead to undesirable allergic reactions and anaphylactic shock (94,96).

In spite of the critical contribution of pseudouridines to mRNA COVID-19 vaccines, little is known about the biological consequences of delivering highly-stabilized m1Ψ-modified mRNA within the cytoplasm of human cells. For instance, an effective immune response necessarily involves the induction of a robust TLR-mediated type I IFN signaling cascade as part of the innate immune system. If this response is ablated, immunopathology during lytic and latent viral infections may result (97-99). Defects in TLR expression have been reported in people with herpesvirus infections (100,101). Mutations in *TLR3* and its downstream signaling molecules have been associated with cases of herpes simplex virus encephalitis (102,103), varicella zoster virus meningoencephalitis (102), and recurrent herpes zoster ophthalmicus (103). Strikingly, an increasingly high number of herpes zoster cases has been reported following mRNA (BNT162b2 and mRNA-1273) but not adenovirus-vectorized or inactivated COVID-19 vaccination (104-109). Such observation is consistent with an impaired TLR-mediated type I IFN response triggered by m1Ψ-modified mRNA. Multimodal single-cell profiling of peripheral blood of patients with acute COVID-19 and healthy volunteers before and after receiving the BNT162b2 mRNA (Pfizer/BioNTech) injection also revealed dramatic differences in response to both immune challenges. In COVID-19 patients, immune responses were characterized by a highly augmented type I IFN response, which was largely absent in vaccine recipients. Increased IFN signaling likely contributed to the drastic upregulation of cytotoxic genes in the peripheral T cells and innate-like lymphocytes observed in COVID-19 patients. Analysis of B and T cell repertoires revealed that while the majority of clonal lymphocytes in COVID-19 patients were effector cells, in vaccine recipients, clonal expansion was primarily restricted to circulating memory cells (110).

Despite the above mentioned, there is no ample consensus on whether type I IFN activity is robust (23,110,111) or compromised (112,113) during SARS-CoV-2 infection. For instance, a study using primary cells from macaque lung bronchoalveolar lavage (113) provided evidence that the SARS-CoV-2 S1 spike subunit directly suppresses the expression of ACE2 and type I IFNs, contributing to SARS-CoV-2-associated lung disease. Additionally, COVID-19 diagnosis in ≥ 50 -year-olds has been associated with an increased risk of developing herpes zoster (114,115). This apparent controversy could be partially explained by the fine tuning between acute antiviral immune responses that quickly achieve infection clearance through high IFN secretion, and those that lead to longer and more robust inflammatory patterns (i.e., severe forms of COVID-19) with functional exhaustion of IFN responses (116). Notwithstanding, peripheral lymphopenia (described in both severe COVID-19 patients and COVID-19 vaccinees) could alternatively (or additionally) justify the reactivation of latent herpes zoster infections in both COVID-19 patients and people who received the COVID-19 mRNA vaccines.

Notably, TLRs are expressed not only in immune cells but also in tumor cells, where they can both inhibit and promote malignancy (117). Copious studies in humans and mice underline the importance of endogenous type I IFN, produced by both immune and tumor cells, in the control of tumor growth and in the response to antitumor therapies (118-120). Seneff and collaborators (23) extensively discuss the complexity and the role of type I IFNs, particularly IFN- α , in cancer surveillance and cancer suppression. The authors point out the dazzling range of anticancer effects initiated by IFN- α through both direct (i.e., cell cycle arrest, apoptosis, activation of natural killer and CD8⁺ T cells) and indirect (i.e., gene transcription activation of the JAK/STAT pathway) mechanisms (23). The Janus Kinase Signal Transducer and Activator of Transcription (JAK/STAT) pathway is dysregulated in several hematologic malignancies, and this has been shown to increase the metastatic potential in animal models of melanoma, colorectal cancer, and lymphoma (121). Defects in lymphocyte IFN signaling arise in

patients with breast cancer, melanoma and gastrointestinal cancer, and these defects may represent a common cancer-associated mechanism of immune dysfunction (120). Consistently, the exogenous administration of type I IFN and/or the use of type I IFN inducers boost the innate and adaptive immune responses against solid tumors (122,123).

Impairment of type I IFN responses is also observed in other diseases, including chronic infections (i.e., HIV/AIDS) and autoimmune conditions (i.e., multiple sclerosis -MS-). By interfering with type I IFN responses, HIV-1 can circumvent host antiviral signaling and establish persistent viral reservoirs. HIV-1-mediated defects in the IFN pathway include the impairment of protein receptors involved in pathogen detection, downstream signaling cascades required for type I IFN upregulation, and expression or function of key type I IFN-inducible, antiviral proteins (124,125). Remarkably, people infected with HIV have a substantially higher risk of some types of cancer compared with the general population including Kaposi sarcoma, non-Hodgkin lymphoma, cervical cancer (50) and, to a lesser extent, cancers of the anus, liver, oral cavity/pharynx, lung, and Hodgkin lymphoma (51). Similarly, patients with MS that have a suppressed type I IFN signaling and respond well to IFN-therapy (126,127) are also at greater risk of developing cancer than the general population (128). This increased risk is particularly apparent for prostate, breast, colorectal, and anal cancers, as well as cancers of the trachea, bronchus, and lung.

Overall, the exceedingly complicated and pleiotropic roles of TLR and type I IFN responses in tumor biology prompts caution when introducing synthetic (i.e., m1Ψs) mRNAs for *in vivo* therapeutic applications. Of relevance, disrupted TLR-mediated type I IFN responses following SARS-CoV-2 infection and mRNA vaccination may not be comparable for the following reasons. First, synthetic m1Ψ-modified mRNA, unlike viral RNA, has the ability to ablate TLR activity. Second, recent studies suggest that endogenous production of synthetic spike persists for a long time (> 6 months) within the human body (87-89). Third, whereas most of the viral S protein likely remains in the respiratory tract, vaccine-induced S protein production takes place in internal organs and tissues, thus being in the position to exert more systemic effects (129). Indeed, biodistribution studies of the BNT162b2 mRNA (Pfizer/BioNTech) vaccine in animal models revealed that the vaccine does not remain at the site of injection but rather accumulates in different organs (i.e., liver, spleen, lungs, ovaries, etc.) 48h post-inoculation (130-133). Last, compliance with multiple-dose vaccine schedules at relatively short intervals (8) may conceivably increase the risk of adverse effects in vaccine recipients. Further studies should thus shed light on relevant TLR-dependent pro- and anti-tumorigenic pathways that may be dysregulated as a result of mRNA vaccination and/or SARS-CoV-2 infection.

Codon optimization of COVID-19 vaccines may lead to the dysregulation of the G4-RNA-protein binding system, altering the translational regulation of cellular microRNAs.

The design of COVID-19 vaccines involves different types of optimizations, including codon-optimization (134,135). Codon optimization is a gene-engineering approach that uses synonymous codon changes to increase protein production in hosts that do not naturally express the gene. This process generally increases GC content, which correlates with an increased level of transcription, possibly as a result of decreased transcriptional pausing (136). Some authors advise that codon optimization compromises the safety and efficacy of biotech therapeutics (137). McKernan (138), Seneff (23), and others describe that the significant enrichment of GC content in COVID-19 mRNA vaccines (as compared to the native SARS-CoV-2 spike mRNA) might lead to an increase of secondary structures such as the G-quadruplexes (G4s) during translation. Specifically, McKernan and collaborators present a series of *in silico* approaches such as RNAfold and QGRSMapper that show changes to the secondary structure in the vaccine derived RNAs compared to the native virus (138). Of note is the increased number of G4 formations in the codon optimized mRNA vaccines (i.e., 19 and 9 G4 motifs in the

334 Moderna and Pfizer/BioNTech mRNAs, respectively, *versus* 4 G4 motifs in the spike coding region of
335 the SARS-CoV-2 virus). The abundance of G4 structures in the vaccinal mRNA likely amplifies the
336 attachment of RNA-binding proteins and micro RNAs that normally target human-expressed G4s for
337 normal regulation of human gene expression. Moreover, the use of N1-methylpseudouridines (m1Ψ) in
338 the vaccinal mRNAs further obscures the folding predictions as m1Ψ promiscuous base pairing
339 facilitates translation errors (139-141) and stabilizes G4s (142,143), thus exacerbating the impact of G4
340 formation with codon optimization (138).

341 Dysregulation of the G4-RNA-protein binding system might dramatically downregulate cellular
342 microRNA expression, which is involved in many pathological conditions such as cardiovascular
343 disease, onset of neurodegeneration, and cancer progression (23). One example, vital for cellular normal
344 housekeeping, is that of Mouse double minute 2 (MDM2) homolog, a physical negative regulatory
345 protein of p53 (which is a well-known tumor suppressor protein, as described below in further detail).
346 Dysregulation of micro RNAs that control the intricate interplay between MDM2 and p53, predictably
347 leads to an increased risk to a range of cancers (23,138, 144-146). Another example is the amplification
348 of G4 RNA repeats in amyotrophic lateral sclerosis/frontotemporal dementia -ALS/FTD- (*C9ORF72*
349 gene) and Fragile X syndrome (*FMR1* gene) (147). In these diseases, changes in the expression levels of
350 or mutations in RNA G4-binding proteins are also reported, suggesting that these proteins cannot exert
351 their critical function for normal neuron physiology when mutated or in cells with RNA G4 expansions
352 (147).

353 Largely, these observations highlight the evolved complexity of codon usage and challenge the scientific
354 bases for codon-optimization in human therapeutics.

355

356 **The lipid nanoparticles (LNPs) used in the mRNA vaccines are highly inflammatory in mice.**

357 Lipid nanoparticles (LNPs) are a vital component of mRNA-based COVID-19 vaccines, playing a
358 key role in improving the *in vivo* stability of mRNA and enhancing delivery to the cytosol of
359 antigen-presenting cells (148). LNPs consist of four main components: a neutral phospholipid,
360 cholesterol, a polyethylene-glycol lipid, and an ionizable cationic lipid (149).

361

362 The highly inflammatory properties of cationic LNPs have been known since 2010 (150). A recent
363 report (150) specifically showed that LNPs used in preclinical nucleoside-modified mRNA COVID-19
364 vaccines studies are highly inflammatory in mice. Intradermal injection of these LNPs led to massive
365 neutrophil infiltration, rapid and robust activation of diverse inflammatory pathways, and production of
366 various inflammatory cytokines and chemokines. Intranasal delivery led to similar inflammatory
367 responses in the lung (151). While the intrinsic adjuvant activity of LNPs may contribute to elicit
368 protective immunity, uncontrolled activation of various distinct and convergent inflammatory pathways
369 and the secretion of inflammatory cytokines and chemokines might lead to severe inflammation and
370 cytotoxicity. Extensive studies are therefore needed to map the interactions between cationic LNPs and
371 intracellular pattern-recognition receptors to unravel integrated and multifaceted mechanisms by which
372 these lipids induce inflammasome activation (152). In addition, while it is probable that intramuscular
373 injection of the COVID-19 vaccine LNP-mRNA complexes triggers similar responses in humans (151),
374 the exact nature of such responses and how much they overlap with the inflammatory signatures
375 documented in mice remain unknown. Relevantly, adenovirus-vectorized injections, unlike mRNA
376 vaccines, don't induce severe innate immune responses (i.e., cytokine storm), hyperinflammation, or
377 major damage in the targeted cells (153). Conversely, severe COVID-19 (which affects about 5% of the
378 SARS-CoV-2-infected population) (154), triggers a cytokine storm in pulmonary tissues which may be
379 accompanied by immunopathology, viremia, and systemic multiorgan collapse (155-157).

380

In the context of cancer, inflammation predisposes to the development of disease and promotes all stages of tumorigenesis (158). Tumor-extrinsic inflammation is caused by many factors including bacterial and viral infections, autoimmune diseases, obesity, tobacco smoking, asbestos exposure, and excessive alcohol consumption (158). Around 15-20% of all cancer cases are preceded by infection, chronic inflammation or autoimmunity at the same tissue or organ site (158-164). In such cases, cancer-promoting inflammation is induced and exists long before tumor formation. In contrast, cancer-intrinsic or cancer-elicited inflammation can be triggered by cancer-initiating mutations, contributing to malignant progression through the recruitment and activation of inflammatory cells (158). Both extrinsic and intrinsic inflammation can result in immunosuppression, thereby providing a preferred background for tumor development. Of note, neutrophils are actively involved in a network of inflammatory reactions that promote all the stages of tumor initiation, progression, angiogenesis and metastasis (165-170). Neutrophils form Neutrophil extracellular traps (NETs) that, when dysregulated, lead to the exacerbation of inflammation (171,172), unconstrained cancer progression, reawakening of DCCs, and metastatic dissemination, both in animal models and cancer patients (173). In addition, the tumor microenvironment, which is largely orchestrated by inflammatory cells, fosters proliferation, survival and migration of neoplastic cells. Markedly, inflammatory responses are aggravated on a background of pre-existing inflammatory conditions, as was recently demonstrated in a mouse model after administration of mRNA-LNPs (174). This effect was proven to be specific to the LNPs, acting independently of the mRNA cargo. Given that LNPs often accumulate in tumors, due to enhanced permeability and retention effect (EPR) (175-178), protecting cancer cells from transformation-related stress stimuli, including inflammation and the pro-tumorigenic action of NETs, is of paramount importance. Understanding the interactions between LNPs and neutrophils (179) should thus be critical for the development of safe and effective nanomaterials.

Potential reverse-transcription and genomic integration of foreign RNA are a source of genomic instability

A new study by Acevedo-Whitehouse and Bruno (180) discusses the possibility that parts of the SARS-CoV-2 genome might undergo reverse-transcription and genomic integration within infected cells, leading to persistent transcription of the integrated sequences. This hypothesis is based on an *in vitro* study that detected the presence of reverse-transcribed copies of SARSCoV-2 sequences in transfected human cells and found active transcription of the integrated sub-genomic segments (181). Acevedo-Whitehouse and Bruno speculate that the same phenomenon could occur in human cells that received COVID-19 mRNA vaccines. Indeed, a current study by Alden and collaborators (182) reported that an endogenous retrotransposon, namely Long Interspersed Nuclear Element-1 (LINE-1), was unsilenced following BNT162b2 mRNA (Pfizer/BioNTech) vaccine entry to the cell. This led to reverse transcription of full-length vaccine mRNA sequences and subsequent nuclear entry.

If these results are confirmed *in vivo*, the sustained activity of unsilenced LINE-1, which is normally repressed in somatic cells, might increase the risk of insertional mutagenesis of the reverse-transcribed molecules which, in turn, might disrupt coding regions, enhance the risk of mutations in tumor suppressor genes, and lead to sustained DNA damage in cells and tissues targeted by the vaccine (180). LINE-1 retrotransposition is indeed a major hallmark of cancer (183) and correlates with p53 mutations, copy number alterations, and cell cycle S phase checkpoints (184). Importantly, activation of LINE-1 increases the risk of epithelial-mesenchymal transition and metastasis in epithelial cancer, which accounts for 80-90% of all known human cancers (185). There is hence a pressing need for clarity on the potential COVID-19- and COVID-19 vaccine-induced activation of LINE-1 and its repercussions in cancerous and/or pre-cancerous cells with intrinsic high levels of LINE-1 expression.

Moreover, if SARS-CoV-2 spike mRNA vaccine sequences are reverse-transcribed, integrated into the genome of targeted cells, and expressed as chimeric transcripts that combine viral and cellular sequences, dysregulation of the G4-RNA-protein binding system might further promote malignancy. Indeed, experimental studies and bioinformatics predictions support the view that G4s are involved in different cellular functions associated to both DNA processes (i.e., telomere elongation, recombination and transcription) and RNA post-transcriptional mechanisms (i.e., pre-mRNA processing, mRNA turnover, targeting and translation) (186). As previously described, an increasing number of different diseases (i.e., neoplastic transformation, neurodegeneration) have been associated with the inappropriate regulation of RNA G4s, exemplifying the potential importance of these structures on human health. Notably, G4 structure formation, if not regulated efficiently, can stimulate genome instability, inducing mutations, deletions, and complex gross chromosomal rearrangements (187). A computational study that compared the location of potential G4 forming sites with cancer-associated breakpoints revealed a significant overlap, in particular in those cancers that harbor mutations in TP53 (the gene that codes for p53). This is underlined by computational studies in melanoma cells that linked G4 regions with mutational hot spots (188). Additionally, Hänsel-Hertsch and collaborators identified a direct correlation of G4s with mutational changes in different breast cancer entities (189). This supports the notion that G4 formation stimulates and influences mutation rates in different cancers.

The S2 subunit of SARS-CoV-2 spike glycoprotein interacts with tumor suppressor proteins p53 and BRCA-1/2 *in silico*.

Using bioinformatic (*in silico*) analyses, Singh and Bharara (190) proved that the S2 subunit of SARS-CoV-2 strongly interacts with well-known tumor suppressor proteins p53 and BRCA-1/2, which are frequently mutated in human cancers. These proteins provide a major barrier to neoplastic transformation and tumor progression by their unique ability to act as extremely sensitive collectors of stress inputs, and to coordinate a complex framework of diverse effector pathways and processes that protect cellular homeostasis and genome integrity. p53 and BRCA-1/2 act predominantly in the cell nucleus regulating cell-cycle progression, DNA-damage repair and recombination, and gene transcription (191-193). However, these proteins also play critical roles in the cytoplasm, triggering apoptosis and inhibiting autophagy thereby contributing to their mission as tumor suppressors (194,195). Wild-type p53 has been reported to be abnormally sequestered in the cytoplasm of a subset of primary human tumors (196). A myriad of cancer-associated mutations that disrupt nuclear targeting of BRCA-1, restrict the protein to the cytosol and diminish its nuclear function in homologous recombination repair of DNA breaks (197). Notably, BRCA-1 cytosolic accumulation promotes breast cancer metastasis (198) and independently predicts survival, tumor grade, and recurrence in low-grade basal-like sporadic breast cancers (199).

If, as *in silico*, the S2 subunit of spike interacts with tumor suppressor proteins *in vivo*, such a demonstration would have implications not only for the long-term health of those impacted by COVID-19 but also of those who received COVID-19 vaccination and repeated booster doses. Indeed, both mRNA and adenovirus-vectorized vaccines carry the genetic material that instruct the host cells to express S. As described above, biodistribution studies of the BNT162b2 mRNA (Pfizer/BioNTech) vaccine revealed its accumulation in different organs 48h post-inoculation (130-133). Most importantly, lipid nanoparticles, which are a vital component of the mRNA vaccines, preferentially accumulate in tumor tissue over healthy tissue due to enhanced permeability and retention (EPR) effect (175-178). Based on these findings, it is essential to decipher the range, detailed role, and biological consequences of the potential interactions between S2 and tumor suppressor proteins (i.e., p53, BRCA-1/2) in COVID-19 patients and vaccinees; particularly if these interactions confer a selective advantage (i.e., promotion of cancer cell survival, invasion, metastasis, chemoresistance) to cancer and/or precancerous cells.

Cancers associated with TP53 (the gene that codes for p53) mutations include breast cancer, bone and soft tissue sarcomas, brain tumors and adrenocortical carcinomas. Other less frequent cancers include leukemia, stomach cancer and colorectal cancer (200). Cancers associated with impaired BRCA1 activity include breast, uterine, and ovarian cancer in women; prostate and breast cancer in men; and a modest increase in pancreatic cancer for both men and women (201,202). The most commonly reported cancers with BRCA2 mutations include pancreas, prostate in men, and melanoma (203).

Dysregulation and/or aberrant changes in p53 levels/activity (204,205) as well as cytoplasmatic sequestration of BRAC-1 (206) have also been linked to neuronal dysfunction. Therefore, the potential *in vivo* interaction between S2 and tumor suppressor proteins might have consequences not only for rapidly cycling cancer cells but also for non-cycling cells (notably neurons) and thus for long-latency neurodegenerative diseases (207,208).

CD147 transmembrane protein, a novel entry route for SARS-CoV-2 infection to host cells, is correlated with various cancers

Recently, a novel SARS-CoV-2 entry route was proposed, namely utilization of the cluster of differentiation 147 (CD147) transmembrane glycoprotein (209). Despite lesser affinity towards the spike protein of SARS-CoV-2, as compared to ACE2, CD147 might be a complementary receptor in mediating virus infection (210). Although unequivocal evidence supporting a direct interaction between spike and CD147 is currently missing (211), confirmation of CD147 as a novel SARS-CoV-2 viral target might have serious implications for oncologic patients. CD147 has been correlated with various cancers (212-214) and has been shown to participate in the upregulation of the tumor microenvironment and cancer progression by several mechanisms, namely the control of glycolysis and its well-known ability to induce proteinases leading to matrix degradation, tumor cell invasion, metastasis and angiogenesis (215). As previously described for ACE2, the possible interaction of SARS-CoV-2 spike glycoprotein with CD147 receptors could, through activation of tumorigenic pathways, pave the way for cancer progression and/or recurrence.

DISCUSSION

COVID-19 vaccination is the largest emergency immunization campaign ever attempted in human history. Although the pandemic has largely vanished from public discourse, approximately 2,000-3,000 Americans are still dying from COVID-19 every week (216) and the same trend is observed in the U.K (217). Therefore, the protection of millions continues to be a tremendous challenge and responsibility. While vaccines may have had a significant impact in averting deaths, serious health outcomes from vaccines may go unrecognized in clinical trials and/or passive surveillance systems such as VAERS (218), especially if they are mid/long-latency and do not require immediate hospitalization. In this context, SARS-CoV-2 spike glycoprotein-based vaccines have the potential to induce DNA damage, promote inflammation, activate oncogenic pathways, and disrupt the fine tuning of the immune response. These dysregulated mechanisms and signaling pathways underlie most types of cancer.

While we understand that much of the discussion about cancer and COVID-19 vaccination was done under high pressure to protect this cohort from severe disease and death, a more balanced risk-benefit evaluation is urgently needed. This is especially relevant for people with poor immune responses, such as those with hematologic malignancies (219,220), for which the benefits of vaccination are dubious and the cumulative risks of successive boosters unknown (although conceivably increased with each dose received). Of particular concern is the observation that some anticancer drugs render COVID-19 vaccines ineffective (221,222). In addition, the coadministration of complex anticancer regimes and COVID-19 vaccines (222-224) might pave the way for intercurrent or synergistic toxic effects. Indeed, a

recent article (224) on the effects of BNT162b2 mRNA vaccine in oncologic patients under checkpoint inhibitors (CPIs) describes that CPI therapy resulted in a constant and variable increase of all COVID-19 vaccination side effects, which is alarming. Additionally, reactive axillary lymphadenopathy secondary to COVID-19 vaccines may mimic cancer metastasis, posing diagnostic dilemma and increasing anxiety in patients with breast cancer who received COVID-19 immunization (225-229). In contrast, a few rare cases of temporary or prolonged cancer remission after COVID-19 (230) and mRNA-based COVID-19 vaccination (231) have been reported, possibly as a result of the intense immune-inflammatory response that may have prompted anticancer immunity in these individuals. Overall, cancer is one of the most complex, heterogeneous and dynamic human diseases (232,233) and as such, a universal “one-size-fits-all” approach is flawed.

Unfortunately, most current cancer statistics worldwide (i.e., Japan, Australia, Canada, Europe) don’t extend beyond 2020 (234-239). This makes it imperative to build global pharmacovigilance databases that help in making decisions based on the best evidence available at each moment. In the U.S., from January 7, 2018 to July 2, 2022, the CDC mortality and morbidity weekly reports (MMWR) listed approximately 13,000 cancer deaths per week (range = 12,221–14,845), with peaks occurring in January 2021 (14,284 deaths) and January 2022 (14,845 deaths) (240). While the public health agency specified that the number of cancer deaths (with cancer as the underlying cause) increased slightly from 2018 to 2022, it mostly attributed the excess cancer deaths to noncancer underlying causes, such as COVID-19. Indisputably, the cancer mortality peaks observed in 2021 and 2022 correlate well with COVID-19’s winter surges. However, they also follow two major COVID-19 vaccination and booster campaigns. As noted earlier, both SARS-CoV-2 and SARS-CoV-2 spike protein-based vaccines promote the production of spike within human cells which, in light of the above, might facilitate malignant transformation. Chaotic death recording during pandemic waves might have also created a distortion of facts, misguiding efforts to prevent leading causes of cancer (and other) deaths. Indeed, research has found that, even under normal circumstances, critical errors in death certificates are quite common in the U.S., with the frequency of errors ranging from 18% to 85% or higher in hospital-based studies (241).

In short, despite the fact that many institutions (242,243) and authors (244,245) maintain that COVID-19 vaccines are safe and (partially) effective in patients with cancer, these claims are unsupported and recommendations are largely inferred from vaccine safety and effectiveness in the general population; performance of other vaccines in patients with cancer; and immune alterations inherent in current cancer treatments (246). Given the converging evidence of temporal association and biological plausibility, the contribution of genetic COVID-19 vaccines to cancer progression and recurrence cannot be excluded at present. Yet, one might argue that the oncogenic potential of spike should also be exerted during SARS-CoV-2 infection. While this is partially true, we already discussed that COVID-19 genetic vaccines and, in particular, mRNA injections, are radically different from SARS-CoV-2 viral infection. Hence, the role of COVID-19 vaccination and SARSCoV-2 infection in the pathways that potentially promote malignancy may not be comparable and merit further investigation. In addition, if harm can be conclusively attributed to the LNP vehicle itself and/or to the synthetic modified mRNA (regardless of the toxicity, or lack of thereof, of spike), this may have implications for the development of new mRNA products based on the same core technology (247).

In view of the current state of the art, our suggestion is that individuals with cancer or a history of cancer should receive the genetic COVID-19 vaccines only if the benefits clearly outweigh any risks and after careful evaluation case by case. Multidisciplinary clinical and basic research comparing the cellular and molecular basis of COVID-19- and COVID-19 vaccine-induced oncogenic effects may help rebalancing the risk-benefit profile of these products. Direct approaches, such as the use of animal models, should take advantage of the recent development of mice expressing human ACE2 receptors (248-250) and the

availability of cancer mouse models (250). Studies investigating the efficacy and safety of COVID-19 vaccination in cancer patients, both prospectively and retrospectively, are strongly encouraged. Patient-associated and treatment-associated factors merit specific consideration. The need for more reliable databases that include widely measured immune parameters as well as data on spike protein levels in blood has been pointed out by others (251). Taken together, these studies should provide robust data to guide clinical implementation, including the development of therapeutic alternatives (i.e., LNPs with different chemistry: a closed-form of spike not prone to ACE2 binding (252); non-spike targeting vaccines (253); platforms such as COH04S1 (254) with high tolerability and immunogenicity in immunosuppressed patients; non-pharmacological interventions (255), etc.), for those who do not benefit from active COVID-19 vaccination (and those who are allergic to some of the vaccine components).

CONCLUSION

Based on the comprehensive bibliographic research depicted here, we hypothesize that COVID-19 genetic vaccines, and particularly mRNA vaccines, have the potential to elicit a pro-tumorigenic milieu favorable to cancer progression and/or (metastatic) recurrence. Proving this hypothesis wrong is a necessary step towards satisfying the first principle of medicine: “Primum non nocere” (“First do no harm”). Indeed, all global crises pose tremendous challenges to health and welfare; however, such exceptionalities shouldn’t be a justification for lowering scientific standards. This is particularly relevant for prophylactic drugs intended to protect vulnerable high-risk populations across the world. Most importantly, because some of the outlined pro-oncogenic mechanisms are antigen-independent, current safety concerns (247, 256) should be promptly addressed before mRNA-based nanomedicines further transform the way diseases are managed and prevented in the future.

Table I. Concentration and persistence in the body of spike antigens after mRNA-mediated vaccination

ANTIGEN	VACCINE TYPE	CONCENTRATION (PG/ML)	TIME IN THE BODY (DAYS)	CITATION
S	mRNA-1273 mRNA-BNT162b	days 1–2 after 1 st dose - median S levels: 47 pg/mL (plasma) day 7 after 1 st dose - median S levels: 1.7 pg/mL (plasma) days 1-2 after 2 nd dose - median S levels: 1.2 pg/mL (plasma)	Present as late as 60 days post-second dose in germinal centers (lymph nodes) Present at least 1-2 days post-second dose (plasma)	107
S, S1	mRNA-1273	Mean S1 peak levels: 68±21pg/mL Mean S peak levels: 62±13pg/mL	S1 present up to 14 days post-first dose. Undetectable after 2 nd dose Peak levels at 5 days (plasma) S present up to 15 days post-first dose. Undetectable after 2 nd dose (plasma)	108
S fragments	mRNA-1273 mRNA-BNT162b		69-187 days post-vaccination (plasma)	109

FIGURE LEGENDS

Figure 1. Cancer-promoting molecular mechanisms and pathways potentially mediated by SARS-CoV-2 and/or certain COVID-19 vaccines.

Figure 2. Spike-mediated ACE2 downregulation and cell signaling might promote cancer progression in COVID-19 patients and vaccinees. ACE2 downregulation and its subsequent AT₁R-mediated response has the potential to encourage cancer progression and metastasis through its growth-promoting and proangiogenic activities.

ACE2 R: angiotensin-converting enzyme 2 acting as entry receptor for SARS-CoV-2; ACE2: angiotensin-converting enzyme 2; AT II: angiotensin II; AT₁R: angiotensin II type 1 receptor; PI3K: phosphatidylinositol 3-kinase; MAPK: mitogen-activated protein kinase; ERK: extracellular signal-regulated kinase; NF-kB: nuclear factor kB; IL-6: interleukin 6; AP-1: activating protein 1.

Figure 1

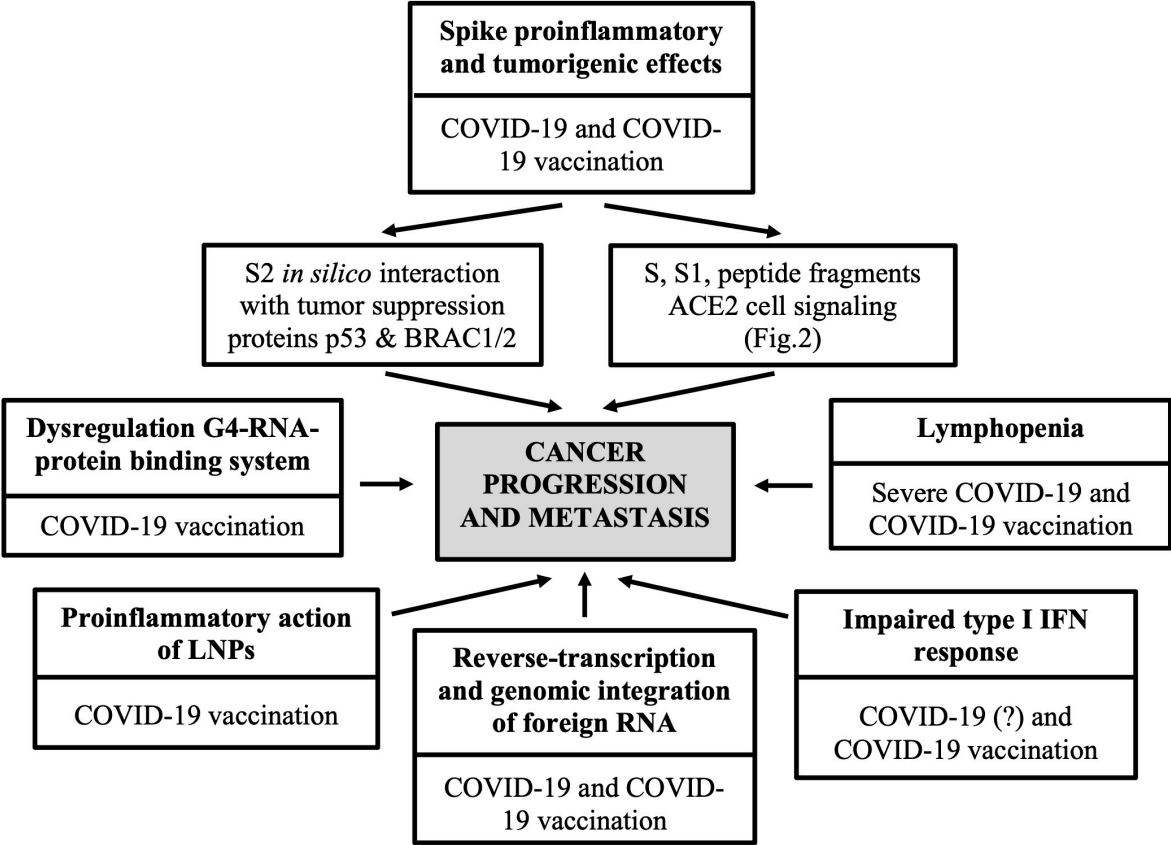
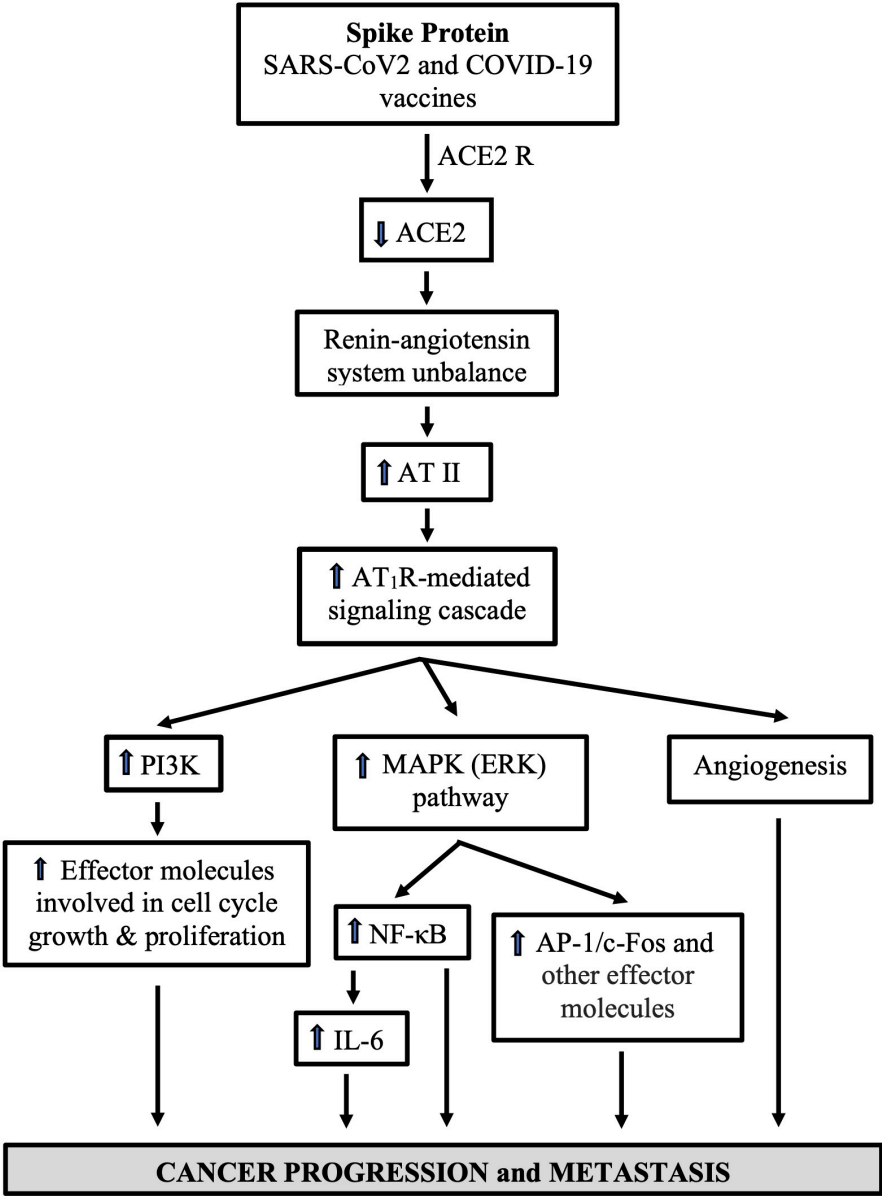


Figure 2



AUTHOR CONTRIBUTIONS

RV and YP contributed to the conception and design of the study. RV wrote the manuscript. YP provided essential contribution in reviewing and editing the manuscript. All authors made a substantial, direct, and intellectual contribution to the article and approved the submitted version.

ACKNOWLEDGMENTS

We thank Dr. Peter S. Spencer (Department of Neurology, Oregon Health & Science University, Portland, OR, USA) for useful discussion and Valerie S. Palmer (Department of Neurology, Oregon Health & Science University, Portland, OR, USA) and Enric Baya Castells for their encouragement and continuous support.

A preliminary version of the manuscript has previously appeared online as a preprint (257).

FUNDING

This research did not receive any specific grant from funding agencies in the public, commercial, or not-for-profit sectors.

CONFLICT OF INTEREST

The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

REFERENCES

1. Meslé MM, Brown J, Mook P, Hagan J, Pastore R, Bundle N et al. Estimated number of deaths directly averted in people 60 years and older as a result of COVID-19 vaccination in the WHO European Region, December 2020 to November 2021. *Euro Surveill* (2021) 26(47):2101021. doi: 10.2807/1560-7917.ES.2021.26.47.2101021. Erratum in: *Euro Surveill* (2022) 27(21): Erratum in: *Euro Surveill* (2022) 27(24).
2. Gupta S, Cantor J, Simon KI, Bento AI, Wing C, Whaley CM. Vaccinations against COVID-19 may have averted up to 140,000 deaths in the United States. *Health Aff (Millwood)* (2021) 40(9):1465-1472. doi: 10.1377/hlthaff.2021.00619.
3. Watson OJ, Barnsley G, Toor J, Hogan AB, Winskill P, Ghani AC. Global impact of the first year of COVID-19 vaccination: a mathematical modelling study. *Lancet Infect Dis* (2022) 22(9):1293-1302. doi: 10.1016/S1473-3099(22)00320-6.
4. Savinkina A, Bilinski A, Fitzpatrick M, Paltiel AD, Rizvi Z, Salomon J, Thornhill T et al. Estimating deaths averted and cost per life saved by scaling up mRNA COVID-19 vaccination in low-income and lower-middle-income countries in the COVID-19 Omicron variant era: a modelling study. *BMJ Open* (2022) 12(9):e061752. doi: 10.1136/bmjopen-2022-061752.
5. Steele MK, Couture A, Reed C, Iuliano D, Whitaker M, Fast H et al. Estimated number of COVID-19 infections, hospitalizations, and deaths prevented among vaccinated persons in the US, December 2020 to September 2021. *JAMA Netw Open* (2022) 5(7):e2220385. doi: 10.1001/jamanetworkopen.2022.20385.
6. Jones M, Khader K, Branch-Elliman W. Estimated impact of the US COVID-19 vaccination campaign. Getting to 94% of deaths prevented. *JAMA Netw Open* (2022) 5(7):e2220391. doi: 10.1001/jamanetworkopen.2022.20391.

7. Centers for Disease Control and Prevention (CDC). COVID-19 Booster Shot (2022). <https://www.cdc.gov/coronavirus/2019-ncov/vaccines/booster-shot.html> [Accessed May 2, 2022].
8. National Comprehensive Cancer Network (NCCN): Cancer and COVID-19 vaccination. Recommendations of the National Comprehensive Cancer Network® (NCCN) Advisory Committee on COVID-19 vaccination and pre-exposure prophylaxis (2022). https://www.nccn.org/docs/default-source/covid-19/covid-vaccine-and-cancer-05.pdf?sfvrsn=45cc3047_2 [Accessed May 2, 2022].
9. Ferdinands JM, Rao S, Dixon BE, Mitchell PK, DeSilva MB, Irving SA, et al. Waning 2-dose and 3-dose effectiveness of mRNA vaccines against COVID-19-associated emergency department and urgent care encounters and hospitalizations among adults during periods of delta and omicron variant predominance. VISION Network, 10 States, August 2021-January 2022. Morb Mortal Wkly Rep (2022) 71(7):255-63. doi: 10.15585/mmwr.mm7107e2.
10. Cancer Research UK. The immune system and cancer (2020). <https://www.cancerresearchuk.org/about-cancer/what-is-cancer/body-systems-and-cancer/the-immune-system-and-cancer> [Accessed September 13, 2022].
11. ClinicalTrials.gov. U.S. National Library of Medicine. National Institutes of Health. A study to evaluate efficacy, safety, and immunogenicity of mRNA-1273 vaccine in adults aged 18 years and older to prevent COVID-19. <https://clinicaltrials.gov/study/NCT04470427> [Accessed July 2, 2023].
12. ClinicalTrials.gov. U.S. National Library of Medicine. National Institutes of Health. A phase 1/2/3, placebo-controlled, randomized, observer-blind, dose-finding study to evaluate the safety, tolerability, immunogenicity, and efficacy of sars-cov-2 RNA vaccine candidates against COVID-19 in healthy individuals. <https://classic.clinicaltrials.gov/ct2/show/NCT04368728> [Accessed July 2, 2023].
13. ClinicalTrials.gov. U.S. National Library of Medicine. National Institutes of Health. A phase III randomized, double-blind, placebo-controlled multicenter study in adults, to determine the safety, efficacy, and immunogenicity of AZD1222, a non-replicating ChAdOx1 vector vaccine, for the prevention of COVID-19. <https://classic.clinicaltrials.gov/ct2/show/NCT04516746> [Accessed July 2, 2023].
14. Janssen Vaccines & Prevention B.V. A randomized, double-blind, placebo-controlled phase 3 study to assess the efficacy and safety of Ad26.COV2.S for the prevention of SARS-CoV-2-mediated COVID-19 in adults aged 18 years and older. Ensemble. Protocol VAC31518COV3001; AMENDMENT 3. <https://www.jnj.com/coronavirus/ensemble-1-study-protocol> [Accessed July 2, 2023].
15. He Z, Erdengasileng A, Luo X, Xing A, Charness N, Bian J. How the clinical research community responded to the COVID-19 pandemic: An analysis of the COVID-19 clinical studies in ClinicalTrials.gov. medRxiv [Preprint] (2020). doi: 10.1101/2020.09.16.20195552. Update in: JAMIA Open. 2021 Apr 20;4(2):ooab032 [Accessed July 2, 2023].
16. Goldman S, Bron D, Tousseyn T, Vierasu I, Dewispelaere L, Heimann P, et al. Rapid progression of angioimmunoblastic T cell lymphoma following BNT162b2 mRNA vaccine booster shot: A case report. Front Med (Lausanne) (2021) 8:798095. doi: 10.3389/fmed.2021.798095.
17. Adin ME, Wu J, Isufi E, Tsui E, Pucar D. Ipsilateral malignant axillary lymphadenopathy and contralateral reactive lymph nodes in a COVID-19 vaccine recipient with breast cancer. J Breast Cancer (2022) 25(2):140-144. doi: 10.4048/jbc.2022.25.e12.
18. Ang SY, Huang YF, Chang CT. Ph-positive B-cell acute lymphoblastic leukemia occurring after receipt of bivalent SARS-CoV-2 mRNA vaccine booster: A case report. Medicina (Kaunas) (2023) 59(3):627. doi: 10.3390/medicina59030627.

19. Plüß M, Mitteldorf C, Szusies CJ, Tampe B. Case report: Acquired hemophilia A following mRNA-1273 booster vaccination against SARS-CoV-2 with concurrent diagnosis of pleomorphic dermal sarcoma. *Front Immunol* (2022) 13:868133. doi: 10.3389/fimmu.2022.868133.
20. Norimatsu Y, Yoshizaki A, Yamada T, Akiyama Y, Toyama S, Sato S. Pemphigus vulgaris with advanced hypopharyngeal and gastric cancer following SARS-CoV-2 vaccination. *J Dermatol* (2023) 50(2):e74-e75. doi: 10.1111/1346-8138.16539.
21. Veeraballi S, Patel A, Are G, Ramahi A, Chittamuri S, Shaaban H. A case of chronic myelomonocytic leukemia unmasked after receiving J&J COVID-19 vaccine. *Cureus* (2022) 14(6):e26070. doi: 10.7759/cureus.26070.
22. Vaccine Adverse Event Reporting System (VAERS). <https://vaers.hhs.gov/> [Accessed Dec 2, 2022].
23. Seneff S, Nigh G, Kyriakopoulos AM, McCullough PA. Innate immune suppression by SARS-CoV-2 mRNA vaccinations: The role of G-quadruplexes, exosomes, and microRNAs. *Food Chem Toxicol* (2022) 164:113008. doi: 10.1016/j.fct.2022.113008.
24. Damen MPF, van Rheenen J, Scheele CLGJ. Targeting dormant tumor cells to prevent cancer recurrence. *FEBS J* (2021) 288(21):6286-303. doi: 10.1111/febs.15626.
25. Blasco MT, Espuny I, Gomis RR. Ecology and evolution of dormant metastasis. *Trends Cancer* (2022) 8(7):570-82. doi: 10.1016/j.trecan.2022.03.002.
26. American Cancer Society. Cancer treatment & survivorship. Facts and figures 2019-2021 (2022). <https://www.cancer.org/content/dam/cancer-org/research/cancer-facts-and-statistics/cancer-treatment-and-survivorship-facts-and-figures-2019-2021.pdf> [Accessed April 19, 2022].
27. Dai M, Liu D, Liu M, Zhou F, Li G, Chen Z, et al. Patients with cancer appear more vulnerable to SARS-CoV-2: A multicenter study during the COVID-19 outbreak. *Cancer Discov* (2020) 10(6):783-791. doi: 10.1158/2159-8290.CD-20-0422.
28. Poland GA, Ovsyannikova IG, Kennedy RB. SARS-CoV-2 immunity: review and applications to phase 3 vaccine candidates. *Lancet* (2020) 396 (10262):1595–606. doi: 10.1016/S0140-6736(20)32137-1.
29. Saini G, Aneja R. Cancer as a prospective sequela of long COVID-19. *Bioessays* (2021) 43(6):e2000331. doi: 10.1002/bies.202000331.
30. Francescangeli F, De Angelis ML, Baiocchi M, Rossi R, Biffoni M, Zeuner A. COVID-19-induced modifications in the tumor microenvironment: Do they affect cancer reawakening and metastatic relapse? *Front Oncol* (2020) 10:592891. doi:10.3389/fonc.2020.592891.
31. Sharpless NE. Q&A: Ned Sharpless on COVID-19 and cancer prevention. *Cancer Prev Res (Phila)* (2021) 14(6):615–8. <https://doi.org/10.1158/1940-6207.CAPR-21-0146>.
32. Cassata C. Five unhealthy pandemic habits and how to break them. *Healthline* (2022). <https://www.healthline.com/health-news/5-unhealthy-pandemic-habits-and-how-to-break-them> [Accessed September 6, 2022].
33. National Cancer Institute (NCI). Study suggests a link between stress and cancer coming back. (2021). <https://www.cancer.gov/news-events/cancer-currents-blog/2021/cancer-returning-stress-hormones> [Accessed April 21, 2022].
34. Tavakolpour S, Rakhshandehroo T, Wei EX, Rashidian M. Lymphopenia during the COVID-19 infection: What it shows and what can be learned. *Immunol Lett* (2020) 225:31-2. doi: 10.1016/j.imlet.2020.06.013.
35. Adamo S, Chevrier C, Cervia C, Zurbuchen Y, Raeber ME, Yang L et al. Lymphopenia-induced T cell proliferation is a hallmark of severe COVID-19. *bioRxiv* [Preprint] (2020). <https://doi.org/10.1101/2020.08.04.236521> [Accessed Feb 2, 2023].

36. Härter G, Spinner CD, Roider J, Bickel M, Krzmaric I, Grunwald S, et al. COVID-19 in people living with human immunodeficiency virus: a case series of 33 patients. *Infection* (2020) 48(5):681-6. doi: 10.1007/s15010-020-01438-z.
37. Zhang S, Asquith B, Szydlo R, Tregoning JS, Pollock KM. Peripheral T cell lymphopenia in COVID-19: potential mechanisms and impact. *Immunother Adv* (2021) 1(1):ltab015. doi: 10.1093/immadv/ltab015.
38. Guo Z, Zhang Z, Prajapati M, Li Y. Lymphopenia caused by virus infections and the mechanisms beyond. *Viruses* (2021) 13(9):1876. doi: 10.3390/v13091876.
39. Shen XR, Geng R, Li Q, Chen Y, Li SF, Wang Q, et al. ACE2-independent infection of T lymphocytes by SARS-CoV-2. *Signal Transduct Target Ther* (2022) 7(1):83. doi: 10.1038/s41392-022-00919-x.
40. Zhang Z, Zheng Y, Niu Z, Zhang B, Wang C, Yao X, et al. SARS-CoV-2 spike protein dictates syncytium-mediated lymphocyte elimination. *Cell Death Differ* (2021) 28(9):2765-77. doi: 10.1038/s41418-021-00782-3.
41. Mulligan MJ, Lyke KE, Kitchin N, Absalon J, Gurtman A, Lockhart et al. Phase I/II study of Covid-19 RNA vaccine BNT162b1 in adults. *Nature* (2020) 586 (7830):589-93. <https://doi.org/10.1038/s41586-020-2639-4>.
42. Folegatti PM, Ewer KJ, Aley PK, Angus B, Becker S, Belij-Rammerstorfer S et al. Safety and immunogenicity of the ChAdOx1 nCoV-19 vaccine against SARS-CoV-2: a preliminary report of phase I/II, single-blind, randomized controlled trial. *Lancet* (2020) 396(10249):467-78. doi: 10.1016/S0140-6736(20)31604-4.
43. Hunter PR, Brainard J. Estimating the effectiveness of the Pfizer COVID-19 BNT 162b2 vaccine after a single dose. A reanalysis of a study of “real-world” vaccination outcomes from Israel. *MedRxiv [Preprint]* (2021). doi: <https://doi.org/10.1101/2021.02.01.21250957> [Accessed Feb 10, 2023].
44. Lopez Bernal J, Andrews N, Gower C, Stowe J, Robertson C, Tessier E et al. Early effectiveness of COVID-19 vaccination with BNT162b2 mRNA vaccine and ChAdOx1 adenovirus vector vaccine on symptomatic disease, hospitalizations and mortality in older adults in England. *MedRxiv [Preprint]* (2021). doi: <https://doi.org/10.1101/2021.03.01.21252652> [Accessed Feb 10, 2023].
45. Chiuppesi F, Zaia JA, Frankel PH, Stan R, Drake J, Williams B, et al. Safety and immunogenicity of a synthetic multiantigen modified vaccinia virus Ankara-based COVID-19 vaccine (COH04S1): an open-label and randomized, phase 1 trial. *Lancet Microbe* (2022) 3(4):e252-e264. doi: 10.1016/S2666-5247(22)00027-1.
46. Volz A, Sutter G. Modified vaccinia virus Ankara: History, value in basic research, and current perspectives for vaccine development. *Adv Virus Res* (2017) 97:187-243. doi: 10.1016/bs.aivir.2016.07.001.
47. Ménétrier-Caux C, Ray-Coquard I, Blay JY, Caux C. Lymphopenia in cancer patients and its effects on response to immunotherapy: an opportunity for combination with cytokines? *J Immunother Cancer* (2019) 7(1):85. doi: 10.1186/s40425-019-0549-5.
48. Warny M, Helby J, Nordestgaard BG, Birgens H, Bojesen SE. Incidental lymphopenia and mortality: a prospective cohort study. *CMAJ* (2020) 192(2):E25-33. doi: 10.1503/cmaj.191024.
49. Penn I. Depressed immunity and the development of cancer. *Cancer Detect Prev* (1994) 18(4):241-52.
50. Hernández-Ramírez RU, Shiels MS, Dubrow R, Engels EA. Cancer risk in HIV-infected people in the USA from 1996 to 2012: a population-based, registry-linkage study. *Lancet HIV* (2017) 4(11):e495-e504. doi: 10.1016/S2352-3018(17)30125-X.
51. Wang CC, Silverberg MJ, Abrams DI. Non-AIDS-defining malignancies in the HIV-infected population. *Curr Infect Dis Rep* (2014) 16(6):406. doi: 10.1007/s11908-014-0406-0.

52. Eyles J, Puaux AL, Wang X, Toh B, Prakash C, Hong M et al. Tumor cells disseminate early, but immunosurveillance limits metastatic outgrowth, in a mouse model of melanoma. *J Clin Invest* (2010) 120(6):2030–9. doi: 10.1172/JCI42002.
53. Romero I, Garrido C, Algarra I, Collado A, Garrido F, Garcia-Lora AM. T lymphocytes restrain spontaneous metastases in permanent dormancy. *Cancer Res* (2014) 74(7):1958–68. doi: 10.1158/0008-5472.CAN-13-2084.
54. Kim, R. Effects of surgery and anesthetic choice on immunosuppression and cancer recurrence. *J Transl Med*. (2018) 16(1):8. doi: <https://doi.org/10.1186/s12967-018-1389-7>.
55. Gutierrez-Dalmau A, Campistol JM. Immunosuppressive therapy and malignancy in organ transplant recipients: a systematic review. *Drugs* (2007) 67(8):1167-98. doi: 10.2165/00003495-200767080-00006.
56. Tontonoz M. Stealth Mode: How metastatic cancer cells evade detection by the immune system. Memorial Sloan Kettering Cancer Center (2016) <https://www.mskcc.org/news/stealth-mode-how-metastatic-cancer-cells-evade-detection-immune-system> [Accessed April 20, 2022].
57. Fleuren J, Arko Gorter A, Kuppen PJK. Immune Surveillance 1998. Editor(s): Peter J. Delves. *Encyclopedia of Immunology* (Second Edition). Elsevier. Pages 1243-1247. <https://doi.org/10.1006/rwei.1999.0321>.
58. Grossman SA, Ellsworth S, Campian J, Wild AT, Herman JM, Laheru D, et al. Survival in patients with severe lymphopenia following treatment with radiation and chemotherapy for newly diagnosed solid tumors. *J Natl Compr Canc Netw* (2015) 13(10):1225-31. doi: 10.6004/jnccn.2015.0151.
59. Yu H, Chen F, Lam KO, Yang L, Wang Y, Jin JY, et al. Potential determinants for radiation-induced lymphopenia in patients with breast cancer using interpretable machine learning approach. *Front Immunol* (2022) 13:768811. doi:10.3389/fimmu.2022.768811.
60. Xie Y, Karki CB, Du D, Haotian L, Wang J, Sobitan A et al. Spike proteins of SARS-CoV and SARS-CoV-2 utilize different mechanisms to bind with human ACE2. *Front Mol Biosci* (2020) 7:591873. doi:10.3389/fmolb.2020.591873.
61. Zhang H, Penninger JM, Li Y, Zhong N, Slutsky AS. Angiotensin-converting enzyme 2 (ACE2) as a SARS-CoV-2 receptor: molecular mechanisms and potential therapeutic target. *Intensive Care Med* (2020) 46(4):586-90. doi: 10.1007/s00134-020-05985-9.
62. Huang Y, Yang C, Xu XF, Xu W, Liu SW. Structural and functional properties of SARS-CoV-2 spike protein: potential antiviral drug development for COVID-19. *Acta Pharmacol Sin* (2020) 41(9):1141-1149. doi: 10.1038/s41401-020-0485-4.
63. Suzuki YJ, Gychka SG. SARS-CoV-2 spike protein elicits cell signaling in human host cells: Implications for possible consequences of COVID-19 vaccines. *Vaccines (Basel)* (2021) 9(1):36. doi: 10.3390/vaccines9010036.
64. Zhang W, Liu HT. MAPK signal pathways in the regulation of cell proliferation in mammalian cells. *Cell Res* (2002) 12:9–18. doi: 10.1038/sj.cr.7290105.
65. Patra T., Meyer K., Geerling L, Isbell TS, Hoft DF, Brien J et al. SARS-CoV-2 spike protein promotes IL-6 trans-signaling by activation of angiotensin II receptor signaling in epithelial cells. *PLoS Pathog* (2020) 16:e1009128. doi: 10.1371/journal.ppat.1009128.
66. Taniguchi K, Karin M. NF-kappaB, inflammation, immunity and cancer: coming of age. *Nat Rev Immunol* (2018) 8(5):309–24. doi: 10.1038/nri.2017.142.
67. De Luca A, Maiello MR, D'Alessio A, Pergameno M, Normanno N. The RAS/RAF/MEK/ERK and the PI3K/AKT signaling pathways: role in cancer pathogenesis and implications for therapeutic approaches. *Expert Opin Ther Targets* (2012) 16 Suppl 2:S17-27. doi:10.1517/14728222.2011.639361.
68. National Cancer Institute (NCI). The Ras initiative. <https://www.cancer.gov/research/key-initiatives/ras> [Accessed April 24, 2022].

69. Braicu C, Buse M, Busuioc C, Drula R, Gulei D, Raduly L, et al. A comprehensive review on MAPK: A promising therapeutic target in cancer. *Cancers (Basel)* (2019) 11(10):1618. doi:10.3390/cancers11101618.
70. Dhillon AS, Hagan S, Rath O, Kolch W. MAP kinase signaling pathways in cancer. *Oncogene* (2007) 26(22):3279-90. doi: 10.1038/sj.onc.1210421.
71. Bryant KL, Stalnecker CA, Zeitouni D, Klomp JE, Peng S, Tikunov AP, et al. Combination of ERK and autophagy inhibition as a treatment approach for pancreatic cancer. *Nature medicine* (2019) 25:628–40. doi:10.1038/s41591-019- 0368-8.
72. Hung AC, Tsai CH, Hou MF, Chang WL, Wang CH, Lee YC et al. The synthetic β -nitrostyrene derivative CYT-Rx20 induces breast cancer cell death and autophagy via ROS-mediated MEK/ERK pathway. *Cancer Letters* (2016) 371:251–61. doi: <https://doi.org/10.1016/j.canlet.2015.11.035>.
73. Kinsey CG, Camolotto SA, Boespflug AM, Guillen KP, Foth M, Truong A, et al. Protective autophagy elicited by RAF→MEK→ERK inhibition suggests a treatment strategy for RAS-driven cancers. *Nat Med* (2019) 25:620–7. doi:10.1038/s41591-019-0367-9.
74. Guo YJ, Pan WW, Liu SB, Shen ZF, Xu Y, Hu LL. ERK/MAPK signaling pathway and tumorigenesis. *Exp Ther Med* (2020) 19(3):1997-2007. doi: 10.3892/etm.2020.8454.
75. Gimple RC, Wang X. RAS: Striking at the core of the oncogenic circuitry. *Front Oncol* (2019) 9:965. doi: 10.3389/fonc.2019.00965.
76. Semesiuk N, Zhylchuk A, Bezdenzhnykh N, Lykhova A, Vorontsova AL, Zhylchuk VE, et al. Disseminated tumor cells and enhanced level of some cytokines in bone marrow and peripheral blood of breast cancer patients as predictive factors of tumor progression. *Exp Oncol* (2013) 35(4):295–302.
77. Meyer F, Samson É, Douville P, Duchesne T, Liu G, Bairati I. Serum prognostic markers in head and neck cancer. *Clin Cancer Res* (2010) 16(3):1008–15. doi: 10.1158/1078-0432.CCR-09-2014.
78. Finkel KA, Warner KA, Kerk S, Bradford CR, McLean SA, Prince ME, et al. IL-6 inhibition with MEDI5117 decreases the fraction of head and neck cancer stem cells and prevents tumor recurrence. *Neoplasia* (2016) 18(5):273–81. doi: 10.1016/j.neo.2016.03.004.
79. Lai SC, Su YT, Chi CC, Kuo YC, Lee KF, Wu YC, et al. DNMT3b/OCT4 expression confers sorafenib resistance and poor prognosis of hepatocellular carcinoma through IL-6/STAT3 regulation. *J Exp Clin Cancer Res* (2019) 38(1):1–18. doi: 10.1186/s13046-019-1442-2.
80. Liao D, Liu Z, Wrasidlo WJ, Luo Y, Nguyen G, Chen T, et al. Targeted therapeutic remodeling of the tumor microenvironment improves an HER-2 DNA vaccine and prevents recurrence in a murine breast cancer model. *Cancer Res* (2011) 71(17):5688–96. doi: 10.1158/0008-5472.CAN-11-1264.
81. Yang J, Nie J, Ma X, Wei Y, Peng Y, Wei X. Targeting PI3K in cancer: mechanisms and advances in clinical trials. *Mol Cancer* (2019) 18(1):26. doi: 10.1186/s12943-019-0954-x.
82. Shi X, Wang J, Lei Y, Cong C, Tan D, Zhou X. Research progress on the PI3K/AKT signaling pathway in gynecological cancer (Review). *Mol Med Rep* (2019) 19(6):4529-4535. doi: 10.3892/mmr.2019.10121.
83. Zhang Q, Lu S, Li T, Yu L, Zhang Y, Zeng H, et al. ACE2 inhibits breast cancer angiogenesis via suppressing the VEGFa/VEGFR2/ERK pathway. *J Exp Clin Cancer Res* (2019) 38(1):173. doi: 10.1186/s13046-019-1156-5.
84. Feng Y, Wan H, Liu J, Zhang R, Ma Q, Han B, et al. The angiotensin-converting enzyme 2 in tumor growth and tumor-associated angiogenesis in non-small cell lung cancer. *Oncol Rep* (2010) 23(4):941–8. doi: 10.3892/or_00000718.
85. Yu C, Tang W, Wang Y, Shen Q, Wang B, Cai C, et al. Downregulation of ACE2/Ang-(1-7)/Mas axis promotes breast cancer metastasis by enhancing store-operated calcium entry. *Cancer Lett* (2016) 376(2):268–77. doi: 10.1016/j.canlet.2016.04.006.

86. Zhang Z, Li L, Li M, Wang X. The SARS-CoV-2 host cell receptor ACE2 correlates positively with immunotherapy response and is a potential protective factor for cancer progression. *Comput Struct Biotechnol J* (2020) 18:2438-44. doi:10.1016/j.csbj.2020.08.024.
87. Röltgen K, Nielsen SCA, Silva O, Younes SF, Zaslavsky M, Costales C, et al. Immune imprinting, breadth of variant recognition, and germinal center response in human SARS-CoV-2 infection and vaccination. *Cell* (2022) 185(6):1025-40.e14. doi: 10.1016/j.cell.2022.01.018.
88. Ogata AF, Cheng CA, Desjardins M, Senussi Y, Sherman AC, Powell M et al. Circulating severe acute respiratory syndrome coronavirus 2 (SARS-CoV-2) vaccine antigen detected in the plasma of mRNA-1273 vaccine recipients. *Clin Inf Dis* (2022) 74(4):715-8. doi: <https://doi.org/10.1093/cid/ciab465>.
89. Cristoni S, Brogna C, Frongillo A, Marino G, Montano L, Piscopo M. Detection of recombinant spike protein in plasma from vaccinated against SARS-CoV-2 individuals. *medRxiv* (2022). doi: <https://doi.org/10.5281/zenodo.5831816>.
90. Ogata AF, Maley AM, Wu C, Gilboa T, Norman M, Lazarovits R, et al. Ultra-sensitive serial profiling of SARS-CoV-2 antigens and antibodies in plasma to understand disease progression in COVID-19 patients with severe disease. *Clin Chem* (2020) 66(12):1562-72. doi: 10.1093/clinchem/hvaa213.
91. El-Zayat SR, Sibaii H, Mannaa FA. Toll-like receptors activation, signaling, and targeting: an overview. *Bull Natl Res Cent* (2019) 43:187. doi: <https://doi.org/10.1186/s42269-019-0227-2>.
92. Karikó K, Buckstein M, Ni H, Weissman D. Suppression of RNA recognition by Toll-like receptors: the impact of nucleoside modification and the evolutionary origin of RNA. *Immunity* (2005) 23(2):165-75. doi: 10.1016/j.immuni.2005.06.008.
93. Freund I, Eigenbrod T, Helm M, Dalpke AH. RNA modifications modulate activation of innate Toll-like receptors. *Genes* (2019) 10(2):92. doi: <https://doi.org/10.3390/genes10020092>.
94. Morais P, Adachi H, Yu YT. The critical contribution of pseudouridine to mRNA COVID-19 vaccines. *Front Cell Dev Biol* (2021) 9:789427. doi:10.3389/fcell.2021.789427.
95. Nance KD, Meier JL. Modifications in an emergency: The role of N1-methylpseudouridine in COVID-19 vaccines. *ACS Cent Sci* (2021) 7(5):748-56. doi:10.1021/acscentsci.1c00197.
96. Heinz FX, Stiasny K. Distinguishing features of current COVID-19 vaccines: knowns and unknowns of antigen presentation and modes of action. *NPJ Vaccines* (2021) 6(1):104. doi:10.1038/s41541-021-00369-6.
97. Uematsu S and Akira S. Toll-like receptors and type I interferons. *J Biol Chem* (2007) 282(21):15319-23. doi: <https://doi.org/10.1074/jbc.R700009200>.
98. Schoggins JW, Wilson SJ, Panis M, Murphy MY, Jones CT, Bieniasz P, et al. A diverse range of gene products are effectors of the type I interferon antiviral response. *Nature* (2011) 472(7344):481-5. doi: 10.1038/nature09907. Erratum in: *Nature* (2015) 525(7567):144.
99. Murira A, Lamarre A. Type-I interferon responses: From friend to foe in the battle against chronic viral infection. *Front Immunol* (2016) 7:609. doi:10.3389/fimmu.2016.00609.
100. Immune Deficiency Foundation (IDF). Innate immune defects (2022). <https://primaryimmune.org/about-primary-immunodeficiencies/specific-disease-types/innate-immune-defects> [Accessed April 28, 2022].
101. Jouanguy E, Béziat V, Mogensen TH, Casanova JL, Tangye SG, Zhang SY. Human inborn errors of immunity to herpes viruses. *Curr Opin Immunol* (2020) 62:106–22. doi:10.1016/j.coi.2020.01.004.
102. Sironi M, Peri AM, Cagliani R, Forni D, Riva S, Biasin M, et al. TLR3 mutations in adult patients with Herpes simplex virus and Varicella-Zoster virus encephalitis. *J Infect Dis* (2017) 215(9):1430-4. doi:10.1093/infdis/jix166.

103. Liang F, Glans H, Enoksson SL, Kolios AGA, Loré K, Nilsson J. Recurrent Herpes-Zoster ophthalmicus in a patient with a novel Toll-like receptor 3 variant linked to compromised activation capacity in fibroblasts. *J Infect Dis* (2020) 221(8):1295-303. doi:10.1093/infdis/jiz229.
104. Katsikas Triantafyllidis K, Giannos P, Mian IT, Kyrtsonis G, Kechagias KS. Varicella zoster virus reactivation following COVID-19 vaccination: a systematic review of case reports. *Vaccines (Basel)* (2021) 9 (9):1013. doi:10.3390/vaccines9091013.
105. Psychogiou M, Samarkos M, Mikos N, Hatzakis A. Reactivation of Varicella zoster virus after vaccination for SARS-CoV-2. *Vaccines (Basel)* (2021) 9(6):572. doi: 10.3390/vaccines9060572.
106. Lladó I, Fernández-Bernáldez A, Rodríguez-Jiménez P. Varicella zoster virus reactivation and mRNA vaccines as a trigger. *JAAD Case Rep* (2021) 15:62-63. doi: 10.1016/j.jdc.2021.07.011.
107. Iwanaga J, Fukuoka H, Fukuoka N, Yutori H, Ibaragi S, Tubbs RS. A narrative review and clinical anatomy of Herpes zoster infection following COVID-19 vaccination. *Clin Anat*. (2021) 35 (1):45-51. doi:10.1002/ca.23790.
108. Préta LH, Contejean A, Salvo F, Treluyer JM, Charlier C, Chouchana L. Association study between herpes zoster reporting and mRNA COVID-19 vaccines (BNT162b2 and mRNA-1273). *Br J Clin Pharmacol* (2022) 88(7):3529-34. doi:10.1111/bcp.15280.
109. Shafiq A, Salameh MA, Laswi I, Mohammed I, Mhaimeed O, Mhaimeed N, et al. Neurological immune-related adverse events after COVID-19 vaccination: A systematic review. *J Clin Pharmacol* (2022) 62(3):291-303. doi: 10.1002/jcph.2017.
110. Ivanova EN, Devlin JC, Buus TB, Koide A, Shwetar J, Cornelius A, et al. B. SARS-CoV-2 mRNA vaccine elicits a potent adaptive immune response in the absence of IFN-mediated inflammation observed in COVID-19. *medRxiv [Preprint]* (2021). doi:10.1101/2021.04.20.21255677 [Accessed April 24, 2023].
111. Lee JS, Shin EC. The type I interferon response in COVID-19: implications for treatment. *Nat Rev Immunol* (2020) 20:585–6. <https://doi.org/10.1038/s41577-020-00429-3>.
112. Hadjadj J, Yatim N, Barnabei L, Corneau A, Boussier J, Smith N, et al. Impaired type I interferon activity and inflammatory responses in severe COVID-19 patients. *Science* (2020) 369:718–24. doi: 10.1126/science.abc6027.
113. Sui Y, Li J, Venzon DJ, Berzofsky JA. SARS-CoV-2 spike protein suppresses ACE2 and Type I interferon expression in primary cells from macaque lung bronchoalveolar lavage. *Front Immunol* (2021) 12:658428. doi:10.3389/fimmu.2021.658428.
114. Thompson D. Could COVID increase your risk for shingles? *HealthDay Reporter* (2022). <https://www.webmd.com/skin-problems-and-treatments/shingles/news/20220411/could-covid-infection-after-age-50-leave-you-vulnerable-to-shingles> [Accessed May 5, 2022].
115. Bhavsar A, Lonnet G, Wang C, Chatzikonstantinidou K, Parikh R, Brabant Y, et al. Increased risk of Herpes Zoster in adults ≥50 years old diagnosed with COVID-19 in the United States. *Open Forum Infect Dis* (2022) 9(5):ofac118. doi:10.1093/ofid/ofac118.
116. Greene TT, Zuniga EI. Type I interferon induction and exhaustion during viral infection: Plasmacytoid dendritic cells and emerging COVID-19 findings. *Viruses* (2021)13(9):1839. doi: 10.3390/v13091839.
117. Braunstein MJ, Kucharczyk J, Adams S. Targeting Toll-like receptors for cancer therapy. *Target Oncol* (2018) 13(5):583-98. doi: 10.1007/s11523-018-0589-7.
118. Aricò E, Castiello L, Capone I, Gabriele L, Belardelli F. Type I interferons and cancer: An evolving story demanding novel clinical applications. *Cancers (Basel)* (2019) 11(12):1943. doi:10.3390/cancers11121943.

119. Dunn GP, Bruce AT, Sheehan KC, Shankaran V, Uppaluri R, Bui JD, et al. A critical function for type I interferons in cancer immunoediting. *Nat Immunol* (2005) 6(7):722-9. doi: 10.1038/ni1213.
120. Critchley-Thorne RJ, Simons DL, Yan N, Miyahira AK, Dirbas FM, Johnson DL, et al. Impaired interferon signaling is a common immune defect in human cancer. *Proc Natl Acad Sci USA* (2009) 106(22):9010-5. doi: 10.1073/pnas.0901329106.
121. Zeinalzadeh E, Valerievich Yumashev A, Rahman HS, Marofi F, Shomali N, Kafil HS, et al. The role of Janus Kinase/STAT3 pathway in hematologic malignancies with an emphasis on epigenetics. *Front Genet* (2021) 12:703883. doi:10.3389/fgene.2021.703883.
122. Cao X, Liang Y, Hu Z, Li H, Yang J, Hsu EJ, et al. Next generation of tumor-activating type I IFN enhances anti-tumor immune responses to overcome therapy resistance. *Nat Commun* (2021) 12(1):5866. doi: <https://doi.org/10.1038/s41467-021-26112-2>.
123. Yu R, Zhu B, Chen D. Type I interferon-mediated tumor immunity and its role in immunotherapy. *Cell Mol Life Sci* (2022) 79:191. doi: <https://doi.org/10.1007/s00018-022-04219-z>.
124. Gargan S, Ahmed S, Mahony R, Bannan C, Napoletano S, O'Farrelly C, Borrow P, Bergin C, Stevenson NJ. HIV-1 promotes the degradation of components of the Type 1 IFN JAK/STAT pathway and blocks anti-viral ISG induction. *EBioMedicine* (2018) 30:203-216. doi: 10.1016/j.ebiom.2018.03.006.
125. Sandstrom TS, Ranganath N, Angel JB. Impairment of the type I interferon response by HIV-1: Potential targets for HIV eradication. *Cytokine Growth Factor Rev* (2017) 37:1-16. doi: 10.1016/j.cytogfr.2017.04.004.
126. Reder AT, Feng X. Aberrant type I interferon regulation in autoimmunity: Opposite directions in MS and SLE, shaped by evolution and body ecology. *Front Immunol* (2013) 4:281. doi: 10.3389/fimmu.2013.00281.
127. Harari D, Abramovich R, Kallweit N, Pouly S, Zozulya-Weidenfeller A, Smith P et al. 113: Type I interferon signaling is suppressed in experimental autoimmune encephalomyelitis (EAE): Implications for multiple sclerosis. *Cytokine* (2013) 63 (3):269-270. doi: <https://doi.org/10.1016/j.cyto.2013.06.116>.
128. Bosco-Lévy P, Foch C, Grelaud A, Sabidó M, Lacueille C, Jové J, et al. Incidence and risk of cancer among multiple sclerosis patients: A matched population-based cohort study. *Eur J Neurol* (2022) 29(4):1091-1099. doi: 10.1111/ene.15226.
129. Cosentino M, Marino F. The spike hypothesis in vaccine-induced adverse effects: questions and answers. *Trends Mol Med* (2022) 28(10):797-799. doi:10.1016/j.molmed.2022.07.009.
130. European Medicines Agency. Assessment report. Comirnaty. Common name: COVID-19 mRNA vaccine (nucleoside-modified). Procedure No. EMEA/H/C/005735/0000. E MA/707383/2020 Corr.1*1 Committee for Medicinal Products for Human Use (2021). https://www.ema.europa.eu/en/documents/assessment-report/comirnaty-epar-public-assessment-report_en.pdf [Accessed April 11, 2022].
131. Non-clinical overview. FDA-CBER-2021-5683-0013861 (2021). https://phmpt.org/wp-content/uploads/2022/03/125742_S1_M2_24_nonclinical-overview.pdf [Accessed April 2, 2022].
132. Pfz SARS-CoV-2 mRNA Vaccine (BNT162, PF-07302048) (2022). <https://www.docdroid.net/xq0Z8B0/pfizer-report-japanese-government-pdf#page=17> [Accessed April 11, 2022].
133. Zhouyi R, Hongcheng M, Saketh K, Puellas VG, Czogalla J, Schadler J et al. SARS-CoV-2 spike protein accumulation in the skull-meninges-brain axis: Potential implications for

- long-term neurological complications in post-COVID-19. *bioRxiv* [Preprint] (2023). doi: <https://doi.org/10.1101/2023.04.04.535604> [Accessed April 22, 2023].
134. Kowarz E, Krutzke L, Klp M, Streb P, Larghero P, Reis J et al. Vaccine-induced COVID-19 mimicry syndrome. *Elife* (2022) 11:e74974. doi:10.7554/eLife.74974.
135. Xia X. Detailed dissection and critical evaluation of the Pfizer/BioNTech and Moderna mRNA vaccines. *Vaccines* (Basel) (2021) 9(7):734. doi:10.3390/vaccines9070734.
136. Newman ZR, Young JM, Ingolia NT, Barton GM. Differences in codon bias and GC content contribute to the balanced expression of TLR7 and TLR9. *Proc Natl Acad Sci USA* (2016) 113(10):E1362-71. doi: 10.1073/pnas.1518976113.
137. Mauro VP, Chappell SA. A critical analysis of codon optimization in human therapeutics. *Trends Mol Med* (2014) 20(11):604-13. doi:10.1016/j.molmed.2014.09.003.
138. McKernan K., Kyriakopoulos A.M., McCullough P.A. Differences in vaccine and SARS-CoV-2 replication derived mRNA: implications for cell biology and future disease. *OSF Prepr* [Preprint] (2021). doi: 10.31219/osf.io/bcsa6 [Accessed April 24, 2023].
139. Xia X. Detailed dissection and critical evaluation of the Pfizer/BioNTech and Moderna mRNA vaccines. *Vaccines* (Basel) (2021) 9(7) .doi: 10.3390/vaccines9070734.
140. Svidritskiy E, Madireddy R, Korostelev AA. Structural basis for translation termination on a pseudouridylated stop codon. *J Mol Biol* (2016) 428(10 Pt B):2228-36. doi: 10.1016/j.jmb.2016.04.018.
141. Eyler DE, Franco MK, Batool Z, Wu MZ, Dubuke ML, Dobosz-Bartoszek M et al. Pseudouridinylation of mRNA coding sequences alters translation. *Proc Natl Acad Sci U S A* (2019) 116(46):23068-23074. doi: 10.1073/pnas.
142. Hagen T, Laski A, Brmmer A, Pruka A, Schlsser V, Clry A et al. Inosine substitutions in RNA activate latent G-quadruplexes. *J Am Chem Soc* (2021) 143(37):15120-15130. doi: 10.1021/jacs.1c05214.
143. Jara-Espejo M, Fleming AM, Burrows CJ. Potential G-quadruplex forming sequences and N(6)-methyladenosine colocalize at human pre-mRNA intron splice sites. *ACS Chem Biol* 2020 15(6):1292-300. doi: 10.1021/acscchembio.0c00260.
144. Quante T, Otto B, Brazdova M, et al. Mutant p53 is a transcriptional co-factor that binds to G-rich regulatory regions of active genes and generates transcriptional plasticity. *Cell Cycle* (2012) 11(17):3290-303. doi: 10.4161/cc.21646.
145. Lago S, Nadai M, Ruggiero E, et al. The MDM2 inducible promoter folds into four-tetrad antiparallel G-quadruplexes targetable to fight malignant liposarcoma. *Nucleic Acids Res* (2021) 49(2):847-63. doi: 10.1093/nar/gkaa1273.
146. Ozaki T, Nakagawara A. Role of p53 in cell death and human cancers. *Cancers* (Basel) (2011) 3(1):994-1013. doi: 10.3390/cancers3010994.
147. Fay MM, Lyons SM, Ivanov P. RNA G-quadruplexes in biology: Principles and molecular mechanisms. *J Mol Biol* (2017) 429(14):2127-2147. doi: 10.1016/j.jmb.2017.05.017.
148. Hou X, Zaks T, Langer R, Dong Y. Lipid nanoparticles for mRNA delivery. *Nat Rev Mater* (2021) 6:1078–94. doi: <https://doi.org/10.1038/s41578-021-00358-0>.
149. Schoenmaker L, Witzigmann D, Kulkarni JA, Verbeke R, Kersten G, Jiskoot W, et al. mRNA-lipid nanoparticle COVID-19 vaccines: Structure and stability. *Int J Pharm* (2021) 601:120586. doi:10.1016/j.ijpharm.2021.120586.
150. Kedmi R, Ben-Arie N, Peer D. The systemic toxicity of positively charged lipid nanoparticles and the role of Toll-like receptor 4 in immune activation. *Biomaterials* (2010) 31(26):6867-75. doi: 10.1016/j.biomaterials.2010.05.027.
151. Ndeupen S, Qin Z, Jacobsen S, Bouteau A, Estanbouli H, Igyrt BZ. The mRNA-LNP platform's lipid nanoparticle component used in preclinical vaccine studies is highly inflammatory. *iScience* (2021) 24 (12):103479. doi: 10.1016/j.isci.2021.103479.

152. Moghimi SM, Simberg D. Pro-inflammatory concerns with lipid nanoparticles. *Mol Ther* (2022) 30(6):2109-2110. doi: 10.1016/j.ymthe.2022.04.011.
153. Sakurai F, Tachibana M, Mizuguchi H. Adenovirus vector-based vaccine for infectious diseases. *Drug Metab Pharmacokinet* (2022) 42:100432. doi: 10.1016/j.dmpk.2021.100432.
154. Preskorn SH. The 5% of the population at high risk for severe COVID-19 infection is identifiable and needs to be taken into account when reopening the economy. *J Psychiatr Pract* (2020) 26(3):219-227. doi: 10.1097/PRA.0000000000000475.
155. Ye Q, Wang B, Mao J. The pathogenesis and treatment of the 'Cytokine Storm' in COVID-19. *J Infect* (2020) 80(6):607-613. doi: 10.1016/j.jinf.2020.03.037.
156. Gupta A, Madhavan MV, Sehgal K, Nair N, Mahajan S, Sehrawat TS, et al. Extrapulmonary manifestations of COVID-19. *Nat Med* (2020) 26(7):1017-1032. doi: 10.1038/s41591-020-0968-3.
157. Chen G, Wu D, Guo W, Cao Y, Huang D, Wang H, et al. Clinical and immunological features of severe and moderate coronavirus disease 2019. *J Clin Invest* (2020) 130(5):2620-2629. doi:10.1172/JCI137244.
158. Singh N, Baby D, Rajguru JP, Patil PB, Thakkannavar SS, Pujari VB. Inflammation and cancer. *Ann Afr Med* (2019) 18(3):121-126. doi:10.4103/aam.aam_56_18.
159. Coussens LM, Werb Z. Inflammation and cancer. *Nature* (2002) 420(6917):860-7. doi: 10.1038/nature01322.
160. Mantovani A, Allavena P, Sica A, Balkwill F. Cancer-related inflammation. *Nature* (2008) 454(7203):436-44. doi: 10.1038/nature07205.
161. de Martel C, Franceschi S. Infections and cancer: established associations and new hypotheses. *Crit Rev Oncol Hematol* (2009) 70(3):183-94. doi: 10.1016/j.critrevonc.2008.07.021.
162. Ji Z, He L, Regev A, Struhl K. Inflammatory regulatory network mediated by the joint action of NF- κ B, STAT3, and AP-1 factors is involved in many human cancers. *Proc Natl Acad Sci USA* (2019) 116(19):9453-62. doi: 10.1073/pnas.1821068116.
163. Grivennikov SI, Greten FR, Karin M. Immunity, inflammation, and cancer. *Cell* (2010) 140(6):883-99. doi: 10.1016/j.cell.2010.01.025.
164. Mantovani A, Ponzetta A, Inforzato A, Jaillon S. Innate immunity, inflammation and tumor progression: double-edged swords. *J Intern Med* (2019) 285(5):524-532. doi: 10.1111/joim.12886.
165. Ocana A, Nieto-Jimenez C, Pandiella A, Templeton AJ. Neutrophils in cancer: prognostic role and therapeutic strategies. *Mol Cancer* (2017) 16(1):137. doi:10.1186/s12943-017-0707-7.
166. Granot Z. Neutrophils as a therapeutic target in cancer. *Front Immunol* (2019) 10:1710. doi:10.3389/fimmu.2019.01710.
167. Cold Spring Harbor Laboratory (CSH). How a sleeping cancer awakens and metastasizes. (2018). doi: <https://www.cshl.edu/how-a-sleeping-cancer-awakens-and-metastasizes/> [Accessed April 16, 2022].
168. Wu L, Saxena S, Awaji M, Singh RK. Tumor-associated neutrophils in cancer: Going pro. *Cancers (Basel)* (2019) 11(4):564. doi:10.3390/cancers11040564.
169. Leach J, Morton JP, Sansom OJ. Neutrophils: Homing in on the myeloid mechanisms of metastasis. *Mol Immunol* (2019) 110:69-76. doi:10.1016/j.molimm.2017.12.013.
170. Rapoport BL, Steel HC, Theron AJ, Smit T, Anderson R. Role of the neutrophil in the pathogenesis of advanced cancer and impaired responsiveness to therapy. *Molecules* (2020) 25(7):1618. doi:10.3390/molecules25071618.
171. Papayannopoulos, V. Neutrophil extracellular traps in immunity and disease. *Nat Rev Immunol* (2018) 18:134-47. doi: 10.1038/nri.2017.105.

172. Demkow U. Neutrophil extracellular traps (NETs) in cancer invasion, evasion and metastasis. *Cancers (Basel)* (2021) 13(17):4495. doi: 10.3390/cancers13174495.
173. Masucci MT, Minopoli M, Del Vecchio S, Carriero MV. The emerging role of neutrophil extracellular traps (NETs) in tumor progression and metastasis. *Front Immunol* (2020) 11:1749. doi: 10.3389/fimmu.2020.01749.
174. Parhiz H, Brenner JS, Patel PN, Papp TE, Shahnawaz H, Li Q, Shi R, et al. Added to pre-existing inflammation, mRNA-lipid nanoparticles induce inflammation exacerbation (IE). *J Control Release* (2022) 344:50-61. doi:10.1016/j.jconrel.2021.12.027.
175. Alavi M, Hamidi M. Passive and active targeting in cancer therapy by liposomes and lipid nanoparticles. *Drug Metab Pers Ther* (2019) 34(1). doi: 10.1515/dmpt-2018-0032.
176. Bennie LA, McCarthy HO, Coulter JA. Enhanced nanoparticle delivery exploiting tumor-responsive formulations. *Cancer Nanotechnol* (2018) 9(1):10. doi: 10.1186/s12645-018-0044-6.
177. Maeda H. The enhanced permeability and retention (EPR) effect in tumor vasculature: the key role of tumor-selective macromolecular drug targeting. *Adv Enzyme Regul* (2001) 41:189-207. doi: 10.1016/s0065-2571(00)00013-3.
178. Tenchov R. Lipid nanoparticles - key players in cancer treatment. CAS. A division of the American Chemical Society (2022) <https://www.cas.org/resources/blog/lipid-nanoparticles-cancer-therapy> [Accessed August 25, 2022].
179. Hwang TL, Aljuffali IA, Hung CF, Chen CH, Fang JY. The impact of cationic solid lipid nanoparticles on human neutrophil activation and formation of neutrophil extracellular traps (NETs). *Chem Biol Interact* (2015) 235:106-14. doi: 10.1016/j.cbi.2015.04.011.
180. Acevedo-Whitehouse K, Bruno R. Potential health risks of mRNA-based vaccine therapy: a hypothesis. *Med Hypotheses* (2023) 171:111015. doi: 10.1016/j.mehy.2023.111015.
181. Zhang L, Richards A, Barrasa MI, Hughes SH, Young RA, Jaenisch R. Reverse transcribed SARS-CoV-2 RNA can integrate into the genome of cultured human cells and can be expressed in patient-derived tissues. *Proc Natl Acad Sci USA* (2021) 118. doi: <https://doi.org/10.1073/pnas.2105968118>.
182. Aldén M, Olofsson Falla F, Yang D, Barghouth M, Luan C, Rasmussen M, et al. Intracellular reverse transcription of Pfizer BioNTech COVID-19 mRNA vaccine BNT162b2 in vitro in human liver cell line. *Curr Issues Mol Biol* (2022) 44:1115-26. doi: <https://doi.org/10.3390/cimb44030073>.
183. Zhang X, Zhang R, Yu J. New understanding of the relevant role of LINE-1 retrotransposition in human disease and immune modulation. *Front Cell Dev Biol* (2020) 8:657. doi: 10.3389/fcell.2020.00657.
184. McKerrow W, Wang X, Mendez-Dorantes C, Mita P, Cao S, Grivainis M, et al. LINE-1 expression in cancer correlates with p53 mutation, copy number alteration, and S phase checkpoint. *Proc Natl Acad Sci USA* (2022) 119(8):e2115999119. doi:10.1073/pnas.2115999119.
185. Rangasamy D, Lenka N, Ohms S, Dahlstrom JE, Blackburn AC, Board PG. Activation of LINE-1 retrotransposon increases the risk of epithelial-mesenchymal transition and metastasis in epithelial cancer. *Curr Mol Med* (2015) 15(7):588-97. doi: 10.2174/1566524015666150831130827.
186. Cammas A, Millevoi S. RNA G-quadruplexes: emerging mechanisms in disease. *Nucleic Acids Res* (2017) 45(4):1584-95. doi: 10.1093/nar/gkw1280.
187. Kosiol N, Juranek S, Brossart P, Heine A, Paeschke K. G-quadruplexes: a promising target for cancer therapy. *Mol Cancer* (2021) 20(1):40. doi:10.1186/s12943-021-01328-4.
188. Katapadi VK, Nambiar M, Raghavan SC. Potential G-quadruplex formation at breakpoint regions of chromosomal translocations in cancer may explain their fragility. *Genomics* (2012) 100(2):72-80. doi: 10.1016/j.ygeno.2012.05.008.

189. Hänsel-Hertsch R, Simeone A, Shea A, Hui WWI, Zyner KG, Marsico G, et al. Landscape of G-quadruplex DNA structural regions in breast cancer. *Nat Genet* (2020) 52(9):878–83. doi: <https://doi.org/10.1038/s41588-020-0672-8>.
190. Singh N, Bharara Singh A. S2 subunit of SARS-nCoV-2 interacts with tumor suppressor protein p53 and BRCA: an *in silico* study. *Transl Oncol* (2020) 13(10):100814. doi:10.1016/j.tranon.2020.100814.
191. Mantovani F, Collavin L, Del Sal G. Mutant p53 as a guardian of the cancer cell. *Cell Death Differ* (2019) 26(2):199–212. doi: 10.1038/s41418-018-0246-9.
192. Brosh R, Rotter V. When mutants gain new powers: news from the mutant p53 field. *Nat Rev Cancer* (2009) 9:701–13.
193. Venkitaraman AR. Cancer suppression by the chromosome custodians, BRCA1 and BRCA2. *Science* (2014) 343(6178):1470–5.
194. Green DR, Kroemer G. Cytoplasmic functions of the tumour suppressor p53. *Nature* (2009) 458(7242):1127–30. doi: 10.1038/nature07986.
195. Henderson BR. Regulation of BRCA1, BRCA2 and BARD1 intracellular trafficking. *Bioessays* (2005) 27(9):884–93. doi: 10.1002/bies.20277.
196. Moll UM, Ostermeyer AG, Haladay R, Winkfield B, Frazier M, Zambetti G. Cytoplasmic sequestration of wild-type p53 protein impairs the G1 checkpoint after DNA damage. *Mol Cell Biol* (1996) 16(3):1126–37. doi: 10.1128/MCB.16.3.1126.
197. Rodriguez JA, Au WW, Henderson BR. Cytoplasmic mislocalization of BRCA1 caused by cancer-associated mutations in the BRCT domain. *Exp Cell Res* (2004) 293(1):14–21. doi: 10.1016/j.yexcr.2003.09.027.
198. Santivasi WL, Wang H, Wang T, Yang Q, Mo X, Brogi E, et al. Association between cytosolic expression of BRCA1 and metastatic risk in breast cancer. *Br J Cancer* (2015) 113(3):453–9. doi:10.1038/bjc.2015.208.
199. Rakha EA, El-Sheikh SE, Kandil MA, El-Sayed ME, Green AR, Ellis IO. Expression of BRCA1 protein in breast cancer and its prognostic significance. *Hum Pathol* (2008) 39(6):857–65. doi: 10.1016/j.humpath.2007.10.011.
200. Petitjean A, Achatz MI, Borresen-Dale AL, Hainaut P, Olivier M. TP53 mutations in human cancers: functional selection and impact on cancer prognosis and outcomes. *Oncogene* (2007) 26(15):2157–65. doi: 10.1038/sj.onc.1210302.
201. National Cancer Institute (NCI). BRCA gene mutations: Cancer risk and genetic testing (2020). <https://www.cancer.gov/about-cancer/causes-prevention/genetics/brca-fact-sheet> [Accessed April 26, 2022].
202. Facing Our Risk of Cancer Empowered (FORCE). Cancer risk associated with inherited BRCA1 gene mutations (2022). <https://www.facingourrisk.org/info/hereditary-cancer-and-genetic-testing/hereditary-cancer-genes-and-risk/genes-by-name/brca1/cancer-risk> [Accessed April 15, 2022].
203. Mersch J, Jackson MA, Park M, Nebgen D, Peterson SK, Singletary C, et al. Cancers associated with BRCA1 and BRCA2 mutations other than breast and ovarian. *Cancer* (2015) 121(2):269–75. doi: 10.1002/cncr.29041. Erratum in: *Cancer* (2015) 121(14):2474–5.
204. Abate G, Frisoni GB, Bourdon JC, Piccirella S, Memo M, Uberti D. The pleiotropic role of p53 in functional/dysfunctional neurons: focus on pathogenesis and diagnosis of Alzheimer's disease. *Alzheimers Res Ther* (2020) 12(1):160. doi: 10.1186/s13195-020-00732-0.
205. Szybińska A, Leśniak W. P53 dysfunction in neurodegenerative diseases. The cause or effect of pathological changes? *Aging Dis* (2017) 8(4):506–518. doi: 10.14336/AD.2016.1120.
206. Nakamura M, Kaneko S, Dickson DW, Kusaka H. Aberrant accumulation of BRCA1 in Alzheimer disease and other tauopathies. *J Neuropathol Exp Neurol* (2020) 79(1):22–33. doi: 10.1093/jnen/nlz107.

207. Columbia University. Irving Medical Center. Small study finds Alzheimer's-like changes in some COVID patients' brains (2022). <https://www.cuimc.columbia.edu/news/small-study-finds-alzheimers-changes-some-covid-patients-brains> [Accessed May 7, 2022].
208. Alzheimer's Association. International brain study: SARS-CoV-2 impact on behavior and cognition (2022). https://www.alz.org/research/for_researchers/partnerships/sars-cov2-global-brain-study [Accessed May 8, 2022].
209. Wang K, Chen W, Zhang Z, Deng Y, Lian JQ, Du P et al. CD147-spike protein is a novel route for SARS-CoV-2 infection to host cells. *Signal Transduct Target Ther* (2020) 5(1):283. doi: 10.1038/s41392-020-00426-x.
210. Behl T, Kaur I, Aleya L, Sehgal A, Singh S, Sharma N, et al. CD147-spike protein interaction in COVID-19: Get the ball rolling with a novel receptor and therapeutic target. *Sci Total Environ* (2022) 808:152072. doi:10.1016/j.scitotenv.2021.152072.
211. Shilts J, Crozier TWM, Greenwood EJD, Lehner PJ, Wright GJ. No evidence for basigin/CD147 as a direct SARS-CoV-2 spike binding receptor. *Sci Rep* (2021) 11(1):413. doi: 10.1038/s41598-020-80464-1.
212. Xu T, Zhou M, Peng L, Kong S, Miao R, Shi Y, et al. Upregulation of CD147 promotes cell invasion, epithelial-to-mesenchymal transition and activates MAPK/ERK signaling pathway in colorectal cancer. *Int J Clin Exp Pathol* (2014) 7(11):7432-41.
213. Yang H, Chen B. CD147 in ovarian and other cancers. *Int J Gynecol Cancer* (2013) 23(1):2-8. doi: 10.1097/IGC.0b013e3182749139.
214. Peng J, Jiang H, Guo J, Huang J, Yuan Q, Xie J et al. CD147 expression is associated with tumor proliferation in bladder cancer via GSDMD. *Biomed Res Int* (2020) 2020:7638975. doi: 10.1155/2020/7638975.
215. Landras A, Reger de Moura C, Jouenne F, Lebbe C, Menashi S, Mourah S. CD147 is a promising target of tumor progression and a prognostic biomarker. *Cancers (Basel)* (2019) 11(11):1803. doi:10.3390/cancers11111803.
216. Centers for Disease Control and Prevention (CDC). Trends in numbers of Covid-19 cases and deaths in the US reported to CDC, by state/territory. Covid data tracker (2023). https://covid.cdc.gov/covid-data-tracker/#trends_weeklydeaths_select_00 [Accessed Feb 10, 2023].
217. UK Health Security Agency. Coronavirus (COVID-19) in the UK. Simple summary for England. https://coronavirus.data.gov.uk/easy_read [Accessed Feb 10, 2023].
218. Improving detection of and response to adverse events. In: Institute of Medicine (US) Board on Health Promotion and Disease Prevention. Vaccine Safety Forum: Summaries of Two Workshops. Washington (DC): National Academies Press (US) (1997). <https://www.ncbi.nlm.nih.gov/books/NBK232983/> [Accessed May 8, 2022].
219. Agha M, Blake M, Chilleo C, Wells A, Haidar G. Suboptimal response to COVID-19 mRNA vaccines in hematologic malignancies patients. *medRxiv [Preprint]* (2021). doi:10.1101/2021.04.06.21254949 [Accessed Feb 10, 2023].
220. Maneikis K, Šablauskas K, Ringelevičiūtė U, Vaitekėnaitė V, Čekauskienė R, Kryžauskaitė L, et al. Immunogenicity of the BNT162b2 COVID-19 mRNA vaccine and early clinical outcomes in patients with haematological malignancies in Lithuania: a national prospective cohort study. *Lancet Haematol* (2021) 8(8):e583-e592. doi: 10.1016/S2352-3026(21)00169-1.
221. Shree T, Shankar V, Lohmeyer JJK, Czerwinski DK, Schroers-Martin JG, Rodriguez GM et al. CD20-targeted therapy ablates de novo antibody response to vaccination but spares preestablished immunity. *Blood Cancer Discov* (2022) 3(2):95-102. doi: 10.1158/2643-3230.BCD-21-0222.

222. Pich O, Muiños F, Lolkema MP, Steeghs N, Gonzalez-Perez A, Lopez-Bigas N. The mutational footprints of cancer therapies. *Nat Genet* (2019) 51(12):1732-40. doi:10.1038/s41588-019-0525-5.
223. Korompoki E, Gavriatopoulou M, Kontoyiannis DP. COVID-19 vaccines in patients with cancer. A welcome addition, but there is need for optimization. *JAMA Oncol* (2021) 7(8):1113-4. doi:10.1001/jamaoncol.2021.1218.
224. Waissengrin B, Agbarya A, Safadi E, Padova H, Wolf I. Short-term safety of the BNT162b2 mRNA COVID-19 vaccine in patients with cancer treated with immune checkpoint inhibitors. *Lancet Oncol* (2021) 2045:581–3. doi: [https://doi.org/10.1016/S1470-2045\(21\)00155-8](https://doi.org/10.1016/S1470-2045(21)00155-8).
225. Garreffa E, Hamad A, O'Sullivan CC, Hazim AZ, York J, Puri S, et al. Regional lymphadenopathy following COVID-19 vaccination: Literature review and considerations for patient management in breast cancer care. *Eur J Cancer* (2021) 159:38-51. doi: 10.1016/j.ejca.2021.09.033.
226. Wolfson S, Kim E. Breast cancer screening and axillary adenopathy in the era of COVID-19 vaccination. *Radiology* (2023) 306(2):e222040. doi: 10.1148/radiol.222040.
227. Seban RD, Richard C, Nascimento-Leite C, Ghidaglia J, Provost C, Gonin J, et al. Absolute lymphocyte count after COVID-19 vaccination is associated with vaccine-induced hypermetabolic lymph nodes on ¹⁸F-FDG PET/CT: A focus in breast cancer care. *J Nucl Med* (2022) 63(8):1231-1238. doi: 10.2967/jnumed.121.263082.
228. Özütemiz C, Potter DA, Özütemiz AÖ, Steinberger D. Lymphadenopathy after the third Covid-19 vaccine. *Curr Probl Cancer Case Rep* (2021) 4:100127. doi: 10.1016/j.cpcr.2021.100127.
229. Treglia G, Cuzzocrea M, Giovanella L, Elzi L, Muoio B. Prevalence and significance of hypermetabolic lymph nodes detected by 2-[¹⁸F]FDG PET/CT after COVID-19 vaccination: A systematic review and a meta-analysis. *Pharmaceuticals (Basel)* (2021) 14(8):762. doi: 10.3390/ph14080762.
230. Meo C, Palma G, Bruzzese F, Budillon A, Napoli C, de Nigris F. Spontaneous cancer remission after COVID-19: insights from the pandemic and their relevance for cancer treatment. *J Transl Med* (2023) 21(1):273. doi: 10.1186/s12967-023-04110-w.
231. Sousa LG, McGrail DJ, Li K, Marques-Piubelli ML, Gonzalez C, Dai H et al. Spontaneous tumor regression following COVID-19 vaccination. *J Immunother Cancer* (2022) 10(3):e004371. doi: 10.1136/jitc-2021-004371.
232. Krzyszczyk P, Acevedo A, Davidoff EJ, Timmins LM, Marrero-Berrios I, Patel M et al. The growing role of precision and personalized medicine for cancer treatment. *Technology (Singap World Sci)* (2018) 6(3-4):79-100. doi: 10.1142/S2339547818300020.
233. American Association for Cancer Research (AACR). Honey K. Complexities of cancer explained for patient benefit (2015). <https://www.aacr.org/blog/2015/05/26/complexities-of-cancer-explained-for-patient-benefit/> [Accessed April 20, 2022].
234. International Agency for Research of Cancer - World Health Organization (WHO). Cancer over time (1965-2023). <https://gco.iarc.fr/overtime/en> [Accessed April 22, 2023].
235. Australian Institute of Health and Welfare (AIHW). Australian Government. Cancer (2023). <https://www.aihw.gov.au/reports-data/health-conditions-disability-deaths/cancer/data> [Accessed April 22, 2023].
236. Cancer statistics in Japan. Table download (2022). https://ganjoho.jp/reg_stat/statistics/data/dl/en.html#anchor2 [Accessed April 22, 2023].
237. Office for National Statistics. Cancer Registration Statistics, England. (2019). <https://www.ons.gov.uk/peoplepopulationandcommunity/healthandsocialcare/conditionsanddiseases>

- [ses/datasets/cancerregistrationstatistics/cancerregistrationstatisticsengland](#) [Accessed April 22, 2023].
238. Canadian Cancer Society. Canadian Cancer Statistics (2023). <https://cancer.ca/en/research/cancer-statistics/canadian-cancer-statistics> [Accessed April 22, 2023].
 239. European Commission. European Cancer Information System (ECIS) [https://ecis.jrc.ec.europa.eu/explorer.php?\\$0-0\\$1-All\\$2-All\\$4-1,2\\$3-0\\$6-0,85\\$5-2020,2020\\$7-7\\$CEstByCountry\\$X0_8-3\\$X0_19-AE27\\$X0_20-No\\$CEstBySexByCountry\\$X1_8-3\\$X1_19-AE27\\$X1_-1-1\\$CEstByIndiByCountry\\$X2_8-3\\$X2_19-AE27\\$X2_20-No\\$CEstRelative\\$X3_8-3\\$X3_9-AE27\\$X3_19-AE27\\$CEstByCountryTable\\$X4_19-AE27](https://ecis.jrc.ec.europa.eu/explorer.php?$0-0$1-All$2-All$4-1,2$3-0$6-0,85$5-2020,2020$7-7$CEstByCountry$X0_8-3$X0_19-AE27$X0_20-No$CEstBySexByCountry$X1_8-3$X1_19-AE27$X1_-1-1$CEstByIndiByCountry$X2_8-3$X2_19-AE27$X2_20-No$CEstRelative$X3_8-3$X3_9-AE27$X3_19-AE27$CEstByCountryTable$X4_19-AE27) [Accessed April 22, 2023].
 240. Henley SJ, Dowling NF, Ahmad FB, Ellington TD, Wu M, Richardson LC. Covid-19 and other underlying causes of cancer deaths – United States, January 2018-July 2022. *MMWR Morb Mortal Wkly Rep* (2022) 71:1583-8. doi: <http://dx.doi.org/10.15585/mmwr.mm7150a3>.
 241. Schuppener LM, Olson K, Brooks EG. Death certification: errors and interventions. *Clin Med Res* (2020) 18(1):21-6. doi: 10.3121/cmr.2019.1496.
 242. American Cancer Society. Covid-19 vaccines in people with cancer (2022). <https://www.cancer.org/treatment/treatments-and-side-effects/physical-side-effects/low-blood-counts/infections/covid-19-vaccines-in-people-with-cancer.html> [Accessed September 6, 2022].
 243. National Cancer Institute (NCI). Covid-19 vaccines and people with cancer: A Q&A with Dr. Steven Pergam (2022) <https://www.cancer.gov/about-cancer/coronavirus/covid-19-vaccines-people-with-cancer> [Accessed September 6, 2022].
 244. Hwang JK, Zhang T, Wang AZ, Li Z. COVID-19 vaccines for patients with cancer: benefits likely outweigh risks. *J Hematol Oncol* (2021) 14(1):38. doi: 10.1186/s13045-021-01046-w.
 245. Goldman JD, Gonzalez MA, Rütthrich MM, Sharon E, von Lilienfeld-Toal M. COVID-19 and cancer: Special considerations for patients receiving immunotherapy and immunosuppressive cancer therapies. *Am Soc Clin Oncol Educ Book* (2022) 42:1-13. doi: 10.1200/EDBK_359656.
 246. Hassan AO, Case JB, Winkler ES, Thackray LB, Kafai NM, Bailey AL, et al. A SARS-CoV-2 infection model in mice demonstrates protection by neutralizing antibodies. *Cell* (2020) 182(3):744-53.e4. doi: 10.1016/j.cell.2020.06.011.
 247. Halma MTJ, Rose J, Lawrie T. The novelty of mRNA viral vaccines and potential harms: A scoping review. *J* (2023) 6(2):220-235. doi: <https://doi.org/10.3390/j6020017>.
 248. Zhao J, Li K, Wohlford-Lenane C, Agnihothram SS, Fett C, Zhao J, Gale MJ Jr, et al. Rapid generation of a mouse model for Middle East respiratory syndrome. *Proc Natl Acad Sci USA* (2014) 111(13):4970-5. doi: 10.1073/pnas.1323279111.
 249. Rathnasinghe R, Strohmeier S, Amanat F, Gillespie VL, Krammer F, García-Sastre A, et al. Comparison of transgenic and adenovirus hACE2 mouse models for SARS-CoV-2 infection. *bioRxiv [Preprint]* (2020). <https://doi.org/10.1101/2020.07.06.190066> [Accessed Feb 10, 2023].
 250. Lamprecht Tratar U, Horvat S, Cemazar M. Transgenic mouse models in cancer research. *Front Oncol* (2018) 8:268. doi: 10.3389/fonc.2018.00268.20.
 251. Trivanović D, Peršurić Ž, Agaj A, Jakopović M, Samaržija M, Bitar L et al. The interplay of lung cancer, COVID-19, and vaccines. *Int J Mol Sci* (2022) 23(23):15067. doi: 10.3390/ijms232315067.
 252. Carnell GW, Ciazynska KA, Wells DA, Xiong X, Aguinam ET, McLaughlin SH, et al. SARS-CoV-2 spike protein stabilized in the closed state induces potent neutralizing responses. *J Virol* (2021) 95(15):e0020321. doi: 10.1128/JVI.00203-21.

253. Wu KJ. Covid-19 vaccine makers are looking beyond the spike protein. The Atlantic (2021). <https://www.theatlantic.com/science/archive/2021/05/spike-protein-vaccines-covid/618954/> [Accessed April 26, 2023].
254. SARS-CoV-2 vaccine (COH04S1) versus emergency use authorization SARS-CoV-2 vaccine for the treatment of COVID-19 in patients with blood cancer. ClinicalTRials.gov. <https://clinicaltrials.gov/ct2/show/NCT04977024> [Accessed April 26, 2023].
255. Liu Y, Yu Q, Wen H, Shi F, Wang F, Zhao Y et al. What matters: non-pharmaceutical interventions for COVID-19 in Europe. Antimicrob Resist Infect Control (2022) 11(1):3. doi: 10.1186/s13756-021-01039-x.
256. Romero E, Fry S, Hooker B. Safety of mRNA vaccines administered during the first twenty-four months of the international COVID-19 vaccination program. IJVTPr (2023) 3:891-910. 10.56098/ijvtpr.v3i1.70.
257. Valdes Angues R and Perea Bustos Y. Navigating uncharted waters: Could certain COVID-19 vaccines promote malignancy? Authorea [Preprint] (2022). <https://www.authorea.com/users/508862/articles/586489-navigating-uncharted-waters-could-certain-covid-19-vaccines-promote-malignancy> [Accessed May 22, 2023].